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Local niche differences predict genotype associations in sister taxa of desert tortoise

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Abstract

Aims: To investigate spatial congruence between ecological niches and genotype in two allopatric species of desert tortoise that are species of conservation concern. **Location:** Mojave and Sonoran Desert ecoregions; California, Nevada, Arizona, Utah, USA.

Methods: We compare ecological niches of *Gopherus agassizii* and *Gopherus morafkai* using species distribution modelling (SDM) and then calibrate a pooled-taxa distribution model to explore local differences in species–environment relationships based on the spatial residuals of the pooled‐taxa model. We use multiscale geographically weighted regression (MGWR) applied to those residuals to estimate local species–en‐ vironment relationships that can vary across the landscape. We identify multivariate clusters in these local species–environment relationships and compare them against models of (a) a geographically based taxonomic designation for two sister species and (b) an environmental ecoregion designation, with respect to their ability to predict a genotype association index for these two species.

Results: We find non‐identical niches for these species, with differences that span physiographic and vegetation niche dimensions. We find evidence for two distinct clusters of local species–environment relationships that when mapped, predict an index of genotype association for the two sister taxa better than did either the geo‐ graphically based taxonomic designation or an environmental ecoregion designation. **Main conclusions:** Exploring local species–environment relationships by coupling SDM and MGWR can benefit studies of biogeography and conservation. We find that niche separation in habitat selection conforms to genotypic differences between sister taxa of tortoise in a recent secondary contact zone. This result may inform de‐ cision making by agencies with regulatory or land management authority for the two sister taxa addressed here.

KEYWORDS

conservation genetics, desert tortoise, ecological niche, multiscale geographically weighted regression, species distribution modelling

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1 | **INTRODUCTION**

The relationships between the distributions of species and their ecological properties have long been central to biogeographic in‐ quiry (Grinnell, 1917; MacArthur, 1972). More recently, quantitative methods to define ecological niches have become essential toolsets for investigating how species are distributed in environmental and geographic space (Araújo & Guisan, 2006; Guisan & Zimmermann, 2000). Species distribution modelling (SDM) is one such toolset that relates locations of species observations to explanatory variables hypothesized to influence or define a species' Hutchinsonian niche (Franklin, 2010). SDM quantifies the relationships between environ‐ mental conditions at locations where a species has been observed to those locations where it has not in order to predict how likely it is to occur at unobserved locations. These relationships, hereafter "spe‐ cies–environment relationships," are represented by model coeffi‐ cients and can range from simple linear parameters in the most basic form of generalized linear modelling (GLM), to complex combinations of basis functions (e.g., polynomials, splines) in generalized additive modelling (GAM) or machine learning frameworks (Franklin, 2010).

In conservation biogeography, metrics such as phylogenetic di‐ versity (Crozier, 1997; Helmus, Bland, Williams, & Ives, 2007; Scoble & Lowe, 2010; Vandergast et al., 2013; Wood et al., 2013) are being recognized as important for conservation planning and managing biological resources (Myers, Mittermeier, Mittermeier, Fonseca, & Kent, 2000; Naeem, Duffy, & Zavaleta, 2012; Rodrigues & Gaston, 2002; Winter, Devictor, & Schweiger, 2013). Spatially structured variation in phylogenetic diversity may foster ecosystem resilience to global change through evolutionary potential (Devictor et al., 2010; Flynn, Mirotchnick, Jain, Palmer, & Naeem, 2011; Legendre, Borcard, & Peres‐Neto, 2005; Tews et al., 2004), and therefore, the ability to map species' distributions and their relatedness has shown to be a valuable tool for conservation (Moritz, 2002; Rodrigues & Gaston, 2002; Scoble & Lowe, 2010; Winter et al., 2013). SDM is widely used for mapping distributions, but also provides a frame‐ work for quantifying differences between species' niches (Peterson, Soberón, & Sanchez‐Cordero, 1999; Warren, Glor, & Turelli, 2008).

For example, under conditions of allopatric speciation, SDM can be used to develop niche models for sister species in order to quantify niche differences using ecological overlap metrics (e.g., Broennimann et al., 2011; Godsoe, 2013; Rödder & Engler, 2011) and statistical tests (e.g., Nunes & Pearson, 2017; Warren et al., 2008; Warren, Glor, & Turelli, 2010). Substantial similarity between niches may suggest niche conservation between the two allopatric species (Warren et al., 2008; Wiens & Graham, 2005), such that the species–environment relationships are maintained through time and across taxa, even in the presence of environmental change or speciation (Wiens & Graham, 2005). However, the assumptions normally imposed in SDM may affect its utility for niche comparisons, including (a) imposing a single scale for all species–environment re‐ lationships and (b) fixing the spatial scale of those species–environ‐ ment relationships across space. The former is well understood, with a consensus that no single scale is most appropriate for studying

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ecological phenomena (Levin, 1992; Wiens, 1989) and that under‐ standing landscape structure and ecosystem processes may re‐ quire multi‐scale approaches (Rahbek & Graves, 2001; Seo, Thorne, Hannah, & Thuiller, 2009; Willis & Whittaker, 2002; Wu, 2004). This is evident in SDM where climate may dominate distributions at global to regional scales, yet topography and surface characteristics may influence species at regional to local scales (Beever, Swihart, & Bestelmeyer, 2006; Mackey & Lindenmayer, 2001).

The second constraint assumes that species–environment rela‐ tionships (and the model coefficients that define them) do not vary across the geographic range of each species. Mixture zones, espe‐ cially those with hybridization, may represent areas where a species' niche differs from the rest of its range due to local adaptation in different habitats (Lenormand, 2012). In these areas, species-environment relationships defined from the entirety of a species' range may not adequately characterize local differences, such that a single set of model coefficients may not represent the variation in species–environment relationships across space (Foody, 2004; Miller, 2012; Osborne, Foody, & Suárez‐Seoane, 2007) because model co‐ efficients may covary with location (Atkinson, 2001; Fotheringham, 1997). SDM practitioners have developed tools for incorporating po‐ tential spatial variation in species–environment relationships by cali‐ brating separate models on subregions of a species' distribution (e.g., Peterson & Holt, 2003) or through the use of additive or multipli‐ cative indicator variables. However, these solutions require a priori knowledge of the configuration of any hypothesized subregions and therefore limit exploration of spatial patterns in model coefficients.

Identifying spatially varying relationships across ecological boundaries (such as between two adjacent species) can highlight differences in data quality spanning those transition zones (e.g., differences in sampling regimes; Cheng & Fotheringham, 2013) and can provide infor‐ mation about the nature of the boundary, such as potential secondary contact between vicariant populations (Endler, 1977; Gay, Crochet, Bell, & Lenormand, 2008; Jiggins & Mallet, 2000). Through a conser‐ vation biogeography lens, boundaries and the gradients spanning them are key concepts that give context to phylogenetic diversity and the conditions that influence speciation (Hoffmann & Blows, 1994).

Tools such as geographically weighted regression (GWR; Fotheringham, Brunsdon, & Charlton, 2002) have been used to ex‐ plore locally varying processes and their spatial scale across bound‐ aries (Cheng & Fotheringham, 2013) and have also been used to investigate locally varying patterns in species richness (Bickford & Laffan, 2006; Foody, 2004; Holloway & Miller, 2015) and spe‐ cies–environment relationships (Kupfer & Farris, 2006; Miller, 2012; Miller, Franklin, & Aspinall, 2007; Osborne et al., 2007). In GWR, the spatial scale of species–environment relationships is represented with bandwidth parameters that determine the degree to which nearby observations are given higher weights than more distant ones (Fotheringham et al., 2002). Large bandwidths approximate classical regression with a single set of coefficients to represent species–environment relationships, while small bandwidths result in highly local models with species–environment relationships that can vary across the landscape. However, previous implementations of **1196 WILEY Diversity** and **Distributions**

FIGURE 1 Study area used to create pooled‐taxa species distribution models (light grey) and region of habitat for the two species of desert tortoise, *Gopherus agassizii* (Agassiz's tortoise; light orange) and *Gopherus morafkai* (Morafkai's tortoise; purple). The focal study area (thin black line) encompassing the contact zone was used to reduce computation time for local models and genotype assessment. The Colorado River (blue) separates California and Arizona and creates the division between the two species of desert tortoise. A Distinct Population Segment defined as the Mojave population includes individuals located west of the Colorado River. USA Contiguous Albers Equal Area Conic projection (SR‐ORG:7301)

GWR have required a single bandwidth for all explanatory variables (Fotheringham et al., 2002), thus precluding a multi‐scale approach. A recent development has enabled estimation of separate spatial scales for each explanatory variable by optimizing multiple band‐ width parameters—multiscale geographically weighted regression (MGWR; Fotheringham, Yang, & Kang, 2017), which allows a multi‐ scale approach to exploring species–environment relationships.

Here we develop a case study of two sister taxa to explore geographic patterns of niche differences between them, with focus on their differing conservation status. These two species, *Gopherus agassizii* (Agassiz's desert tortoise) and *Gopherus morafkai* (Morafka's desert tortoise) diverged approximately 6 Ma due to geographic iso‐ lation by the Bouse embayment, a putative marine transgression of the ancestral Gulf of California along the lower Colorado River, which has resulted in allopatric speciation (Murphy et al., 2011). These two cryptic species were only recently distinguished phylogenetically and taxonomically due to differences in genetics, reproductive ecol‐ ogy and seasonal activity (McLuckie, Lamb, Schwalbe, & McCord, 1999; Murphy et al., 2011), but are not readily distinguished morpho‐ logically. Prior to the taxonomic split, a distinct population segment (DPS) defined as the Mojave population (Figure 1; tortoises west and north of the Colorado River) was listed as threatened with extinction and given legal protection under the U.S. Endangered Species Act (ESA; Department of the Interior, 1990) and has received extensive monitoring yielding a wealth of georeferenced observations. The re‐ maining "Sonoran Population," later elevated as the distinct species, *G. morafkai*, does not have the same legal protection or monitoring effort (Murphy et al., 2011; Service, 2015).

While the Colorado River defines the geographic division be‐ tween the species, recent genetic work has identified a secondary

contact zone where *G. agassizii* (the western species) occurs in a small population east of the Colorado River (Edwards et al., 2015; McLuckie et al., 1999). This secondary contact zone likely emerged only 2.5 ka as a result of avulsion in the Colorado River, but now *G. agassizii* in this zone are isolated from individuals occurring west of the Colorado River. This small population faces threats from in‐ creasing development in the region and is not legally protected. This situation is further complicated by evidence of natural hybrid‐ ization between *G. agassizii* and *G. morafkai* occurring in this second‐ ary contact zone, and by the lack of a clear definition of habitat for this population of *G. agassizii* east of the river. Recent work has also suggested that this population occupies habitat with intermediate characteristics to that of the pure lineages (Edwards et al., 2015). Habitat for *G. agassizii* and *G. morafkai* outside the contact zone is better defined, with known habitat ranging from valley bottoms and alkaline areas surrounding playas in the Mojave, to bajadas and alluvial fans, arroyos, rocky slopes and ridges in the upland regions in the Sonoran desert (Nussear & Tuberville, 2014). Differences in habitat characteristics between the two species span physiography (Nussear & Tuberville, 2014), geology (Burge, 1978), vegeta‐ tion (Bury, Esque, DeFalco, & Medica, 1994) and climate (Nussear, Esque, Inman, Gass, & Thomas, 2009; Tracy et al., 2004). However, no formal tests of niche similarity have been conducted to date, further complicating delineations of these two iconic species.

We use SDM and MGWR in a coupled modelling approach to identify differences in the ecological niches of *G. agassizii* and *G. morafkai*, and explore spatially varying species–environment relationships in the recent secondary contact zone. We (a) formally test for differ‐ ences in their ecological niches, (b) identify boundaries represented by differences in their niches and (c) determine which of three spatial **Diversity** and **Distributions** \rightarrow **WII** \rightarrow **1197**

delineations better describes landscape patterns of genotypic varia‐ tion. These delineations include (a) the Colorado River (the current geo‐ graphic boundary defining each species), (b) the Mojave and Sonoran Basin and Range ecotone, and (c) geographic patterns in local niche differences identified in this study. The results of this study will inform conservation planning across the transition zone of these two species.

2 | **METHODS**

2.1 | **Study area**

Our study area included the known range of *G. agassizii* and G. *morafkai* across 68,323 km² in the Southwestern United States, encompassing parts of California, Arizona, Nevada and Utah (Figure 1). This region is characterized as the Mojave Basin and Range Level III Ecoregion and Sonoran Basin and Range Level III Ecoregion (Wiken, Nava, & Griffith, 2011), hereafter the Mojave Desert and Sonoran Desert, respectively. The subregion encompassing the genetic sampling locations used by Edwards et al. (2015), hereafter referred to as the focal study area (Figure 1), offered an opportunity to explore spatial patterns in species-environment relationships across the ecotone between the Mojave and Sonoran deserts and in the second‐ ary contact zone between *G. agassizii* and *G. morafkai*.

2.2 | **Modelling overview**

We developed a two-step modelling approach drawing on the strengths of both SDM and MGWR (Figure 2) to explore spatial

FIGURE 2 Modelling Overview. Two-step modelling approach using species distribution modelling (SDM) and multiscale geographically weighted regression (MGWR) to explore spatial patterns in species–environment relationships of *Gopherus agassizii* and *Gopherus morafkai*. In step 1, we use SDM to develop range‐wide ecological niche models for each species separately and test hypotheses of niche equivalency. We also pool both species to develop a single model of their combined ecological niche and use the mapped residuals from this pooled model as a measure of local deviation. In step 2, we use MGWR to explore spatial patterns in the relationships between these residuals and explanatory variables that may enumerate differences between the two species and their hybrids within the focal study area

patterns in species–environment relationships of *G. agassizii* and *G. morafkai* across this secondary contact zone. In the first step, we use SDM to develop range‐wide ecological niche models for each species separately and test hypotheses that the niches of these two species are more different than would be expected by chance. We then pool location data for both species and develop a single model of their combined ecological niche and use the mapped residuals from this pooled model as a measure of local deviation. We assume that if the two species exhibit different ecological niches, residu‐ als from a pooled model will represent how poorly their combined niche predicts the probability of presence at a given location. In the second step, we use MGWR to explore spatial patterns in the relationships between these residuals and hypothesized explanatory variables that may enumerate differences between the two species and their hybrids within the focal study area.

2.3 | **Species distribution modelling**

We used MaxEnt v. 3.4.0 (Phillips, Dudik, & Schapire, 2018) to create distribution models for each species separately and for a combined pooled model in a presence–background framework. The latest ver‐ sions of Maxent software (e.g., version 3.4.0) produce an estimate of occurrence probability based on the complementary log–log function (Phillips, Anderson, Dudik, Schapire, & Blair, 2017) which we use as the probability of presence. We allow inclusion of all feature classes (linear, quadratic, product and hinge). Observational data spanning 1970–2013 from 23 datasets encompassing the U.S. portions of the two species' known ranges were compiled from Nussear et al. (2009) and augmented with additional observations from the Arizona Game and Fish Department. Occurrences with spatial precision worse than 1 km were discarded, and the remainder were limited to one per each 1 km 2 grid cell, resulting in 8,728 $\,$ observations for the two species available for model calibration. To reduce the effects of spatial sampling bias, we implement a back‐ ground weight correction with the FactorBiasOut algorithm (Dudik, Phillips, & Schapire, 2005) and use a bias grid as an estimate of the sampling bias by creating a kernel density raster of observations for each species. The bandwidth for each kernel was estimated using cross‐validation to minimize mean square error (Baddeley, Rubak, & Turner, 2015) and was linearly rescaled to 1–20 range to give greater background selection probability to areas with higher densities of observations (Elith et al., 2011).

In order to compare niches of *G. agassizii* and *G. morafkai*, a com‐ mon set of explanatory variables is needed; we therefore considered a set of 13 explanatory variables (Table 1) that represented a suite of physiographic, vegetative and climatic characteristics hypothesized to influence the distribution of both species (Edwards et al., 2015; Inman et al., 2014; Nussear et al., 2009). These variables were cho‐ sen from among 18 by removing variables with Pearson's correlation values greater than 0.6 to reduce multicollinearity. Variable selection started with a single model including the 13 remaining explanatory variables and sequentially removing those contributing the least to model fit using a step-wise jackknife test of training gain (Elith et

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al., 2011). We stopped removing variables when a decrease of 0.05 in the area under the receiver operating characteristic curve (AUC; Fielding & Bell, 1997) was observed with 20% withheld test data. The selected set of explanatory variables was used to calibrate mod‐ els for each species separately using a bootstrap framework with 100 iterations and also to create a pooled model by treating the two species as a single taxon and pooling observations. We report the test AUC for each model, as well as relative contributions for each explanatory variable on training gain.

2.4 | **Niche comparisons**

We hypothesized that the niches of the two species would show similarities due to relatedness and niche conservation, but that dif‐ ferences would also be apparent due to geographic isolation over the past 6 million years. We therefore compared their niches using three methods: (a) correlation of explanatory variable contributions, (b) correlation of species–environment relationships, and (c) rand‐ omization tests of niche similarity. In the first, we used Pearson's correlation coefficient to compare the contributions of the explana‐ tory variables across species. Variable contributions were assessed as the relative per cent contribution to model training gain and can be used as a measure of variable importance (Phillips & Dudik, 2008). We asked if the explanatory variables had the same importance to each species, such that high correlation of these contribution scores across species would indicate niche overlap and provide additional evidence that these two species share niche properties, whereas low correlation would suggest that some explanatory variables are more important to one species than the other.

Our second method compared the marginal species–environ‐ ment relationships of each explanatory variable from the calibrated niche models of each species. We again used Pearson's correlation coefficients to compare the predicted probability of presence at 100 intervals spanning the range of background values between the two species. Marginal species–environment relationships ex‐ press the predicted probability of presence across the range of background values of each explanatory variable while keeping all other explanatory variables at their mean value (Phillips & Dudik, 2008). Differences in the shape of species–environment response curves suggest niche differences between the two species. We hy‐ pothesized that differences due to prolonged geographic isolation over the past 6 million years would be limited to physiographic vari‐ ables as a result of the substantial differences in terrain between occupied habitats. For example, *G. agassizii* occurs more often in valley bottoms and gentle slopes with smaller sediment size, while *G. morafkai* occurs more often in rockier slopes and bajadas (Nussear & Tuberville, 2014). We also assumed that differences in their re‐ sponses to temperature would be minimal because these species have evolved in comparable climates.

Our third method to compare the species' niches used ran‐ domization tests of niche equivalency and asymmetrical similarity proposed by Warren et al. (2008). Here we ask: (a) if the niches of the two species are equivalent? and (b) are they more similar than **1200 WILEY Diversity** and Distributions **Distributions**

would be expected if each species selected habitat at random within their ranges? To address the first question, we compared the actual similarity between *G. agassizii* and *G. morafkai* to a null distribution of similarity scores generated from niche models calibrated on ran‐ dom partitions of the pooled observations with sample sizes pro‐ portional to the species' observation datasets. This null distribution represents hypothetical conditions where the two species are com‐ pletely intermingled throughout their combined ranges, such that they share identical niches. The second question was addressed by comparing the similarity score between *G. agassizii* and *G. morafkai* to a distribution of scores obtained by comparing the probability of presence of *G. agassizii* to the probability of presence from niche models calibrated with random locations within the known range of *G. morafkai*. This null distribution (*G. agassizii* → *G. morafkai*) rep‐ resents hypothetical conditions where *G. morafkai* occupies slight permutations of its current niche, such that any differences in the niches between the two species are due to local variation in environ‐ mental conditions, not geographic differences between their ranges. We repeated this test for *G. morafkai* (*G. morafkai* → *G. agassizii*) and implemented these tests in GRASS 7.4 (Neteler, Bowman, Landa, & Metz, 2012) and R 3.5.1 (R Core Team, 2016) with 100 replications. For each of these randomization tests (equivalency and asymmetrical similarity), we define niche similarity with the expected frac‐ tion of shared presences overlap metric (ESP; Godsoe, 2013), which measures the degree to which two probability of presence surfaces agree.

2.5 | **Local niche models and spatial scale**

In order to further investigate differences in species–environment relationships between *G. agassizii* and *G. morafkai*, we calibrated *local* species–environment relationships within our focal study area around the secondary contact zone to estimate local variation that may exist across this region. We hypothesized that if differences in species–environment relationships were evident between the two species, then spatial gradients in those relationships may also be evident in the ecotone between them. We expected that these species–environment relationships would be expressed at different spatial scales due to differences between climatic and topographic constraints on distributions, where climate may affect distributions at regional scales, while topography may influence local scale pat‐ terns (Beever et al., 2006; Mackey & Lindenmayer, 2001). To identify spatially varying environmental responses that we expected to be multi‐scale, we developed a local regression model with MGWR and explored spatial patterns in species–environment relationships as expressed in locally varying model coefficients. We calibrate MGWR on the model residuals of the pooled niche model. The pooled niche model represents the combined niche of both species, such that model residuals can be interpreted as local measures of how poorly the pooled niche model explains local patterns of each species' niche.

Local regression, including MGWR, can be more suscepti‐ ble to multicollinearity than ordinary least squares regression (Fotheringham et al., 2002), and a carefully chosen set of explanatory

variables is less likely to cause erratic behaviour in model coeffi‐ cients. Because our goal was to explore local variation in the model coefficients (i.e., species–environment relationships), we therefore developed composite, uncorrelated predictors from the explanatory variables considered for the pooled niche model. These predictor variables were the principal component axes summing to at least 80% of the component eigenvalues based on four principal compo‐ nent analyses (PCA; Abdi & Williams, 2010) conducted separately for the physiographic, climatic, soils and vegetation variables (see Tables S1.1–S1.4 in Appendix S1) in GRASS GIS with the "i.pca" mod‐ ule using normalization. We explore eigenvalues and their weight loadings to determine which explanatory variables contributed the most to a given principal component axis, in order to relate MGWR model coefficients for PCA axes to underlying explanatory variables. An additional variable representing non‐habitat land cover was derived from the 2011 National Land Change Database (NLCD) Percent Developed Imperviousness layer (Fry et al., 2011) and the National Hydrography Dataset (Simley & Carswell, 2010), and repre‐ sented the per cent of each grid cell covered by impervious surfaces (such as paved roads and parking lots) or large water bodies (such as lakes and reservoirs).

We selected a subset of these variables for MGWR by removing those with variance inflation factors greater than 10 based on OLS regression, and by using Akaike's information criterion (AIC; Burnham & Anderson, 2002) to select a single parsimonious set. The selected set of variables was then used to explain the spatial pattern of residuals from the pooled niche model using MGWR [\(https://](https://sgsup.asu.edu/sparc/multiscale-gwr) [sgsup.asu.edu/sparc/multiscale-gwr\)](https://sgsup.asu.edu/sparc/multiscale-gwr), implemented in Python 2.7.10 (Python Software Foundation; <http://www.python.org>) with the "mgwr" package (Oshan, Li, Kang, Wolf, & Fotheringham, 2018) with adaptive bandwidths. MGWR uses an iterative back-fitting algorithm that is computationally intensive (Fotheringham et al., 2017). We therefore thinned the calibration data to 3 per 10 km² to reduce computation time, resulting in a dataset with 2,156 records. This thinning also created a uniform density of observations across the study area to minimize bias towards the more heavily sampled spe‐ cies, *G. agassizii*. MGWR can use *adaptive* bandwidths as estimates of spatial scale, and therefore, we use a Gaussian spatial kernel for each explanatory variable to allow each variable to converge on a separate bandwidth using AIC with small sample correction to avoid overfitting (Fotheringham et al., 2017). Non‐linear regression coef‐ ficients were not considered as they have not been implemented in MGWR, and because at very local scales, species–environment re‐ lationships are expected to approximate linear responses due to the limits in the local range of each explanatory variable (Fotheringham et al., 2002). Bandwidths and their approximate spatial scale are re‐ ported for each explanatory variable along with local parameter estimates and model R^2 . We report the spatial scale of each explanatory variable as the product of the average distance between locations in our calibration dataset and the bandwidth, and use inverse dis‐ tance weighted interpolation to create regression coefficient maps from local parameter estimates for areas that were thinned prior to running MGWR.

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2.6 | **Habitat–genotype association**

We hypothesized that landscape patterns in the interpolated MGWR coefficient maps (representing spatially varying species–environ‐ ment relationships) would be congruent with previously reported phylogenetic differences found in the secondary contact zone iden‐ tified by Edwards et al. (2015). We represented the phylogenetic structure of sampled populations using the admixture proportion (Q) of a pure *G. agassizii* genotype from STRUCTURE 2.3.4 (Pritchard, Stephens, & Donnelly, 2000). This index, hereafter genotype association index, represents the probability that an individual contained *G. agassizii* genotypes (Edwards et al., 2015) and was interpolated

across our study area using inverse distance weighting to create a map for the Mojave genotype. We used Kendall's rank correlation coefficient (tau) for paired samples to assess correlations between each explanatory variable's local regression coefficients and the genotype association index. A nonparametric test was chosen be‐ cause our genotype association index did not meet assumptions of normality.

We then asked if natural divisions in the local species-environment relationships exist, and if present, do they coincide spatially with the genotype association index. We identified divisions with K‐medoids optimal partitioning in multivariate space of the local regression coefficient maps for all explanatory variables using the

FIGURE 3 Probability of presence for (a) the pooled species, (b) *Gopherus agassizii* (Agassiz's tortoise and (c) *Gopherus morafkai* (Morafkai's tortoise). Areas with high probability of presence are shown in red, low in blue. Probability of presence was estimated with Maxent v. 3.4.0. The Colorado River (purple) separates California and Arizona and creates the division between the two species of desert tortoise. USA Contiguous Albers Equal Area Conic projection (SR‐ORG:7301)

0.0 0.1 0.2 0.3 0.4

0.6 0.7 0.8 0.9 1.0 **1202 WILEY Diversity** and **Distributions**

package "cluster" (Maechler, Rousseeuw, Struyf, Hubert, & Hornik, 2016) in R (R Core Team, 2016). The optimal number of clusters was estimated by minimizing within‐cluster variance (Hennig & Liao, 2013). Cluster assignments were mapped back to geographic space and compared to the genotype association index. Here we used spatial autoregressive lag models (SAR lag; Anselin, 2001) with the package "spdep" (Bivand & Piras, 2015) in R (R Core Team, 2016) to determine which of three delineations best explained the genotype association index: (a) geographic boundary (Colorado River) for *G. agassizii* and *G. morafkai*, (b) the ecotone between the Mojave and Sonoran Basin and Range U.S. EPA Level III Ecoregions or (c) the cluster assignments that had been mapped back to geographic space. The SAR lag model is well suited for making spatial predictions when spatial dependencies exist among the values of the dependent variable, as is the case for the genotype association index. Three SAR lag models, each with a single explanatory variable of the (a) geographic division, (b) ecoregional division or (c) mapped clusters of species– environment relationships, were calibrated with a random subset of 2,000 locations to reduce processing time and were compared using AIC to identify the most parsimonious model.

3 | **RESULTS**

3.1 | **Species distribution modelling**

The single set of variables selected to describe the ecological niche of each species included the following: precipitation of the driest month (*Ppt_dry*), precipitation seasonality (*Ppt_CV*), surface texture (*Surf_Text*), soil moisture (*S_moist*), temperature evenness (*Isotherm*), photosynthetic activity (*Veg_Amp*), topographic position index (*Topo_ Index*) and surface material (*Surf_Mat*; see Table 1 for descriptions), and resulted in models for *G. agassizii* and *G. morafkai* with test AUC scores of 0.733 and 0.875, respectively (Figure 3). The pooled model showed reduced performance, with a test AUC score of 0.697, suggesting that the ecological niches of each species were different from one another and not well represented with a single, pooled model.

3.2 | **Niche comparisons**

Relative contributions of explanatory variables were not correlated between species (ρ = 0.432, p = 0.286; Table 2). For example, the explanatory variable *Ppt_dry* contributed the most (40.5%) for *G. agassizii,* but for *G. morafkai*, *Ppt_CV* provided the greatest contribution (38.8%). Correlation in the shape of the spe‐ cies–environment relationships between the two species ranged from 0.033 (*Veg_Amp*) to 0.904 (*S_moist*), indicating that *G. agassizii* and *G. morafkai* differed most in their selection of vegetation greenness, but selected for similar soil moisture characteristics (Table 3). *G. agassizii* and *G. morafkai* showed non‐identical niches based on the niche equivalency test (*p* < 0.0001), with an observed similarity score (ESP = 0.328) significantly lower than would be expected under identical niches. In contrast, the asymmetrical niche similarity tests suggested that *G. agassizii* and *G. morafkai* occupy

TABLE 2 Percent (relative) contributions of each explanatory variable used to define the ecological niche of *Gopherus agassizii* (Agassiz's tortoise; MDT) and *Gopherus morafkai* (Morafkai's tortoise; SDT)

Note: Percent contribution represents the average proportion of the change in regularized training gain across the 100 bootstrap replicates for each variable (see Table 1 for descriptions).

TABLE 3 Pearson's correlation coefficient between species– environment relationships for *Gopherus agassizii* and *Gopherus morafkai* ecological niche models for each explanatory variable (see Table 1 for descriptions)

niches that are *more* similar than would be expected if they occu‐ pied their respective ranges randomly, with an observed similarity (ESP = 0.328) significantly higher than the null distributions for the *G. agassizii* \rightarrow *G. morafkai* comparison (μ = 0.311, σ = 0.006) or the *G. morafkai* \rightarrow *G. agassizii* comparison (μ = 0.236, σ = 0.002). This suggests that while these two allopatric species do not occupy identical ecological niches, they do share niche characteristics and are more similar than would be expected if either species selected habitat at random within their ranges.

3.3 | **Local niche models and spatial scale**

The reduced set of nine principal components identified to investi‐ gate spatial patterns in local species–environment relationships in‐ cluded the 1st and 3rd components of the physiographic PCA (PHYS), the 1st and 3rd components of the climate PCA (CLIM), the 2nd and 3rd components of the soils PCA (SOIL), the 1st and 3rd components of the vegetation PCA (VEG) and the land cover variable (LC). The resulting MGWR model based on these nine explanatory variables had an R^2 of 0.722 and showed local R^2 values that ranged from near

FIGURE 4 Local R^2 from selected multiscale geographically weighted regression model of pooled species SDM residuals. The Colorado River (purple) separates California and Arizona and creates the division between *Gopherus agassizii* (Agassiz's tortoise) and *Gopherus morafkai* (Morafkai's tortoise). USA Contiguous Albers Equal Area Conic projection (SR‐ORG:7301)

0 to 0.999 (Figure 4). The approximate spatial scales (bandwidths) for each explanatory variable were estimated as 68.7 km (87) for PHYS1, 77.2 km (110) for PHYS2, 48.8 km (44) for CLIM1, 341.6 km (2154) for CLIM3, 48.8 km (44) for SOIL2, 48.8 km (44) for SOIL3, 336.7 km (2092) for LC, 86.2 km (137) for VEG1 and 99.6 km (183) for VEG3. Local regression coefficient maps are provided in Figure S1.1.

3.4 | **Habitat–genotype association**

Kendall's tau values representing the degree to which local regression coefficient maps from MGWR were correlated with our genotype as‐ sociation index, ranged from −0.43 to 0.40 (Table 4) and indicated a modest overall agreement between any given species–environment relationship and genotype. However, when considered together, we identified two multivariate clusters in these local regression coefficient maps, which, when mapped back to geographic space, were largely coincident with the boundary separating *G. agassizii* and *G. morafkai* (Figure 5). However, this division between the two clus‐ ters did not exactly coincide with the Colorado River, instead suggesting a boundary approximately 40 km to the east of the Colorado River in the northern portion of the focal study area (Figure 5). The mean genotype association index for the two clusters was 0.98 and 0.15 for the Mojave and Sonoran clusters, respectively, indicating that the Mojave cluster was most strongly associated with the Mojave genotype and the Sonoran cluster was not. The mapped clus‐ ters of local species–environment relationships were better able to predict the genotype association index than either the ecoregions or the geographic delineation between the species, with a ∆AIC score of >2 between the next best SAR lag models (Table 5). Overlap be‐ tween the mapped clusters and the Mojave and Sonoran ecoregions

TABLE 4 Kendall's rank correlation value (Tau) for each local parameter surface of the multiscale geographically weighted regression model with the genotype association index

Variable	τ	\boldsymbol{p}
SOIL ₂	0.169	0.001
SOIL ₃	0.406	0.001
VEG3	-0.411	0.001
PHYS1	-0.437	0.001
PHYS2	-0.030	0.001
CLIM1	0.048	0.001
CLIM ₃	0.065	0.001

Note: Environmental variables are physiographic PCA (PHYS), climate PCA (CLIM), soils PCA (SOIL) and vegetation PCA (VEG), with numbers indicating the component number. Descriptions of component loadings and weights can be found in Tables S1.1–S1.4 in Appendix S1.

suggested that the Mojave cluster was more closely aligned with the Mojave ecoregion than with the current geographic delineation of the Mojave population of *G. agassizii* (Table 6)*.* In contrast, the Sonoran cluster most closely aligned with the current geographic delineation of *G. morafkai*, indicating that the current geographic delineation of *G. morafkai* is a better representation of Sonoran habitat in the focal study area than the Sonoran ecoregion alone (Table 6).

4 | **DISCUSSION**

We explore ecological niche differences between two allopatric species of conservation concern, *G. agassizii* and *G. morafkai*, and find that while these two species occupy broadly similar

FIGURE 5 Two multivariate clusters of habitat selection identified from local regression coefficient maps of species– environment relationships. The Mojave cluster (dark grey) includes a region 40 km east of the Colorado River (blue) where *Gopherus agassizii* (Agassiz's tortoise) individuals have been identified but are not protected under the U.S. Endangered Species Act. The Sonoran cluster (light grey) includes a small region west of the Colorado River, though this area contains only marginal habitat. The Mojave Desert and Sonoran Deserts are outlined with dashed line. USA Contiguous Albers Equal Area Conic projection (SR‐ORG:7301)

TABLE 5 Spatial autoregressive (SAR) lag models fit to the genotype association index

SAR lag model	AAIC	AIC	Rho	Rho <i>p</i>	Wald Statistic	Wald p
Mapped cluster		-5.808	0.74198	< 0.001	46.367	< 0.001
Geographic (taxonomic)	2.7	-3.148	0.79395	< 0.001	68.039	< 0.001
Ecoregion	3.5	-2.301	0.75718	< 0.001	51.007	< 0.001

Note: Each model used one of three explanatory variables (rows) and were compared with AIC.

ecological niches in their respective ecoregions, they differ sub‐ tly in their selection of habitat. Moreover, spatial scale differed among key species-environment relationships, confirming expectations that climate may dominate species' distributions at coarse scales, while responses to topography and land surface characteristics may be more apparent at fine scales (Pearson & Dawson, 2003). Habitat barriers such as water and developed surfaces (i.e., lakes, road and cities) had negative effects on habi‐ tat at coarse scales independent of location, while soil conditions, vegetation and physiographic characteristics exhibited local ef‐ fects that varied across the region encompassing the recent secondary contact zone. We also find that local variation in spe‐ cies–environment relationships provided greater support for the phylogenetic differences observed among individuals than does the current geographic delineation between the two species. Our results contribute additional evidence that *G. agassizii* and hybrid‐ ized individuals east of the Colorado River and west of Kingman AZ (Figure 5) exhibit ecological niches that are more similar to *G. agassizii* in the rest of its range than to their proximal sister taxa, *G. morafkai*.

4.1 | **In support of phylogenetic boundaries: Local species–environment relationships**

We found evidence for two, but not three, multivariate clusters in the local regression coefficient maps. A third category, if coinciding

TABLE 6 Overlap (km² and percent area) between each mapped cluster of habitat use with the current geographic delineations between *Gopherus agassizii* (Agassiz's desert tortoise) and *Gopherus morafkai* (Morafka's desert tortoise), and the Mojave and Sonoran Basin and Range U.S. EPA Level III Ecoregions

with regions containing hybrids, might suggest that hybridized individuals select habitat in ways that are locally different than either of the two pure genotypes. Previous work has shown that these hybrids do occupy habitats with characteristics that span those of both their Mojave and Sonoran parental lineages in terms of topographic, surface textural and vegetation characteristics (Edwards et al., 2015). Our delineation of two categories does not counter these findings because here we explore *local* niche differences, that is local differences in species–environment relationships, rather than differ‐ ences in occupied habitat as was explored by Edwards et al. (2015). For example, consider individuals in one region that occupy areas with values near 10 on a hypothetical environmental gradient (on a scale of 1-10). If surrounding areas have values near 5, these individuals will exhibit positive local species–environment relationships because locally available environments have lower values on this hypothetical gradient. However, in another region, individuals occupying areas with values near 5 may also show positive local species– environment relationships if nearby environments show values of 1. In this simple example, these two groups show similar *positive* local species–environment relationships even though they occupy differ‐ ent regions of this hypothetical gradient (i.e., values of 10 vs. 5).

Using a coupled approach with species distribution modelling and multiscale geographically weighted regression, we find that in‐ dividuals in the secondary contact zone exhibit habitat preferences that are more akin to *G. agassizii* than *G. morafkai* even though some of them occupy habitats that are only marginally different from ei‐ ther parental lineage. This is where coupling SDM with local mod‐ elling methods such as MGWR departs from traditional habitat assessments—differences in local habitat *selection* are uncovered rather than differences in local *occupied* habitat. Local habitat *selection* is evaluated in context of nearby environments, while *occupied* habitat is a measure of differences between occupied areas.

In the case of *G. agassizii* and *G. morafkai*, regional differences in occupied habitat are clearly evident. Differences span climate, vegetation, physiography and geology (Nussear & Tuberville, 2014) and are consistent with the subtle differences we found in their ecological niches when quantified with SDM based on range‐wide species-environment relationships. For example, when we modelled distribution of each species separately, we found differences in species–environment relationships for seasonal vegetation ampli‐ tude (*Veg_Amp*; Figure S1.2), an explanatory variable describing the seasonal vegetation green‐up potential (Meier & Brown, 2014). We also found subtle differences in the range‐wide relationships for sur‐ face texture (*Surf_Text*; Figure S1.2) and topographic position index describing physiographic relief (*Topo_Index*; Figure S1.2). *G. agassizii* tend to occupy regions with finer surface textures such as alluvial soils, while *G. morafkai* occur more often in rocky soils and bajadas (Van Devender, 2006). In contrast, both species share similar range‐ wide unimodal species–environment relationships for summer soil moisture content (*S_moist*; Figure S1.2), thereby avoiding very dry and very wet soils. Similarly, both *G. agassizii* and *G. morafkai* ap‐ pear to have range limits defined by cold winter temperatures, as each can tolerate high summer temperatures through behavioural

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aestivation (Nussear & Tuberville, 2014). This suggests that while the two species occupy different habitats, they exhibit similar selec‐ tion for certain environmental conditions.

In contrast, we found substantial differences in species-environment relationships at local scales, where soils (SOIL2, SOIL3), precipitation (CLIM1) and vegetation (VEG1, VEG3) variables were optimized with short bandwidths. This suggests that local varia‐ tions in the species–environment relationships of these explana‐ tory variables contribute to overall niche differences between the two species. For example, the SOIL2 PCA showed a spatial scale of ~50 km and was most influenced by summer and winter potential evapotranspiration. The local regression coefficient map for this variable tended to show positive species–environment relation‐ ships west of the Colorado River, and negative relationships east of the River, suggesting that individuals west of the Colorado River tended to select habitat with higher potential evapotranspiration given locally available conditions (Figure S1.1). Differences in these local species–environment relationships, such as precipita‐ tion (e.g., summer and winter; CLIM1), terrain (e.g., slope and rock‐ iness; PHYS1) and vegetation (e.g., phenology and canopy growth; VEG1 and VEG3), may drive local adaptation and help maintain population structure of genotypes for *G. agassizii* and *G. morafkai*. Ongoing work suggests that genotypic structure within the Mojave population (those west of the Colorado River) may be maintained by selective pressure on key genes from local environmental dif‐ ferences (Sánchez‐Ramírez et al., 2018). Such environmental dif‐ ferences include a pronounced precipitation seasonality gradient across the combined ranges of *G. agassizii* and *G. morafkai*, with western areas exhibiting high winter (November to March) pre‐ cipitation and few summer monsoonal storms, whereas eastern and southern areas are prone to intense monsoonal storms but little winter precipitation. Local adaptation resulting in local niche differences may help maintain population structure and provide an opportunity for selection to result in speciation. Our analyses comparing local species–environment relationships to the geno‐ type association index lend additional support for this hypothesis. Here, we find that the most parsimonious spatial model explaining the landscape pattern of genotype association was the two multi‐ variate clusters of local species–environment relationships rather than the Mojave and Sonoran ecoregions or the current geo‐ graphic delineation of the two species and their protection status.

4.2 | **Importance for conservation**

Efforts to preserve biodiversity have placed new empha‐ sis on measures of biodiversity beyond taxonomic diversity. Understanding landscape patterns in phylogenetic diversity is es‐ pecially important to conservation goals aimed at maximizing the resilience of biodiversity in the face of rapid global change (Flynn et al., 2011; Legendre et al., 2005), and for identifying conditions where recent lineage divergence has contributed to local niche differences that may aid in adapting to changing environments (Ackerly et al., 2010; Crandall, Bininda‐Emonds, Mace, & Wayne, **1206 WILEY Diversity** and **Distributions**

2000; Moritz, 2002). Identifying spatially structured variation in habitat selection, coupled with an understanding of genotypic structure, is therefore important for predicting potential out‐ comes of spatial conservation decisions (Ferrier & Drielsma, 2010; Whittaker et al., 2005). Often, conservation prioritization focuses on hotspots (Myers et al., 2000; Naeem et al., 2012; Winter et al., 2013) delineated on the basis of taxonomic diversity (Ferrier et al., 2004; Myers et al., 2000), phylogenetic diversity (Crozier, 1997; Helmus et al., 2007; Scoble & Lowe, 2010; Vandergast et al., 2013; Wood et al., 2013) or measures of evolutionary potential, such as sequence diversity (Tamura & Nei, 1993) or divergence (Nei & Li, 1979). However, the ability to compare landscape measures of ge‐ netic diversity to measures of local niche differences and habitat selection (e.g., species–environment relationships) presents new opportunities to investigate the confluence of genetics and ecol‐ ogy in context of conservation biogeography.

Conservation managers tasked as stewards of healthy and sustainable ecosystems often request spatially explicit informa‐ tion that supports management objectives. Lake Mead National Recreation Area, a unit within the National Park Service, is the unit responsible for stewardship of ~6 million ha of land in southern Nevada and northwest Arizona. Park managers seek information on tortoise distributions and lineages in order to prioritize pro‐ tection and restoration of tortoise habitat impacted by invasive weeds, fire, road disturbance, recreation and development (Brooks & Esque, 2002; Esque et al., 2010; Lovich et al., 2011). Our work lends additional evidence that the current geographic boundary of the Mojave Distinct Population Segment (DPS; Department of the Interior, 1990) does not capture the full extent of *G. agassizii* and its niche, and further suggests that local habitat selection in and around the secondary contact zone may contribute to the unique‐ ness of *G. agassizii* currently residing on the eastern side of the Colorado River. Protection and restoration of these areas could further park goals of managing and maintaining tortoise habitat.

4.3 | **A novel coupled modelling approach**

The use of local regression to explore spatial variation in species–en‐ vironment relationships is not new to SDM but has been difficult to apply given the widespread reliance on binary (presence–absence or presence–background) calibration data necessitating logistic regres‐ sion. Local models using a logistic regression framework can suffer from complete separation of response classes at fine spatial scales (Fotheringham et al., 2002), thereby forcing models to use large bandwidths approximating range‐wide models (Miller, 2012). This is especially problematic when calibration datasets exhibit extreme sampling bias. We mitigate against this problem by calibrating a local MGWR model on the residuals of a pooled niche model from both taxa to explore local deviation in species–environment relationships. Modelling residuals enable the use of local Gaussian models, and multiscale local regression methods allow more flexible application of the method to ecological data where relationships between re‐ sponse and explanatory variables may be apparent at varying scales.

Cases of allopatric speciation are especially well suited to these coupled methods because gradients in ecological niche conserva‐ tion across two taxa can be explored at a local level within hypoth‐ esized mixture zones. Moreover, delimiting small regions of interest is necessary when computationally intensive MGWR models require extreme processing times due to their use of iterative back-fitting algorithms to fit optimal bandwidth vectors (Fotheringham et al., 2017). In contrast, presence–background SDM methods assume that the entirety of a species' range is sampled (Elith & Leathwick, 2009; Franklin, 2010) and require large study areas. This coupling approach allows each method to use an appropriate spatial domain for its respective model assumptions and computation limitations. In this way, MGWR can be used in a subregion of the species' distribu‐ tion to explore local variation in species–environment relationships as expressed in deviations from these predictions, that is residuals.

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DATA AVAILABILITY

All relevant data are hosted by ScienceBase, a public repository hosted by the U.S. Geological Survey. Data can be accessed at the following DOI: [https://doi.org/10.5066/P91V2S8C.](https://doi.org/10.5066/P91V2S8C)

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BIOSKETCH

Richard Inman's research interests include understanding how anthropogenic induced land‐surface change is altering species distributions through the use of GIS, landscape simulations and species distribution modelling. Interests of the team of authors includes arid systems ecology, spatial analysis, biogeography, and landscape genetics.

Author contributions: R.I., S.F. and J.F. conceived the ideas, and R.I. wrote the first version. All authors contributed to the writing and revised all versions of the manuscript.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

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