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## Sex and the Preimplantation Embryo - Implications of Sexual Dimorphism in the Preimplantation Period for Maternal Programming of Embryonic Development

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### Abstract

The developmental program of the embryo displays a plasticity that can result in long-acting effects that extend into post-natal life. In mammals, adult phenotype can be altered by changes in the maternal environment during the preimplantation period. One characteristic of developmental programming during this time is that the change in adult phenotype is often different for female offspring than for male offspring. In this paper, we propose the hypothesis that sexual dimorphism in preimplantation programming is mediated, at least in part, by sex-specific responses of embryos to maternal regulatory molecules whose secretion is dependent on maternal environment. The strongest evidence for this idea comes from the study of colony-stimulating factor 2 (CSF2). Expression of CSF2 from the oviduct and endometrium is modified by environmental factors of the mother, in particular seminal plasma and obesity. Additionally, CSF2 alters several properties of the preimplantation embryo and has been shown to alleviate negative consequences of culture of mouse embryos on postnatal phenotype in a sex-dependent manner. In cattle, exposure of preimplantation bovine embryos to CSF2 causes sex-specific changes in gene expression, interferon- $\tau$  secretion, and DNA methylation later in pregnancy (day 15 of gestation). It is likely that several embryokines can alter postnatal phenotype through actions directed towards the preimplantation embryo. Identification of these molecules and elucidation of the mechanisms by

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which sexually-disparate programming is established will lead to new insights into the control and manipulation of embryonic development.

### Keywords

Developmental programming; colony stimulating factor 2; preimplantation embryo; protein diet; embryo culture

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### Introduction

As used in this paper, developmental programming refers to the concept that alterations in the environment of the developing organism can result in a permanent or long-term change that is expressed phenotypically in adult life. In species as diverse as amphibians (Berg et al. 2009), reptiles (Schwanz et al. 2013), teleosts (Meier et al. 2010; Celeghein et al. 2011), and mammals (Roseboom et al., 2001; Walker et al., 2012; Ganu et al., 2012; Fleming et al., 2015), alterations in embryonic environment can have long-acting effects on development that extend into post-natal life. Mechanisms responsible for this phenomenon include changes in the epigenome (El-Hajj et al. 2014; Desai et al. 2015) as well as in patterns of organogenesis and differentiation (Fleming et al., 2015; Wood-Bradley et al., 2015). Developmental programming can be beneficial when the deviation in adult phenotype prepares the individual for life in the anticipated postnatal environment but can be harmful when the actual environment is different than expected (Roseboom et al. 2001; Armitage et al. 2005; Gluckman et al. 2007; Warner 2014).

In mammals, adult phenotype can be altered by changes in maternal function early in pregnancy during the preimplantation period (Fleming et al., 2015). One characteristic of programming during this period is that the change in adult phenotype is often different for female offspring than for male offspring. Sexual dimorphism may reflect the differential benefit of altered adult phenotype to males and females (Warner, 2014) as well as differences in evolutionary importance of reproductive success for each sex in various environments (Trivers and Willard, 1973).

The purpose of this review is to describe sexual dimorphism in developmental programming during the preimplantation period and to propose one mechanism by which it occurs. In particular, it is hypothesized that the phenomenon is dictated by disparate actions on male and female embryos of maternally-derived embryo regulatory molecules whose secretion can be modified by maternal environment. The evidence for this mechanism comes from study of the regulation of the preimplantation embryo by colony stimulating factor 2 (CSF2). As will be discussed, CSF2 affects female embryos differently than male embryos and its secretion by the reproductive tract is altered by the maternal environment.

### Developmental programming occurs during the preimplantation period

There is abundant experimental data to support the idea that perturbations in the environment of the preimplantation embryo can modify the phenotype of the resultant offspring in postnatal life (Tables 1 and 2). Among the maternal environmental factors that

can alter postnatal phenotype are a low protein diet, as shown in the rat (Kwong et al. 2000) and mouse (Watkins et al. 2008, 2010, 2011), reduced feed intake, observed in sheep (Hernandez et al. 2010), induction of inflammation through injection of lipopolysaccharide (LPS), shown in the mouse (Williams et al. 2011), and mating of female mice to males in which seminal vesicles have been surgically excised (termed SVX males) (Bromfield et al. 2014). In addition, culture of mouse embryos after removal from the maternal reproductive tract can affect postnatal function (Ecker et al. 2004; Sjöblom et al 2005; Watkins et al. 2007; Sampino et al. 2014). Moreover, the type of culture medium used to support development has been reported to cause changes in the developmental program, as demonstrated by addition of serum to culture medium of mouse embryos (Fernández-Gonzalez et al. 2004; Calle et al. 2012; Serrano et al. 2014). Biopsy of the mouse embryo has also been reported to alter postnatal phenotype (Sampino et al. 2014).

The particular postnatal modifications caused by perturbation in the embryo's environment vary somewhat between studies. This variation reflects, in part, differences between studies in the environmental signals used to change induce programming and, in part, differences in the type of observations made on offspring. Some postnatal changes can be considered to reduce offspring fitness while others could enhance fitness under certain environments. There is also a set of postnatal alterations that are observed repeatedly for several distinct alterations in embryonic environment. Thus, for example, increased body weight and, often, fatness were observed for offspring derived from mothers fed low protein diets (Kwong et al. 2000; Watkins et al. 2008; Watkins et al. 2011), challenged with LPS (Williams et al. 2011) or mated to SVX males (Bromfield et al. 2014). Similar adult phenotypes were caused by embryo biopsy, culture outside the reproductive tract (Sjöblom et al 2005; Sampino et al. 2014) and exposure to serum during culture (Fernández-Gonzalez et al. 2004). There are other indications of deviations in metabolic function in the postnatal period, including delayed glucose clearance, caused by mating to SVX males (Bromfield et al. 2014) and presence of serum in culture medium (Calle et al., 2012), increased plasma leptin concentrations (Bromfield et al. 2014), and increased incidence of liver steatosis (for embryos cultured with serum; Fernández-Gonzalez et al. 2004). Altered cardiovascular function was observed for offspring from mothers fed a low protein diet (Kwong et al. 2000; Watkins et al. 2008; Watkins et al. 2010; Watkins et al. 2011), cultured during the preimplantation period (Watkins et al. 2007) and cultured in the presence of serum (Fernández-Gonzalez et al. 2004; Serrano et al. 2014). Postnatal behavior was also modified by changes in the preimplantation environment for low protein feeding (Watkins et al. 2008), reduced feed intake (Hernandez et al. 2010), LPS injection (Williams et al. 2011), embryo biopsy (Sampino et al. 2014), and culture with serum (Fernández-Gonzalez et al. 2004).

Increased body weight and adiposity, disrupted glucose homeostasis and impaired cardiovascular function are features of the thrifty phenotype exhibited by offspring of mothers exposed to poor nutrition at various times during gestation (Roseboom et al. 2001; Hales and Barker, 2001; Fisher et al. 2012; Ganu et al. 2012). Given that similar programming events have been observed following a variety of perturbations in the environment of the preimplantation embryo, there may be a common set of programming

events in the preimplantation period that can be induced by disparate signals. Research with a wider range of species and environmental perturbations will resolve this question.

Postnatal phenotype of offspring derived from embryos fed a low protein diet (Watkins et al. 2008) or derived from females mated to SVX males (Bromfield et al. 2014) remains disrupted when embryos are transferred to control females. Thus, developmental programming during this time is inherent in the embryo and not the result of alteration in maternal function later in pregnancy. Among the changes in the preimplantation embryo that are likely responsible for modifications of postnatal phenotype are alterations in gene expression (Kwong et al. 2006; Gad et al. 2012; Calle et al. 2012), the epigenome (Lucas, 2013; Chen et al. 2013; Urrego et al. 2014; Sun et al. 2015) and patterns of embryonic growth and differentiation (Kwong et al. 2000; Eckert et al. 2012; Bromfield et al. 2014). Such changes may be particularly profound during the preimplantation period because it is at this time that the basic patterns of development are being established, including re-establishment of DNA methylation marks (Rivera and Ross, 2013) and formation of the first differentiated cell lineages from which subsequent organs and tissues are derived.

Modifications in embryonic development in the preimplantation period can affect function of extraembryonic tissues much later in gestation. Thus, for example, culture of bovine embryos with the WNT antagonist dickkopf-1 from day 5-7 of development resulted in embryos that were longer than control embryos at day 34 of gestation (Denicol et al. 2014). Additionally, the trophoctoderm of blastocysts from mice fed a low protein diet in the preimplantation period had greater capacity for fluid phase endocytosis than embryos from control females (Sun et al. 2014). Increased fluid-phase endocytosis was also observed for primitive endoderm-like lineages of embryoid bodies derived from low-protein embryos (Sun et al. 2014) and for the visceral yolk sac endoderm at days 12 and 17 of gestation (Watkins et al. 2008). Trophoblast giant cells at day 8 of gestation in placentae from mothers fed a low protein diet during the preimplantation period exhibited greater growth than trophoblast giant cells of placentae that developed in control placentae (Watkins et al. 2015). By day 17 of gestation, placentae of conceptuses from mothers fed a low protein diet during the preimplantation period were smaller than for controls (Watkins et al. 2015).

### **Female-male differences in programming of the preimplantation embryo**

One of the characteristics of developmental programming is that postnatal phenotype often varies between male and female offspring. This phenomenon is not limited to cases where maternal environment is modified during the entirety of gestation or during mid- or late gestation (see Micke et al. 2010; Palou et al. 2012, Fisher et al. 2014, and Khalyfa et al. 2015 for examples) but, as is summarized in Table 1, also occurs for programming caused by changes in maternal environment during preimplantation stages of embryonic development.

Some caution should be taken in interpreting interactions between embryonic sex and maternal environment because some apparent cases of sexual dimorphism may simply reflect sampling error. Thus, an effect of maternal environment might be significant in one sex but not the other due to the lack of experimental power. For example, male mice derived from mothers fed a low protein diet during the preimplantation period had significantly

elevated amounts of angiotensin converting enzyme in the lung (Watkins et al. 2010) and lower expression of *Ucp1* in retroperitoneal fat (Watkins et al. 2011). While not statistically significant, numerical differences in these endpoints were also observed in females.

Even accounting for sampling error, however, there are many other endpoints where altered phenotype following preimplantation programming was observed in one sex without any indication for an effect in the other sex. Sexual dimorphism in alterations in postnatal phenotype has been observed in the low protein diet model (Kwong et al. 2000; Watkins et al. 2008, 2010, 2011), in response to reduced feed intake (Hernandez et al. 2010), LPS challenge (Williams et al. 2011), mating of female mice to SVX males (Bromfield et al. 2014), embryo biopsy (Sampino et al. 2014), embryo culture (Sjöblom et al. 2005), and presence of serum in embryo culture (Fernández-Gonzalez et al. 2004; Serrano et al. 2014).

The fact that programming of the preimplantation embryo in response to changes in maternal environment differs between females and males is not surprising when one considers that gene expression differs between sexes in the preimplantation period. Alterations in the transcriptome of the embryo have been seen as early as has been examined – at the eight-cell stage in mice (Lowe et al. 2015) and morula stage in cattle (Denicol et al. 2015). Initially, most differentially expressed genes are on the X or Y chromosomes (Denicol et al. 2015; Lowe et al. 2015) but more autosomal genes become differentially expressed by the blastocyst stage as shown in the mouse (Kobayashi et al. 2006) and bovine (Bermejo-Alvarez et al. 2010). The magnitude of difference in gene expression between sexes is of low magnitude for most genes. There was a 2-fold or greater difference in expression between sexes for only 11 of 391 differentially-expressed genes in the mouse (Kobayashi et al 2006) and 55 of 3,756 genes in the cow (Bermejo-Alvarez et al. 2010).

The transcriptome is defined by the epigenetic environment and this too can differ between female and male embryos. Sexual dimorphism has been shown for specific methylation sites (Bermejo-Alvarez et al. 2008; Gebert et al. 2009) as well as globally. The pattern of histone methylation in embryonic stem cells derived from eight-cell mouse embryos differed between females and males in ways correlated with differences in gene expression (Lowe et al. 2015). Using antibody to 5-methylcytosine to study bovine embryos, Dobbs et al. (2013a) found that differences in global methylation between sexes varied with stage of development. At the 6-8 cell stage, DNA was more methylated for female embryos than male embryos. Methylation increased thereafter but to a greater extent for males. By the blastocyst stage, males exhibited more DNA methylation than females (Fig. 1).

Sex-specific alteration in gene expression or in the structure of the embryonic epigenome caused by changes in maternal environment during the preimplantation period could persist into fetal and postnatal life and result in sexual dimorphism in phenotype. Sex differences in maternal alterations in expression of an imprinted gene have been shown for the low protein diet model in the rat (Kwong et al. 2006). Feeding a low protein diet reduced expression of the imprinted gene *H19* in male blastocysts but not in female blastocysts. Sex differences in the effect of protein feeding on *H19* expression persisted to day 20 of gestation for fetal liver but not for placenta.

## Embryokines as drivers of developmental programming of the preimplantation embryo

Embryokines are defined as molecules produced by the oviduct and endometrium that regulate embryonic growth and development (Hansen et al. 2014a). These include not only protein hormones, growth factors, and cytokines (Hansen et al., 2014b) but also various small molecules such as amino acids, which can regulate embryo development through the mTOR signaling pathway (Martin et al. 2003), prostacyclin I, which increases blastocyst hatching through activation of peroxisome proliferator activated receptor  $\delta$  (Kang et al. 2011) and endogenous cannabinoids, which act through various receptors to regulate development and implantation (Taylor et al. 2010).

As a class, embryokines are likely to be key mediators of developmental programming responses to changes in maternal environment. In addition, alterations in developmental processes caused by embryo culture (Watkins et al. 2007; Farin et al. 2010; Chen et al., 2013; Sampino et al. 2014) could reflect, at least in part, the absence of critical maternal regulatory molecules or, for the case of addition of serum to culture medium (Fernández-Gonzalez et al. 2004; Calle et al. 2012; Serrano et al. 2014), alteration of embryonic function by bioactive molecules not normally involved in control of development. In addition to embryokines, developmental programming in utero or in culture could involve changes in cellular energy metabolism in the developing embryo. It has been shown that alterations in redox potential and mitochondrial activity of the mouse zygote caused by starvation, inclusion of pyruvate or lactate in the culture medium or raising external pH can result in changes in body weight in the postnatal period (Banrezes et al. 2011).

There are two requirements for an embryokine to be considered as participating in developmental programming responses to changes in maternal environment. Firstly, accumulation in the reproductive tract must be modified as a result of changes in maternal function. Secondly, actions of the embryokine on the embryo must cause a deviation in developmental dynamics that alters function of the offspring in postnatal life.

Much of the literature on developmental programming during the preimplantation period is based on the low protein diet model developed by Tom Fleming and colleagues at University of Southampton. This group has proposed that the maternal signal to transmit information about protein nutrition to the embryo is the reduction in uterine luminal concentrations of the branched amino acids leucine, isoleucine and valine, which in turn leads to reduced mTORC1 signaling (Fleming et al. 2015). They have shown that female mice fed a low protein diet for the first 3.5 d of gestation experienced increased circulating concentrations of glucose and reduced circulating concentrations of insulin and free amino acids (Eckert et al. 2012). In addition, there was a reduction in concentrations of several amino acids in uterine fluid, most notably the branched amino acids isoleucine, leucine and valine, and in the blastocyst (asparagine and, at  $P < 0.10$ , hypotaurine, lysine and taurine). Consistent with the idea that changes in amino acid availability alters embryonic function through reduced activation of mTORC1 was the finding that blastocysts of mothers fed a low protein diet had reduced mTORC1 activity, as assessed by phosphorylation of the downstream target S6 ribosomal protein (Eckert et al. 2012). What remains to be seen is whether altered activation of mTORC1 signaling can result in developmental changes that



result in the postnatal phenotypes exhibited by offspring of mothers fed low protein diets during early pregnancy.

Colony stimulating factor 2, also called granulocyte-macrophage colony stimulating factor, is another embryokine that has been implicated in developmental programming. This polypeptide is produced by both the oviduct and endometrium (Giacomini et al. 1995; de Moraes et al. 1999, O'Leary et al. 2004; Nahar et al. 2013) particularly by epithelial cells (Giacomini et al. 1995; de Moraes et al. 1999). Moreover, secretion of CSF2 depends on environmental factors of the mother. In particular, seminal plasma triggers expression of *CSF2* in endometrium of the mouse and pig (Tremellen et al. 1998; O'Leary et al. 2004; Bromfield et al. 2014) as does mating in the ewe (Scott et al. 2007). There is also a report in the cow that metabolic status can affect *CSF2* expression in the oviduct, with obesity causing reduced expression (Nahar et al. 2013).

Importantly, CSF2 can reverse the deleterious effects of embryo culture in the mouse. Sjöblom et al. (2005) compared postnatal characteristics of mice born following natural mating with those derived from embryos cultured from the two-cell to blastocyst stage in medium with or without CSF2. Embryos were transferred into recipients and postnatal characteristics of the offspring determined. As shown in Table 3, addition of CSF2 to culture medium either partially or completely prevented the effect of culture on postnatal growth in females and males, relative brain mass in males and placental weight of female progeny when they themselves became pregnant. Addition of CSF2 to medium did not, however, prevent the effect of culture on fatness of males in the postnatal period.

### Regulation of the preimplantation embryo by CSF2

The experiment described above provides strong evidence that CSF2 can modify postnatal phenotype by altering development of the preimplantation embryo. Indeed, CSF2 exerts several actions on the preimplantation embryo that could result in a long-term change in developmental dynamics. In the bovine, these include altered expression of genes involved in pluripotency, differentiation and cell survival (Loureiro et al. 2011), increased ability of isolated inner cell mass (ICM) to remain in a pluripotent state (Dobbs et al. 2013b), and resistance to pro-apoptotic signals (Loureiro et al. 2011). In the mouse, blastocysts cultured with CSF2 in the medium experienced reduced expression of specific genes involved in stress responses and apoptosis (Chin et al., 2009) and fewer apoptotic cells after freeze/thawing (Desai et al. 2007). In the pig, CSF2 affected gene expression in the blastocyst (Kwak et al. 2012a) and increased the ratio of trophectoderm to inner cell mass (Kwak et al., 2012b). Embryos cultured with CSF2 have increased competence to develop to term after transfer to recipient females in the mouse (Sjöblom et al. 2005), cow (Loureiro et al. 2009; Denicol et al. 2014b; embryos produced with X-sorted sperm), and human (Ziebe et al. 2013).

Colony stimulating factor 2 has also been reported to increase the competence of cultured embryos to develop to the blastocyst stage in the bovine (de Moraes and Hansen 1997; Loureiro et al. 2009; Dobbs et al., 2013b), human (Sjöblom et al. 1999), pig (Cui et al. 2004; Lee et al. 2013), and mouse (Sjöblom et al. 2005). This action of CSF2 has not always been observed, however, as reported for the sheep (Rooke et al. 2005), mouse (Desai et al. 2007;



Elaimi et al. 2012; Kim et al. 2014) and bovine (Dobbs et al. 2013b). In the cow, CSF2 increased the percentage of embryos becoming blastocysts when development in controls was low but reduced percent blastocysts when development in controls was high (Dobbs et al. 2013b).

### **Sexual dimorphism in responses of the preimplantation bovine embryo to CSF2**

Given that many cases of developmental programming are characterized by sexual dimorphism, including programming effects of CSF2 on the mouse embryo (Sjöblom et al. 2005), it is to be expected that some embryokines would regulate preimplantation development of female embryos differently than development of male embryos. Recent experiments in cattle suggest this is true for CSF2 (Siqueira and Hansen 2015). Furthermore, the consequence is that embryonic function later in development is altered by CSF2 in a sex-dependent manner (Dobbs et al. 2014).

As mentioned earlier, addition of CSF2 to culture medium of bovine embryos at day 5 after insemination, when the embryo is at the morula stage of development, can increase the percent of embryos that become blastocysts at day 7 after insemination (De Moraes and Hansen, 1997; Loureiro et al. 2009). The effect of CSF2 is most pronounced when development of untreated control embryos is low (Dobbs et al. 2013b). Recently, it has been shown that the pro-developmental effect of CSF2 also depends on embryo sex. Using X- and Y-sorted spermatozoa to produce embryos that were about 90% female and male, respectively, Siqueira and Hansen (2015) observed that CSF2 increased the percent of fertilized oocytes becoming blastocysts for females ( $14.7 \pm 2.1$  vs  $21.5 \pm 2.1\%$ ) but not for males ( $16.2 \pm 2.0$  vs  $16.3 \pm 2.0\%$ ).

Exposure of the bovine embryo to CSF2 from day 5 to 7 after insemination also results in a sex-dependent change in development later in pregnancy after embryos are transferred to recipient females (Dobbs et al. 2014). The stage examined was Day 15 of gestation when the bovine embryo is undergoing rapid elongation of the trophoblast and secretion of the antiluteolytic molecule interferon- $\tau$  (IFNT) to prevent prostaglandin-induced destruction of the corpus luteum (Spencer et al. 2007). Treatment of in vitro produced embryos with CSF2 from day 5 to 7 of development affected conceptus length and concentrations of IFNT in the uterine lumen at Day 15 of pregnancy differently for females than for males. Treatment with CSF2 decreased embryo length and intrauterine accumulation of IFNT in females but increased length and IFNT in males (Fig. 2). Analysis of the transcriptome of extra-embryonic membranes also indicated that sex affected the response to CSF2. Treatment caused differential regulation of 94 genes in males and 56 genes in females. Only 7 differentially-regulated genes were regulated in both sexes and, in these cases, regulation by CSF2 was always in the opposite direction. Colony stimulating factor 2 also altered the methylome as assessed using a methylation array containing probes for 418,805 CpG located across the genome. Treatment with CSF2 caused hypermethylation for 6,227 probes in females and 9,842 probes in males. Hypomethylation was detected for 3,292 probes in females and 9,322 probes in males. Only 1186 probes were regulated by CSF2 in both sexes, and regulation was in the opposite direction for 772 of these probes.

### Concluding remarks – a model for sexual dimorphism in developmental programming

Developmental plasticity in response to maternal environment is an evolutionary strategy which allows individuals to adapt to the environment during their life, thereby preserving the genotype in the face of transient environmental changes (Gluckman et al. 2007; Warner 2014). Sexual dimorphism in developmental programming may have evolved because the benefit of altered adult phenotype can vary between sexes (Warner 2014) or because the relative importance of females and males for evolutionary success depends on environment (Trivers and Willard 1973). Based on the research reviewed in this paper, it is proposed that sexual dimorphism in developmental programming of mammals during preimplantation development can be dictated by disparate actions of maternally-derived regulatory molecules (i.e., embryokines) on male and female embryos. As a result, alteration in secretion of embryokines in response to environmental change can result in sex-specific deviation in the developmental program.

This hypothesis is shown schematically in Fig. 3 for the example of how changes in maternal environment might differentially affect growth and function of the bovine embryo as a result of altered secretion of CSF2 from the reproductive tract. In this model, an environmental cue acting on the mother (for example, nutrition) leads to a signal that transmits information about the maternal environment to the embryo. In the illustrated example, the signal is a change in secretion of CSF2 from the maternal reproductive tract. The resultant alteration in the trajectory of development is different for female embryos than male embryos so that subsequent characteristics of the embryo and neonate are modified differently for each sex.

There are many unanswered questions about mechanisms for establishment of sexual dimorphism in programming of postnatal phenotype. While CSF2 can prevent alterations of postnatal phenotype in the mouse caused by embryo culture (Sjöblom et al. 2005), it has not yet been demonstrated that changes in secretion of CSF2 mediate alterations in the developmental program caused by modifications in the maternal environment. There are indeed likely to be an array of embryokines that can program development. Their identification will be critical to understanding mechanisms for developmental programming.

It will also be important to understand how a specific maternal signal can affect the developmental program of a female embryo differently from that of a male embryo. The particular transcriptional and post-transcriptional responses of an embryo to maternal cues are defined by characteristics of the embryo's transcriptome and epigenome. Female and male preimplantation embryos differ in both gene expression (Kobayashi et al. 2006; Bermejo-Alvarez et al. 2010; Denicol et al. 2014; Lowe et al. 2015) and the epigenetic landscape (Bermejo-Alvarez et al. 2008; Gebert et al. 2009; Dobbs et al. 2013a). Identification of the specific genes controlling sex differences in response to maternal cues will open up a new understanding of the control of embryonic development in the preimplantation period.

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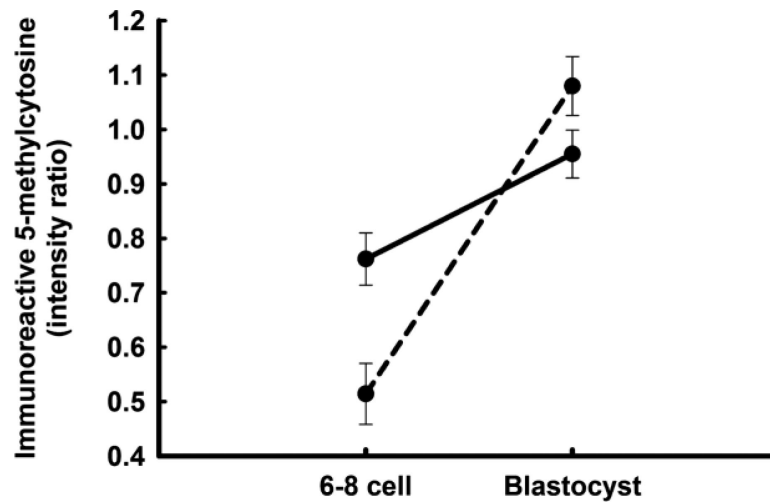
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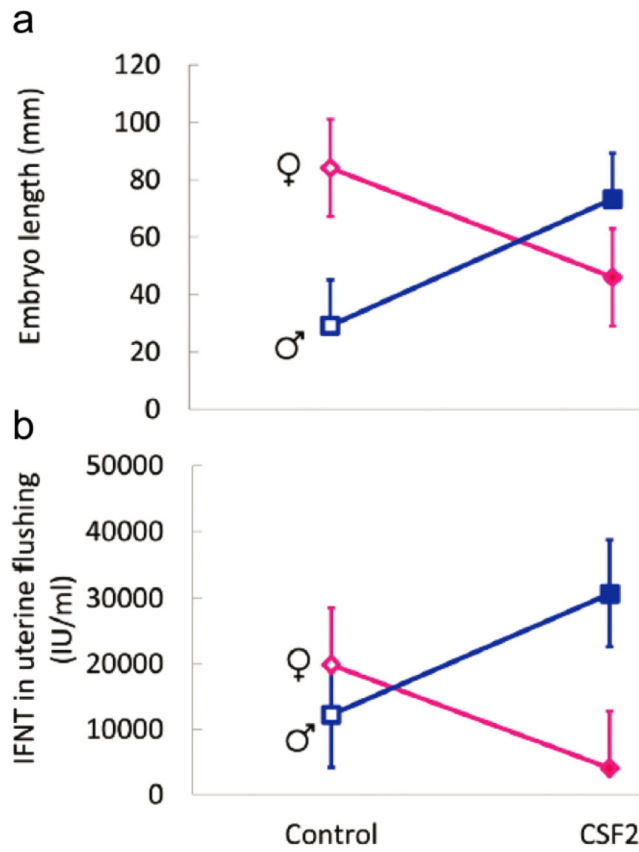
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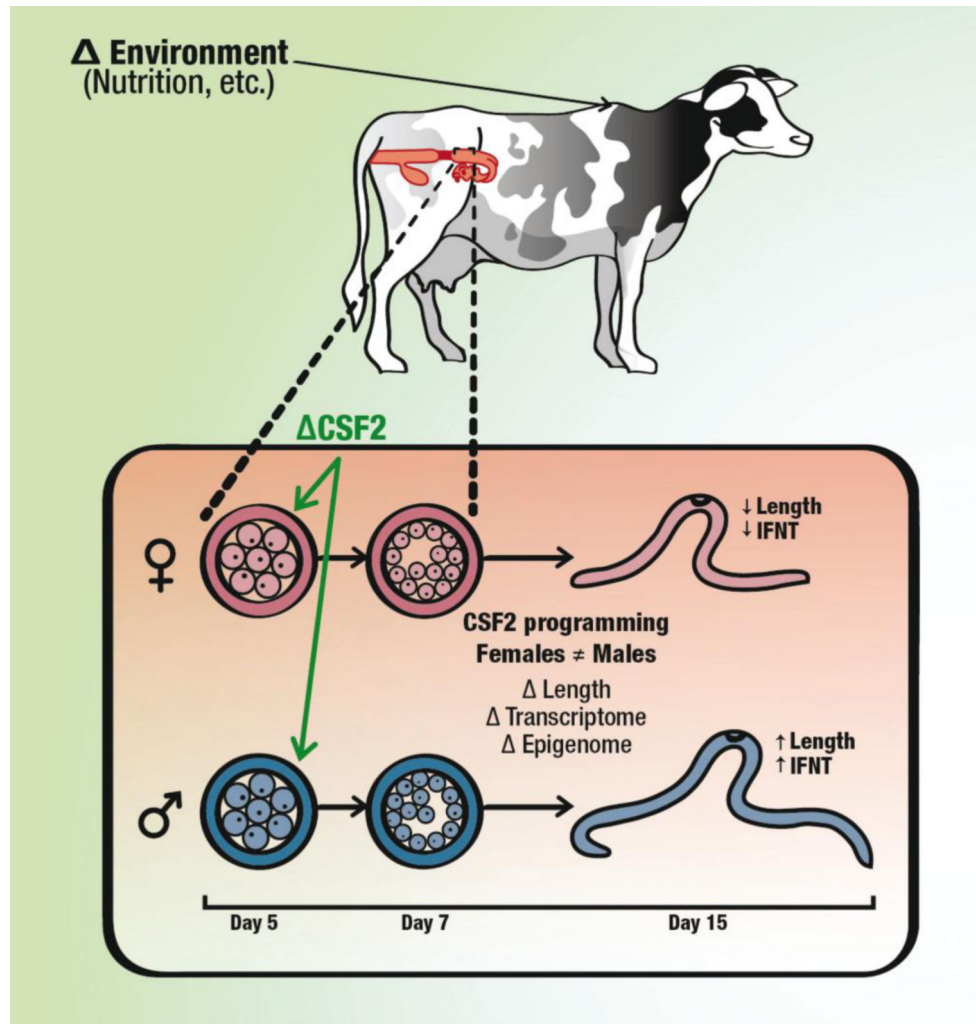




**Fig. 1.** Differences in immunoreactive 5-methylcytosine between female (solid line) and male embryos (dashed line) at the 6–8 cell and blastocyst stages of development in cattle. Data represent fluorescent intensity for anti-5-methylcytosine relative to that for nuclear propidium iodide. The figure is from Dobbs et al. (2013a) and is reproduced from *PLOS One*.



**Fig. 2.** Interactions between CSF2 treatment from day 5-7 of development and sex on embryo length and concentration of IFNT in uterine flushing at Day 15 of pregnancy. The figure is reproduced from *Biology of Reproduction* (Dobbs et al. 2014).



**Fig. 3.** Model illustrating a mechanism for achievement of sex-specific developmental programming in response to a modification in maternal environment. In this model, a change in maternal environment alters secretion of uterine embryokines such as CSF2. The resultant deviation in the trajectory of development is different for female embryos than male embryos so that subsequent characteristics of the embryo and neonate are modified differently for each sex. In the particular example of CSF2, exposure to CSF2 from days 5-7 of after fertilization causes different and often opposite actions on development reflected in characteristics of the embryo at day 15 of development.

**Table 1**

Sexual dimorphism in developmental programming caused by alterations in the maternal environment during the preimplantation period.

Species	Treatment	Altered postnatal phenotype			Reference
		Both sexes	Females only	Males only	
Rat	Low protein diet, 0-4.25 d of gestation	<ul style="list-style-type: none"> <li>•Increased body weight through 7 wk; no difference thereafter</li> </ul>		<ul style="list-style-type: none"> <li>•Increased systolic blood pressure</li> <li>•Increased kidney:body weight and decreased liver:body weight ratio</li> </ul>	Kwong et al. 2000
Mouse	Low protein diet, 0-3.5 d of gestation	<ul style="list-style-type: none"> <li>•Increased systolic blood pressure</li> </ul>	<ul style="list-style-type: none"> <li>•Increased body weight through 26 wk of age</li> <li>•Decreased heart:body weight ratio</li> <li>•Altered behavioral responses</li> </ul>		Watkins et al. 2008
Mouse	Low protein diet, 0-3.5 d of gestation	<ul style="list-style-type: none"> <li>•Elevated lung angiotensin converting enzyme, 28 wk of age (significant for males only)</li> </ul>		<ul style="list-style-type: none"> <li>•Attenuated vasodilation response in mesenteric arteries, 22 wk of age</li> </ul>	Watkins et al. 2010
Mouse	Low protein diet, 0-3.5 d of gestation	<ul style="list-style-type: none"> <li>•Increased systolic blood pressure (significant for females only)</li> <li>•Lower expression of <i>Ucp1</i> in retroperitoneal fat (significant for males only)</li> </ul>	<ul style="list-style-type: none"> <li>•Increased body weight from 11-28 wk of age; no effect afterwards through 52 wk</li> <li>•Higher expression of <i>Insr</i> and <i>Igf1r</i> in retroperitoneal fat</li> </ul>		Watkins et al. 2011
Sheep	Reduced feed intake, d -2 to 30 after mating			<ul style="list-style-type: none"> <li>•Increased cortisol response to isolation, 4 mo of age</li> <li>•Increased escape attempts, behavioral test, 4 mo of age</li> </ul>	Hernandez et al. 2010
Mouse	Lipopolysaccharide (LPS) challenge at zygote stage	<ul style="list-style-type: none"> <li>•Reduced cytokine response to LPS immune challenge</li> </ul>		<ul style="list-style-type: none"> <li>•Altered behavior</li> <li>•Increased body mass index</li> <li>•Increased fat pad: body weight ratio</li> </ul>	Williams et al. 2011
Mouse	Mating of females with males without seminal vesicles	<ul style="list-style-type: none"> <li>•Reduction in prepubertal growth; increase in postpubertal growth</li> </ul>		<ul style="list-style-type: none"> <li>•Increased adiposity</li> <li>•Increased plasma leptin concentration</li> <li>•Delayed glucose clearance</li> </ul>	Bromfield et al. 2014

**Table 2**

Sexual dimorphism in developmental programming caused by culture or manipulation of preimplantation embryos.

Species	Treatment	Altered postnatal phenotype			Reference
		Both sexes	Females only	Males only	
Mouse	Culture, 2-cell to blastocyst	<ul style="list-style-type: none"> <li>Increased body weight (effect persisted for longer after birth for males)</li> </ul>	<ul style="list-style-type: none"> <li>Increased placental weight of progeny when they were pregnant</li> </ul>	<ul style="list-style-type: none"> <li>Increased fatness</li> <li>Decreased brain mass as percent of total body mass</li> </ul>	Sjöblom et al. 2005
Mouse	Culture, 2-cell to blastocyst	<ul style="list-style-type: none"> <li>Increased systolic blood pressure</li> </ul>			Watkins et al. 2007
Mouse	Culture, 8-cell to blastocyst stage	<ul style="list-style-type: none"> <li>Increased body weight at week 2 of age and later through wk 16</li> </ul>			Sampino et al. 2014
Mouse	Biopsy of 8-cell embryos			<ul style="list-style-type: none"> <li>Increased body weight at weeks 6, 10, and 12-16 of age</li> <li>Altered behavioral responses</li> </ul>	Sampino et al. 2014
Mouse	Culture from the 1-cell stage with serum vs culture without serum	<ul style="list-style-type: none"> <li>Altered behavior, elevated plus maze</li> </ul>	<ul style="list-style-type: none"> <li>Increased body weight, wk 31-70 of age</li> <li>Increased incidence of liver steatosis</li> </ul>	<ul style="list-style-type: none"> <li>Altered behavior, open field test</li> <li>Increased incidence of perivascular kidney inflammatory infiltration</li> </ul>	Fernández-Gonzalez et al. 2004
Mouse	Culture from the 1-cell stage with serum vs culture without serum		Not tested	<ul style="list-style-type: none"> <li>Increased apoptosis in testis</li> <li>Reduced sperm motility</li> <li>Reduced pregnancy rates in females that males were exposed to</li> <li>Delayed glucose clearance</li> <li>Insulin resistance</li> </ul>	Calle et al. 2012
Mouse	Culture from the 1-cell stage with serum vs culture without serum	<ul style="list-style-type: none"> <li>Increased body weight</li> <li>Increased mortality to 20 mo of age</li> </ul>	<ul style="list-style-type: none"> <li>Increased systolic blood pressure</li> <li>Increased fat content</li> <li>Increased incidence of hepatomegaly and liver steatosis</li> <li>Decreased expression of <i>Fas</i>, <i>Lep</i>, <i>Ppara</i> and <i>Pparg</i> in white adipose</li> </ul>	<ul style="list-style-type: none"> <li>Increased plasma glucose concentration</li> <li>Increased expression of <i>Acox</i>, <i>Ghr</i>, <i>Ppara</i>, and <i>Ucp1</i> in white adipose</li> </ul>	Serrano et al. 2014

**Table 3**

Addition of CSF2 to culture medium reduces some of the effects of embryo culture on postnatal characteristics of the resultant offsprings.<sup>a</sup>

	Female			Male		
	In vivo	In vitro	In vitro + CSF2	In vivo	In vitro	In vitro + CSF2
Body weight, 5 wk (g) <sup>b</sup>	21.5 ± 0.3	22.4 ± 0.3	21.5 ± 0.2*	-	-	-
Body weight, 9 wk (g) <sup>b</sup>	-	-	-	37.6 ± 0.5	39.0 ± 0.5	37.6 ± 0.4*
Body weight, 12 wk (g)	29.8 ± 0.9	30.6 ± 0.7	30.2 ± 0.8	38.3 ± 0.8	42.7 ± 0.8	41.2 ± 0.8
Mass of abdominal fat, percent of lean body mass	4.0 ± 0.3	4.6 ± 0.3	4.2 ± 0.2	4.0 ± 0.3	5.0 ± 0.2	4.9 ± 0.2
Brain mass, percent of lean body mass	1.83 ± 0.05	1.75 ± 0.05	1.77 ± 0.04	1.37 ± 0.03	1.25 ± 0.03	1.31 ± 0.03*
Placental weight of progeny when they become pregnant (mg)	93 ± 2	111 ± 3	97 ± 2*	-	-	-

<sup>a</sup>From Sjöblom et al. (2005)

<sup>b</sup> maximum differences in weight between groups occurred at 5 wk for females and 9 wk for males

\* Significantly different from in vitro (P<0.05).