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Article

A Divergent Synthesis of Numerous Pyrroloiminoquinone Alkaloids Identifies Promising Antiprotozoal Agents

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Babesia. In combination with evaluations of their human cytotoxicity, we identified several compounds with potent (low-nM IC_{50}) antimalarial and antibabesial activities that are much less toxic toward mammalian cells and are therefore promising lead compounds

intermediate. Critically, this compound was readily diverged via selective methylation of either (or both) of the imine-like or pyrrole nitrogens, which then permitted further divergence by either *O*-demethylation to *o*-quinone natural products or displacement of the methoxy group with a range of amine nucleophiles. Based on a single, early report of their potential utility against the malaria parasite, we assayed these compounds against several strains of *Plasmodium falciparum*, as well as two species of the related protozoan parasite

The diversity of alkaloids generated by the oxidation of

tryptophan or tryptamine is astounding, and their biological

activities show remarkable breadth. Among this rather loose

collection of secondary metabolites, the pyrroloiminoquinone

(PIQ) alkaloids (Figure 1) vary in complexity from the

relatively simple makaluvamines A, C, H, I_{r}^{1} and N $(1-5)_{r}^{2}$

which are barely decorated variants of the basic PIQ ring

system, to the polycyclic, polyfunctional discorhabdins A, B,

Beyond the proposed biosynthetic construction of the core

ring system by oxidation of tryptamine that gives rise to the damirones⁷ and batzellines,^{8,9} further complexity arises via

incorporation of an ammonia equivalent to give the simple

makaluvamines and by inclusion of arylethylamine groups on

the PIQ scaffold to give alkaloids such as makaluvamine G

 $(6)^{10}$ and its congeners. Fused or spirocyclic PIQs such as tsitsikammamine C $(16)^{11}$ and discorhabdins, respectively

then arise via oxidative C-C bond formations of electron-rich

aromatics or benzylic-type C-H bonds. Clearly, nature makes

opportunistic use of oxidizable functionality in the biosynthesis

of this family, and does not stop there, as demonstrated by its most complex members. It is likely that chemists have only scratched the surface of the PIQ family's structural diversity.

C,⁴ and V,⁵ (17–20) and aleutianamine $(22)^6$ (Figure 1).



The most prevalent biological activity reported for the PIQs

B. duncani: 30 nM

B. divergens Rouen: 1.6 nM

methylation is key to activity!

In a single publication by Quinn and co-workers, a small subset of PIQs isolated from a sponge of the genus *Zyzzya* was shown to have promising activity against both chloroquine-sensitive and multidrug-resistant strains (3D7 and Dd2,

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O₂CCF₃

NH



ACS Publications

for antiprotozoal drug discovery.

INTRODUCTION

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Figure 1. Representative examples highlighting structural diversity and biological activity within the pyrroloiminoquinone class of alkaloids. All data given are IC_{50} values.

respectively) of the human-malaria-causing parasite Plasmodium falciparum.¹¹ Makaluvamines G, J, and L (6, 7, and 9), as well as tsitsikammamine C (16) each demonstrated IC_{50} values below 50 nM against each strain. Importantly, these compounds were significantly less toxic toward human embryonic kidney cells (HEK-293); therapeutic indices (TI) with respect to this sample human cell line were thus \sim 50 or greater. Tsitsikammamine C (16) was both the most potent antimalarial and least cytotoxic toward HEK-293 cells, with an SI of \geq 200. Despite this report from over a decade ago, it appears that there has been little follow-up on the antimalarial potential of the PIQs. Notably, a few related discorhabdins were potent inhibitors of D6 and W2 clones of P. falciparum in *vitro*; however, they proved toxic in *P. bergheii*-infected mice.¹⁴ In general, tricyclic PIQs are less potent cytotoxins than the more complex discorhabdins.

Our group has a strong track record in the synthesis of antimalarial natural products and designed analogues, and collaborative investigations into their structure/activity relationships (SAR) and mechanisms of action.^{15–18} Babesiosis is a related, tick-borne disease caused by protozoan parasites of the genus *Babesia*.¹⁹ With its prevalence dramatically increasing worldwide, and with no dedicated therapeutic agents against it, babesiosis is often treated with antimalarial agents, with varying levels of success.^{20–27} As such, there is considerable pressure to develop targeted antibabesial chemotherapeutics.¹⁹

Here, we report how we have leveraged an optimized synthesis of the core heterocyclic scaffold of the PIQ alkaloids, along with a critical selective *N*-methylation strategy, to access numerous members of the class.²⁸ We provide significant new

data on the potent antimalarial and antibabesial activity of these promising compounds.³¹

BACKGROUND

The first synthesis of a marine alkaloid of the PIQ class was accomplished over 30 years ago by Yamamura and co-workers, who demonstrated the utility of an electrophilic methoxy iminoquinone of type 27 (Scheme 1).³²⁻³⁶ This landmark achievement demonstrated the clean reactivity of 27 with primary amine nucleophiles, which is well-suited for convergent synthesis applications. This approach eventually enabled access to a handful of makaluvamines, batzellines and discorhabdins. Thus, many groups have targeted similar intermediates en route to PIQ targets. This is a rich area of heterocycle synthesis that has been frequently, but not comprehensively, reviewed.^{12,13,37-42} In general, this electrophilic PIQ tricycle is formed via one of three overarching approaches (Scheme 1) (A) pyrrole annulation onto a functionalized quinoline intermediate pioneered by Joule, inspiring a handful of similar approaches $^{43-51}$; (B) cyclization of a 4-aminoindole, as was first executed by Yamamura and later used by several other groups^{32,36,43,52,53}; and (C) a biomimetic strategy in which an oxidized tryptamine equivalent enables cyclization of a pendent amine onto an aromatic ring, followed by oxidation to the iminoquinone.⁵⁴

Of these approaches, the biomimetic cyclization strategy has inspired the most innovation because of its value in greatly simplifying the construction of the PIQ core, prompting the application of state-of-the-art C–N bond-forming methodologies. Major advancements to this end include the use of Scheme 1. Strategies for the Synthesis of Versatile PIQ Core Intermediates of Type 27



aryne chemistry, 5^{9-64} transition metal-catalyzed cross-coupling and oxidative coupling reactions. $6^{7-70,64}$ The latter two strategies were employed successfully in very recent total syntheses of PIQ alkaloids by the groups of Tokuyama, Wood, Stoltz and Burns. 65,71-73

Despite the many advances in PIQ total synthesis since Yamamura's first success, only a handful of groups elaborated their tricyclic intermediates to access a range of PIQ alkaloids. The lengthy syntheses required to elaborate commercially available materials to appropriately functionalized tryptamine precursors as a prelude to assembly of the PIQ core apparently limited their practical applications to PIQ alkaloid synthesis. The Burns group made N-Ts damirone C via a short sequence of 6-7 steps, and they advanced this intermediate to complete formal syntheses of several natural products; however, overall, the o-quinone motif salient to the damirones has proven considerably less useful for amine condensation compared to the vinylogous imidate found in compounds like 27.70 One of the key missing elements in an approach able to afford large numbers of PIQ alkaloids from a common core is the selective access to the four different N-methylation patterns found in these alkaloids. Given the apparent impact of the methylation pattern on both antimalarial activity and cytotoxicity (see below), the ability to control methylation is critical to advancing our understanding of the biological potential of the PIQ alkaloids.

RESULTS AND DISCUSSION

Synthesis of PIQ Alkaloids and analogs. The strategy we developed to interrogate antiparasitic PIQ structural space is depicted in Scheme 2. Central to this work was our aim to access members of the structurally related makaluvamine, damirone, batzelline, isobatzelline and tsitsikammamine natural products to establish structure/activity relationships (SAR) among and within these natural product families. While a handful of other groups have used synthetic PIQs to

Scheme 2. Plan to Prosecute Divergent Syntheses of PIQ Alkaloids



interrogate SAR, these studies were confined to anticancer activity. $^{30,74-82}_{\ }$

Despite the significant body of synthetic work focusing on PIQs that hints at the possibility of a divergent synthesis, no single approach has been able to efficiently link many related PIQ natural product families to a common synthetic intermediate and produce a large number of congeners.^{35,66} This problem stems largely from the early installation of key functional groups, especially the *N*-methyl groups. These limitations are also likely related in many cases to long synthetic sequences and perceived instability of the PIQ core ring system.^{54,58} Nonetheless, it was clear that developing a late-stage divergent synthesis would vastly improve our ability to access a library of PIQs with the goal of learning more about their antiprotozoal activity; thus, we embarked upon a concise scalable synthesis of the known electrophilic PIQ core **33**, from which we would implement maximally divergent chemistry.

We were attracted to the hypervalent-iodine-mediated oxidative cyclization chemistry developed by Kita and coworkers to forge the piperidine ring of 27.67,68 However, preparation of the methoxy indole precursor bearing the critical azide requires a lengthy synthesis (see Supporting Information for details) using existing technology.^{30,83} We thus designed a concise synthesis of PIQ 33 (Scheme 2) that centered on a strategic Larock disconnection, which permits rapid access to substituted tryptophols or tryptamine derivatives, thus rapidly setting up for Kita's oxidative cyclization. At the outset of our work, the Larock disconnection had yet to be employed in the preparation of pyrroloiminoquinone alkaloids, despite its identification as an important strategy for the synthesis of tryptophan-based natural products.^{84–86} Very recently, Wood and co-workers documented the utility of a Larock indole synthesis coupled with a Kita-type oxidative cyclization en route to makaluvamines A and K; however, the overall sequence to intermediates of type 27 was >10 steps, LLS, although the indole synthesis was shown to be quite scalable.⁷² Additionally, the Stoltz group reported an attractive tandem Larock/ Buchwald-Hartwig process to assemble a reduced variant of the PIQ core, which was then used to make five alkaloids.⁶⁶ As we show here, our approach compares favorably to these recent achievements, as we are able to procure 16 alkaloids via our highly optimized approach in sequences that are in most cases multiple steps shorter.

The synthesis of tryptophol **39** (Scheme 3) began with the construction of the iodoaniline building block **35** via a

Scheme 3. Multigram Scale Synthesis of Building Block 33



regioselective iodination (4:1 rr, 69% yield of 35) with ICl under biphasic conditions.⁸⁷ Known alkyne building block 36 (prepared in one step)⁸⁸ underwent Larock annulation with 35 in the presence of K_2CO_3 and Pd/C (1 mol %) to generate indole 39 in 56% yield following acidic workup or in 75% yield over two steps by employing a discrete hydrolysis of the crude bis-silylated indole intermediate.⁸⁸ Regardless, the bis-silylated immediate annulation product was sensitive to partial desilylation during purification and we found it most convenient to remove both TES groups prior to chromatographic purification. Deoxyazidation of known tryptophol 3989 using diphenylphosphoryl azide and DBU delivered azide 34 in high yield.⁹⁰ Acetylation of the indole nitrogen of 34 provided the requisite oxidative cyclization precursor 40. The optimized yield for the PIFA-mediated oxidative cyclization of 40 reported by Kita and co-workers is 51%.⁶⁸ Because we were unwilling to concede such a loss of material, we further optimized this key transformation. While conducting a screen of reaction solvents commonly used for this type of transformation, we uncovered the importance of order of reagent addition. Specifically, we observed significantly improved reaction profiles and higher isolated yields when H₂O was added following the addition of the other reagents, thus increasing the yield for PIQ 33 to 78%, thereby easing purification and improving material throughput.

Using the optimized synthetic sequence (Scheme 3), we generated >11 g of the key PIQ intermediate 33 in a week of lab time, starting from aniline 37. Importantly we observed that both the freebase and trifluoroacetic acid (TFA) salt of this compound are significantly more stable than previously reported.⁵⁴ We observed that while 33 and other PIQs (see below) are highly reactive in solution,⁹¹ only minor degradation of a solid sample stored at -20 °C was detected over a period of greater than 6 months. The remarkable stability of the TFA salt of PIQ 33 is highlighted by the ability to generate crystals of sufficient quality for single crystal X-ray

analysis through the slow evaporation of a MeOH solution on the benchtop.

With ample quantities of tricyclic electrophile 33 in hand, we aimed to develop efficient routes to multiple makaluvamine and damirone natural products that differ primarily by Nmethylation pattern. Quinn and co-workers' study suggested that *N*-methylation significantly affects the antimalarial activity PIQ alkaloids, with those bearing N-methylated iminium functions displaying the greatest potency.¹¹ To investigate the generality of this observation, we required a means to selectively methylate both the imine and the pyrrole. In almost all prior work methylation at each of these sites is carried out before the formation of the iminoquinone.^{61,62,92} We looked to develop conditions for the direct methylation of 33 (Scheme 4), thus allowing for strategic diversification first to different Nmethylated variants of 33 and from those intermediates to makaluvamines by amine condensation and damirones by Odemethylation.

Initial attempts at exhaustive methylation of the TFA salt of 33 using the best literature precedent (K_2CO_3 and MeI in $DMF^{61,62}$) were met with nonspecific decomposition. Fortunately, switching to Ag₂CO₃ and MeI reproducibly yielded the desired bis-methylated PIO 41 (48%), along with the imine-methylated PIQ 42 (11%), after careful optimization. Switching to the freebase of 33 enabled selective methylation of the pyrrole and imine N-atoms. The higher nucleophilicity of the imine relative to the pyrrole allowed the imine methylated product 42 to be obtained selectively with MeI in CH₂Cl₂/MeOH. The N-methylated pyrrole 43 was accessed by revisiting the original K₂CO₂, MeI, DMF conditions with the freebase of 33. Careful optimization of the conditions was required to ensure complete conversion while minimizing decomposition, ultimately affording pyrrole N-methylated product 43 in 63% yield. In this way, we gained access to four electrophilic, tricyclic PIQ cores with all possible methylation patterns of the two nitrogen atoms; these could be used in either O-demethylation reactions to deliver the oquinone damirone and batzelline alkaloids, or by amine condensation to afford makaluvamines.

We accessed makaluvamine I (4) and D (44) from the TFA salt of 33 via aminolysis with NH4Cl and tyramine as amine sources, respectively.^{35,49,63} The inclusion of sat. aq. NaHCO₃ ensured the complete conversion of the TFA salt of 33 to aminolysis products, especially when the reactants were also ammonium salts; inconsistent levels of conversion resulted when it was omitted. The synthesis of the ortho-quinone alkaloid damirone C (12) from 33 proceeded via NaImediated demethylation of the vinylogous imidate. This reaction, inspired by the conditions employed by Tokuyama in their synthesis of damirone B (11),⁶¹ provided damirone C (12).¹ If aqueous workup procedures were used, a significant quantity of 6-iodo-damirone C (46, Scheme 5) was isolated in conjunction with 12. Based on control experiments with electrophilic halogenating agents (see below), this reaction likely occurred via the air oxidation of iodide anion. Interestingly, this product was not observed if aqueous workup was omitted. The optimized conditions for demethylation provided damirone C in 48% yield. The chemistry described for the synthesis of makaluvamines I and D, and damirone C, proved general for the synthesis of their differentially methylated congeners as shown in Scheme 4c-e, with the exception of damirone A, which was generated under different conditions (see Supporting Information). This synthesis of 11

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Scheme 4. Synthesis of 11 PIQ Alkaloids^a



 $a^{\prime}(a)$ Selective methylation of PIQ core 33. (b-e) Conversion of each differentially methylated core to PIQ alkaloids. For details, see text and Supporting Information. *Yield calculated on the basis of purity determined by Q-NMR.

Scheme 5. Synthesis of Halogenated PIQs, Tsitsikammamine A, Makaluvamine M, and Makaluvamine L



PIQ alkaloids served as proof-of-principle that divergent methylation of the PIQ core would be a powerful strategy in this area of natural product space.

The halogenated congeners of the makaluvamines and damirones, known as the isobatzellines and batzellines respectively, have only scarcely been evaluated for antimalarial activity.²⁹ Though the electrophilic introduction of halogen atoms via the enaminone of the damirones is well-precedented,^{64,69,73} the direct introduction of halogens onto the iminoquinone framework had not been accomplished. We document that both reactivities are efficient (Scheme 5), by showing chlorination of damirone C (12) to batzelline D (14) under modified literature conditions,⁶⁹ and the conversion of makaluvamine I (4) to makaluvamine N (5) upon treatment with NBS in methanol.

We made three final PIQ natural products. Condensation of 33 with aminoalcohol 47 led to 48, which could be directly dehydrated with BF3:OEt2 in aprotic solvent at elevated temperature to give the conjugated side chain of makaluvamine M (50). Interestingly, treatment of this same intermediate with the same Lewis acid in polar protic solvent at room temperature effected cyclization to pyrroline (51), which was easily oxidized to give the fused tetracyclic core of the tsitsikammamines (in this case tsitsikammamine A, 52). At this point, we had only made these two compounds from nonmethylated electrophilic PIQ core 33; of course, our access to the differentially methylated cores provides a way to make the congeners of makaluvamine M and tsitsikammamine A with different methylation patterns. As a first step to corroborate this assertion we synthesized makaluvamine L(9)from imine-methylated PIQ 42 via the same two-step sequence applied to access makaluvamine M (50).

Overall, our doubly divergent approach led to the first syntheses of makaluvamines H, L, and M. With respect to alkaloids that had been previously made, we accomplished the shortest syntheses of batzelline D; damirones A, B, and C; makaluvamines I, J, K, N, and P; as well as tsitsikammamine A. Further, with the advent of our divergent methylation protocol that permits ready access to all four electrophilic vinylogous imidate PIQ cores, we have a simple blueprint to easily make many analogues by amination. For example, on the basis of the strong activity (see below) of the imine N-methylated makaluvamines bearing phenethylamine-type side chains (makaluvamines J and P), we aminated 42 with O-methyltyramine and tryptamine, arriving at compounds 53 and 54 (Figure 2), the latter of which was very recently reported as a potent inhibitor of PANC-1 cells.⁹² These compounds, as described below, are particularly potent antiprotozoal agents.

Critical Application of Q-NMR for Many Intermediates. Despite the vast literature surrounding PIQ syntheses, few groups have interrogated the purity of their PIQ intermediates or final products beyond NMR spectroscopic homogeneity. In their syntheses of discorhabdin natural



Figure 2. Two unnatural analogs made by amination of 42.

products, Heathcock and co-workers claimed that "inorganic salts often seemed to contaminate the product, so extra care was necessary to ensure that the product was pure".58 However, no further description was given as to the nature of these impurities. Additionally, hygroscopicity of charged PIQs is precedented. In the report of its isolation, discorhabdin C was characterized by X-ray diffraction as the HCl salt, which was hydrated by no less than six water molecules per unit cell.⁴ In our studies, we found that some positively charged PIQ intermediates had unusually high affinity for water and organic solvents. In most cases, bulk water could be removed by interconversion to the corresponding neutral, freebase form, followed by routine desiccation methods. However, when the freebase PIQ was inaccessible (such as the N-Me iminium ions), or when hydroscopic PIQs were obtained, our best efforts were made to assign their purity using quantitative NMR spectroscopy in the presence of an internal standard. Additionally, we seized every opportunity to characterize our products by X-ray diffraction to better understand their solidstate composition. In doing so, we hoped to bolster the validity of our isolated yields as well as the biological assays discussed below.

Biological Assessment of 25 PIQ Alkaloids and Analogues. Our goal for the biology of these PIQ alkaloids was to delve more deeply into the antiprotozoal activity of these compounds, to recapitulate the potent antimalarial activity and high therapeutic index reported by Quinn for makaluvamine J (7) and related compounds, and to gain a better sense of SAR and selectivity with respect to toxicity to a broader range of pathogens and human cell lines. For this purpose, we assayed 25 compounds-a combination of 12 natural products and 13 synthetic analogs-for antimalarial activity (Plasmodium falciparum drug-resistant 3D7, Dd2, and W2 strains), antibabesial activity (Babesia duncani, and B. divergens Rouen strains), several strains of three different species of fungus, and four human cell lines. A summary of the data is presented in Figure 3 with heat mapping/color-coding; IC50 values for the antimalarial and antibabesial activity are shown along with the therapeutic index (TI) calculated against the average cytotoxicity of the compounds against the four human cell lines (complete data sets can be found in the Supporting Information).

The two representative methoxy-PIQ tricycles 33 and 42 are micromolar antiprotozoals, with moderate TIs. o-Quinones damirone C (12), batzelline D (14), and analog PIQ-5 (46) are poorly active antiprotozoals and show virtually no TI. In the cases of the PIQs derived from ammonolysis of the methoxy core, submicromolar activities are observed for makaluvamines I (4) and C (2), but only the latter shows any modest TI. Here, we can begin to observe the importance of methylation patterns on SAR: pyrrole N-methylation trends with poor activity and imine N-methylation seems to bolster activity and attenuate toxicity against mammalian cell lines.

In those PIQs with tyramine-derived side chains, there is a general increase in potency as compared with the simpler compounds. The standouts in all cases are those with the imine *N*-methylated, as in makaluvamines J (7), P (45), PIQ-1 (53), PIQ-2 (54), and makaluvamine L (9). Again, pyrrole *N*-methylation (compare makaluvamines J and P) slightly decreases activity and slightly increases cytotoxicity for an overall decrease in TI. Pyrrole bromination as in PIQ-12 (61) abrogates activity compared to the parent makaluvamine D, as does tyramine phenol bromination, as in PIQ-11 (60) and



Figure 3. Biological data for 25 pyrroloiminoquinones assayed against malaria- and babesiosis-causing protozoan parasites, as well as their therapeutic indices. 3D7, Dd2, and W2 are representative drug-resistant strains of *Plasmodium falciparum*; dun represents *Babesia duncani*, div represents *Babesia divergens* Rouen strain (all numbers are IC₅₀ values in μ M). Av. TI is the therapeutic index (unitless) arrived by comparing the activity against the protozoan parasites with the average value of the cytotoxicities against the human cell lines HCT-116, HEK-293, HeLa, and HepG2.

PIQ-13 (62). O-Methylation (PIQ-1, 53), however, increases activity, with a concomitant increase in cytotoxicity; however, this compound shows 3.5 nM activity against *B. divergens* Rouen and maintains a promising TI. The other compound that is noteworthy is PIQ-2 (54), bearing a tryptamine motif; it shows potencies \leq 31 nM in all assays (9 nM against *P. falciparum* 3D7 and <2 nM against *B. divergens* Rouen). Tsitsikammamine A, the only PIQ with a tetracyclic core that we tested, was significantly less potent than the analogous uncyclized compound makaluvamine D.

It is important to recognize that TI values naturally decrease as the antiprotozoal activity decreases; the raw data in the Supporting Information will indicate, for example, that makaluvamine K, which is roughly an order of magnitude less active than makaluvamine D, is not that much more cytotoxic.

The combination of an *N*-methylated imine core with a lipophilic side chain in many cases leads to potent antiprotozoal activity. In some cases, the increase in potency is accompanied by an increase in cytotoxicity; however, we find that the whole class of compounds are very poorly active in antifungal models (little activity up to 100 μ M concentrations, see the Supporting Information), indicating that they are not just broadly toxic. Clearly, there are levers that can be pulled to try to maximize antiprotozoal potency while minimizing mammalian cell toxicity; these will include the identity of the greasy side chain and the choices of groups with which to alkylate the imine nitrogen atom.

As mentioned above, significant emphasis has previously been placed on the discorhabdin family of PIQs as a result of their potent cytotoxicity. Most—but not all—of the simpler PIQs are significantly less cytotoxic than the discorhabdins, and many have other potentially important activities, are far easier to access, and appear to have promising selectivity. It is still worthy of note that some of the simpler alkaloids—such as makaluvamine J (54 nM PANC-1^{1,1,3,91}) do have potentially valuable anticancer activities. Clearly, an understanding of the mechanisms of action for both their antiprotozoal activity and human cytotoxicity will be of great value in determining the future potential of this class of compounds.

CONCLUSIONS

We have developed an efficient synthesis of the PIQ alkaloids that has permitted extensive biological evaluation of numerous natural products and structural analogues. The effectiveness of this route hinged on a scalable Larock indole synthesis that enabled access to >10 g batches of the known, key electrophilic PIQ core structure 33, which was then diverged in two stages. A critical and enabling advance is found in the selective Nmethylation protocols that delivered four different electrophiles that could then be further diverged either via Odemethylation to afford damirones or substitution with amines to yield makaluvamines. Several other PIQs were made from key intermediate 33, leading to a total of 16 natural alkaloids; multiple unnatural analogues were also obtained. Three of the natural products were synthesized for the first time, and 11 of the remaining 13 were made via the shortest sequence to date, despite the significant quantity of prior art in this area.⁵ Notably, our syntheses of tsitsikammamine A (51) (8 steps LLS) and makaluvamine N (5) (7 steps LLS) access these natural products in just over half the number of steps of previously reported approaches (15 and 12 steps LLS respectively).66,78

Inspired by a seminal report on their potential as antimalarial agents, we evaluated these compounds against drug-resistant strains of Plasmodium falciparum, the related protozoan parasites Babesia duncani and Babesia divergens,^{94,95} several species and strains of yeast, and four human cell lines. These efforts revealed, in particular, that imine N-methylated makaluvamines and analogues are extremely potent antiprotozoal agents. We recapitulated the earlier results of Quinn by confirming that these compounds are indeed potent (sub-50 nM IC₅₀) antimalarials, but we also showed for the first time that they are even more potent (IC₅₀ = 2-4 nM for 53 and 54) against B. divergens Rouen, which causes babesiosis, a tickborne illness of ever-increasing concern that lacks effective front-line therapeutics.94 We also learned that these potent PIQ antiprotozoal agents are not simply broad toxins, as they show little antifungal activity and many have significant therapeutic indices with respect to human cell lines.

This work represents only the beginning of our SAR on PIQ alkaloids with respect to antiprotozoal activity. On the basis of the results described below, we aim to evaluate different pyrrole and imine *N*-capping groups on the makaluvamine scaffold, and we will interrogate the impact of the largely hydrophobic substituents on the N9 position.⁹² The refined SAR obtained from these studies will set the stage for the design and application of natural-product-derived chemical probe compounds for detailed mechanism of action studies, following collaborative workflows that we have recently used successfully in the area of malaria chemotherapeutics.⁹⁶

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/jacs.4c11897.

Supplemental discussion of reactivity studies; experimental procedures for the synthesis of new compounds; tabulated spectral data supporting the structural assignment of these compounds; NMR spectra for these compounds; X-ray diffraction information for compounds **2**, **12**, **13**, **14**, **33**, and **46** and biological assay descriptions and results; and summary of previous synthetic efforts in this area (PDF)

Accession Codes

CCDC 2378804–2378810 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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Notes

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REFERENCES

(1) Schmidt, E. W.; Harper, M. K.; Faulkner, D. J. Makaluvamines H–M and Damirone C from the Pohnpeian Sponge *Zyzzya fuliginosa*. *J. Nat. Prod.* **1995**, *58*, 1861–1867.

(2) Venables, D. A.; Concepción, G. P.; Matsumoto, S. S.; Barrows, L. R.; Ireland, C. M. Makaluvamine N: A New Pyrroloiminoquinone from *Zyzzya fuliginosa. J. Nat. Prod.* **1997**, *60*, 408–410.

(3) Perry, N. B.; Blunt, J. W.; Munro, M. H. G. Cytotoxic Pigments from New Zealand Sponges of the Genus *Latrunculia*: Discorhabdins A. *B and C. Tetrahedron* **1988**, *44*, 1727–1734.

(4) Perry, N. B.; Blunt, J. W.; McCombs, J. D.; Munro, M. H. G. Discorhabdin C, a Highly Cytotoxic Pigment from a Sponge of the Genus *Latrunculia*. J. Org. Chem. **1986**, *51*, 5476–5478.

(5) Antunes, E. M.; Beukes, D. R.; Kelly, M.; Samaai, T.; Barrows, L. R.; Marshall, K. M.; Sincich, C.; Davies-Coleman, M. T. Cytotoxic Pyrroloiminoquinones from Four New Species of South African Latrunculid Sponges. *J. Nat. Prod.* **2004**, *67*, 1268–1276.

(6) Zou, Y.; Wang, X.; Sims, J.; Wang, B.; Pandey, P.; Welsh, C. L.; Stone, R. P.; Avery, M. A.; Doerksen, R. J.; Ferreira, D.; Anklin, C.; Valeriote, F. A.; Kelly, M.; Hamann, M. T. Computationally Assisted Discovery and Assignment of a Highly Strained and PANC-1 Selective Alkaloid from Alaska's Deep Ocean. *J. Am. Chem. Soc.* **2019**, *141*, 4338–4344.

(7) Stierle, D. B.; Faulkner, D. J. Two New Pyrroloquinoline Alkaloids from the Sponge *Damiria* sp. J. Nat. Prod. **1991**, 54, 1131–1133.

(8) Sakemi, S.; Sun, H. H.; Jefford, C. W.; Bernardinelli, G. Batzellines A, B, and C. Novel Pyrroloquinoline Alkaloids from the Sponge *Batzella* sp. *Tetrahedron Lett.* **1989**, *30*, 2517–2520.

(9) Chang, L. C.; Otero-Quintero, S.; Hooper, J. N. A.; Bewley, C. A. Batzelline D and Isobatzelline E from the Indopacific Sponge *Zyzzya fuliginosa. J. Nat. Prod.* **2002**, *65*, 776–778.

(10) Carney, J. R.; Scheuer, P. J.; Kelly-Borges, M. Makaluvamine G, a Cytotoxic Pigment from an an Indonesian Sponge *Histodermella* sp. *Tetrahedron* **1993**, *49*, 8483–8486.

(11) Davis, R. A.; Buchanan, M. S.; Duffy, S.; Avery, V. M.; Charman, S. A.; Charman, W. N.; White, K. L.; Shackleford, D. M.; Edstein, M. D.; Andrews, K. T.; Camp, D.; Quinn, R. J. Antimalarial Activity of Pyrroloiminoquinones from the Australian Marine Sponge *Zyzzya* sp. J. Med. Chem. **2012**, 55, 5851–5858.

(12) Králová, P.; Soural, M. Biological Properties of Pyrroloquinoline and Pyrroloisoquinoline Derivatives. *Eur. J. Med. Chem.* 2024, 269, No. 116287.

(13) Lin, S.; McCauley, E.; Lorig-Roach, N.; Tenney, K.; Naphen, C.; Yang, A.-M.; Johnson, T.; Hernadez, T.; Rattan, R.; Valeriote, F.; Crews, P. Another Look at Pyrroloiminoquinone Alkaloids— Perspectives on Their Therapeutic Potential from Known Structures and Semisynthetic Analogues. *Mar. Drugs* **2017**, *15*, 98.

(14) Na, M.; Ding, Y.; Wang, B.; Tekwani, B. L.; Schinazi, R. F.; Franzblau, S.; Kelly, M.; Stone, R.; Li, X.-C.; Ferreira, D.; Hamann, M. T. Anti-Infective Discorhabdins from a Deep-Water Alaskan Sponge of the Genus *Latrunculia. J. Nat. Prod.* **2010**, *73*, 383–387.

(15) Daub, M. E.; Prudhomme, J.; Le Roch, K.; Vanderwal, C. D. Synthesis and Potent Antimalarial Activity of Kalihinol B. J. Am. Chem. Soc. 2015, 137, 4912–4915.

(16) Daub, M. E.; Prudhomme, J.; Ben Mamoun, C.; Le Roch, K. G.; Vanderwal, C. D. Antimalarial Properties of Simplified Kalihinol Analogues. *ACS Med. Chem. Lett.* **2017**, *8*, 355–360.

(17) Roosen, P. C.; Karns, A. S.; Ellis, B. D.; Vanderwal, C. D. Evolution of a Short and Stereocontrolled Synthesis of (+)-7,20-Diisocyanoadociane. J. Org. Chem. 2022, 87, 1398–1420.

(18) Dwulet, N. C.; Chahine, Z.; Le Roch, K. G.; Vanderwal, C. D.
An Enantiospecific Synthesis of Isoneoamphilectane Confirms Its
Strained Tricyclic Structure. J. Am. Chem. Soc. 2023, 145, 3716-3726.
(19) (a) Vannier, E. G.; Diuk-Wasser, M. A.; Ben Mamoun, C.;
Krause, P. J. Basesiosis. Infect. Dis. Clin. North. Am. 2015, 29, 357-370.
(b) Renard, I.; Ben Mamoun, C. Treatment of Human Babesiosis:
Then and Now. Pathogens 2021, 10, 1120.

(20) Krause, P. J.; Lepore, T.; Sikand, V. K.; Gadbaw, J.; Burke, G.; Telford, S. R.; Brassard, P.; Pearl, D.; Azlanzadeh, J.; Christianson, D.; McGrath, D.; Spielman, A. Atovaquone and Azithromycin for the Treatment of Babesiosis. N. *Engl. J. Med.* **2000**, *343*, 1454–1458.

(21) Gray, J. S.; Pudney, M. Activity of Atovaquone against Babesia Microti in the Mongolian Gerbil. *Meriones unguiculatus. J. Parasitol.* **1999**, 85, 723–728.

(22) Wittner, M.; Lederman, J.; Tanowitz, H. B.; Rosenbaum, G. S.; Weiss, L. M. Atovaquone in the Treatment of *Babesia microti* Infections in Hamsters. *Am. J. Trop. Med. Hyg.* **1996**, *55*, 219.

(23) Wormser, G. P.; Prasad, A.; Neuhaus, E.; Joshi, S.; Nowakowski, J.; Nelson, J.; Mittleman, A.; Aguero-Rosenfeld, M.; Topal, J.; Krause, P. J. Emergence of Resistance to Azithromycin-Atovaquone in Immunocompromised Patients with *Babesia microti* Infection. *Clin. Infect. Dis.* **2010**, *50*, 381–386.

(24) Dorman, S. E.; Cannon, M. E.; Telford, S. R., III; Frank, K. M.; Churchill, W. H. Fulminant Babesiosis Treated with Clindamycin, Quinine, and Whole-Blood Exchange Transfusion. *Transfusion (Paris)* **2000**, *40*, 375–380.

(25) Marley, S. E.; Eberhard, M. L.; Steurer, F. J.; Ellis, W. L.; McGreevy, P. B.; Ruebush, T. K. Evaluation of Selected Antiprotozoal Drugs in the *Babesia microti*-Hamster Model. *Antimicrob. Agents Chemother.* **1997**, *41*, 91–94.

(26) Shih, C. M.; Wang, C. C. Ability of Azithromycin in Combination with Quinine for the Elimination of Babesial Infection in Humans. *Am. J. Trop. Med. Hyg.* **1998**, *59*, 509–512.

(27) Weiss, L. M.; Wittner, M.; Wasserman, S.; Oz, H. S.; Retsema, J.; Tanowitz, H. B. Efficacy of Azithromycin for Treating Babesia Microti Infection in the Hamster Model. *J. Infect. Dis.* **1993**, *168*, 1289–1292.

(28) Radisky, D. C.; Radisky, E. S.; Barrows, L. R.; Copp, B. R.; Kramer, R. A.; Ireland, C. M. Novel Cytotoxic Topoisomerase II Inhibiting Pyrroloiminoquinones from Fijian Sponges of the Genus *Zyzzya. J. Am. Chem. Soc.* **1993**, *115*, 1632–1638.

(29) Hu, J.-F.; Schetz, J. A.; Kelly, M.; Peng, J.-N.; Ang, K. K. H.; Flotow, H.; Leong, C. Y.; Ng, S. B.; Buss, A. D.; Wilkins, S. P.; Hamann, M. T. New Antiinfective and Human 5-HT2 Receptor Binding Natural and Semisynthetic Compounds from the Jamaican Sponge *Smenospongia aurea. J. Nat. Prod.* **2002**, *65*, 476–480.

(30) Wada, Y.; Harayama, Y.; Kamimura, D.; Yoshida, M.; Shibata, T.; Fujiwara, K.; Morimoto, K.; Fujioka, H.; Kita, Y. The Synthetic and Biological Studies of Discorhabdins and Related Compounds. *Org. Biomol. Chem.* **2011**, *9*, 4959–4976.

(31) Venables, D. A.; Barrows, L. R.; Lassota, P.; Ireland, C. M. Veiutamine. A New Alkaloid from the Fijian Sponge *Zyzzya fuliginosa*. *Tetrahedron Lett.* **1997**, *38*, 721–722.

(32) Cheng, J.-F.; Nishiyama, S.; Yamamura, S. Synthetic Studies on Novel Sulfur-Containing Alkaloids, Prianosins Isolated from the Marine Sponge *Prianos melanos*. Synthesis of the Spirodienone Moiety by Phenolic Oxidation. *Chem. Lett.* **1990**, *19*, 1591–1594.

(33) Nishiyama, S.; Cheng, J.-F.; Tao, X. L.; Yamamura, S. Synthetic Studies on Novel Sulfur-Containing Alkaloids, Prianosins and Discorhabdins: Total Synthesis of Discorhabdin C. *Tetrahedron Lett.* **1991**, *32*, 4151–4154.

(34) Tao, X. L.; Nishiyama, S.; Yamamura, S. Total Syntheses of Batzelline C and Isobatzelline C, the Novel Pyrroloquinoline Alkaloids Isolated from the Marine Sponge *Batzella* sp. *Chem. Lett.* **1991**, *20*, 1785–1786.

(35) Izawa, T.; Nishiyama, S.; Yamamura, S. Total Syntheses of Makaluvamines A, B, C and D, Metabolites of the Fijian Sponge *Zyzza* cf. *marsailis* Exhibiting Inhibitory Activities against Topoisomerase II. *Tetrahedron Lett.* **1994**, *35*, 917–918.

(36) Liang Tao, X.; Cheng, J.-F.; Nishiyama, S.; Yamamura, S. Synthetic Studies on Tetrahydropyrroloquinoline-Containing Natural Products: Syntheses of Discorhabdin C, Batzelline C and Isobatzelline C. *Tetrahedron* **1994**, *50*, 2017–2028.

(37) Antunes, E. M.; Copp, B. R.; Davies-Coleman, M. T.; Samaai, T. Pyrroloiminoquinone and Related Metabolites from Marine Sponges. *Nat. Prod. Rep.* **2005**, *22*, 62–72.

(38) Harayama, Y.; Kita, Y. Pyrroloiminoquinone Alkaloids: Discorhabdins and Makaluvamines. *Curr. Org. Chem.* 2005, *9*, 1567–1588.

(39) Wada, Y.; Fujioka, H.; Kita, Y. Synthesis of the Marine Pyrroloiminoquinone Alkaloids, Discorhabdins. *Mar. Drugs* **2010**, *8*, 1394–1416.

(40) Nijampatnam, B.; Dutta, S.; Velu, S. E. Recent Developments in the Isolation, Synthesis, and Bioactivities of Bispyrroloquinone Alkaloids of Marine Origin. *Chin. J. Nat. Med.* **2015**, *13*, 561–577.

(41) Kalinski, J.-C. J.; Polyzois, A.; Waterworth, S. C.; Siwe Noundou, X.; Dorrington, R. A. Current Perspectives on Pyrroloiminoquinones: Distribution, Biosynthesis and Drug Discovery Potential. *Molecules* **2022**, *27*, 8724.

(42) Hu, J.-F.; Fan, H.; Xiong, J.; Wu, S.-B. Discorhabdins and Pyrroloiminoquinone-Related Alkaloids. *Chem. Rev.* 2011, 111, 5465–5491.

(43) Somei, M.; Yamada, F.; Hamabuchi, S.; Shimizu, A. Simple Total Syntheses of Marine Alkaloids, Batzelline C, Isobatzelline C, Damirone A, and Makaluvamine A. *Heterocycles* **1995**, *41*, 1905.

(44) Venemalm, L.; Estéves, C.; Alvarez, M.; Joule, J. A. Synthesis of a 1,3,4,5-Tetrahydropyrrolo[4,3,2-de]Quinoline from a Quinoline. *Tetrahedron Lett.* **1993**, *34*, 5495–5496.

(45) Roberts, D.; Venemalm, L.; Alvarez, M.; Joule, J. A. Synthesis of Damirones A and B from a Quinoline. *Tetrahedron Lett.* **1994**, *35*, 7857–7860.

(46) Roberts, D.; Alvarez, M.; Joule, J. A. Synthesis of 6-Chloro-1,3,4,5-Tetrahydro-7,8-Dimethoxy-1-Methylpyrrolo[4,3,2-de]-Quinoline from a Quinoline; Formal Total Syntheses of Batzelline C, Isobatzelline C, Discorhabdin C and Makaluvamine D. *Tetrahedron Lett.* **1996**, 37, 1509–1512.

(47) Roberts, D.; Joule, J. A.; Bros, M. A.; Alvarez, M. Synthesis of Pyrrolo[4,3,2-de]Quinolines from 6,7-Dimethoxy-4-Methylquinoline. Formal Total Syntheses of Damirones A and B, Batzelline C, Isobatzelline C, Discorhabdin C, and Makaluvamines A–D. J. Org. Chem. **1997**, *62*, 568–577.

(48) Alvarez, M.; Bros, M. A.; Joule, J. A. Synthesis of Isobatzelline B. *Tetrahedron Lett.* **1998**, *39*, 679–680.

(49) Alvarez, M.; Bros, M. A.; Gras, G.; Ajana, W.; Joule, J. A. Syntheses of Batzelline A, Batzelline B, Isobatzelline A, and Isobatzelline B. *Eur. J. Org. Chem.* **1999**, *1999*, *1173–1183*.

(50) Peat, A. J.; Buchwald, S. L. Novel Syntheses of Tetrahydropyrroloquinolines: Applications to Alkaloid Synthesis. J. Am. Chem. Soc. **1996**, 118, 1028–1030.

(51) Kraus, G. A.; Selvakumar, N. Synthetic Routes to Pyrroloiminoquinone Alkaloids. A Direct Synthesis of Makaluvamine C. J. Org. Chem. **1998**, 63, 9846–9849.

(52) Makosza, M.; Stalewski, J.; Maslennikova, O. S. Synthesis of 7,8-Dimethoxy-2-Oxo-1,3,4,5-Tetrahydropyrrolo[4,3,2-de]Quinoline: A Key Intermediate En Route to Makaluvamines, Discorhabdin C and Other Marine Alkaloids of This Group via Vicarious Nucleophilic Substitution of Hydrogen. *Synthesis* **1997**, *1997*, 1131–1133.

(53) Zhao, R.; Lown, J. W. A Concise Synthesis of the Pyrroloquinoline Nucleus of the Makaluvamine Alkaloids. *Synth. Commun.* **1997**, *27*, 2103–2110.

(54) Kita, Y.; Tohma, H.; Inagaki, M.; Hatanaka, K.; Yakura, T. Total Synthesis of Discorhabdin C: A General Aza Spiro Dienone Formation from O-Silylated Phenol Derivatives Using a Hypervalent Iodine Reagent. J. Am. Chem. Soc. **1992**, 114, 2175–2180.

(55) Sadanandan, E. V.; Cava, M. P. Total Syntheses of Damirone A and Damirone B. *Tetrahedron Lett.* **1993**, *34*, 2405–2408.

(56) Wang, H.; Al-Said, N. H.; Lown, J. W. Convenient Syntheses of Pyrroloiminoquinone and Its Lexitropsin-Linked Derivative. *Tetrahedron Lett.* **1994**, 35, 4085–4086.

(57) Sadanandan, E. V.; Pillai, S. K.; Lakshmikantham, M. V.; Billimoria, A. D.; Culpepper, J. S.; Cava, M. P. Efficient Syntheses of the Marine Alkaloids Makaluvamine D and Discorhabdin C: The 4,6,7-Trimethoxyindole Approach. J. Org. Chem. **1995**, 60, 1800– 1805.

(58) Aubart, K. M.; Heathcock, C. H. A Biomimetic Approach to the Discorhabdin Alkaloids: Total Syntheses of Discorhabdins C and E and Dethiadiscorhabdin D. J. Org. Chem. **1999**, *64*, 16–22.

(59) Iwao, M.; Motoi, O.; Fukuda, T.; Ishibashi, F. New Synthetic Approach to Pyrroloiminoquinone Marine Alkaloids. Total Synthesis of Makaluvamines A, D, I, and K. *Tetrahedron* **1998**, *54*, 8999–9010. (60) Moro-oka, Y.; Fukuda, T.; Iwao, M. The First Total Synthesis of Veiutamine, a New Type of Pyrroloiminoquinone Marine Alkaloid. *Tetrahedron Lett.* **1999**, *40*, 1713–1716.

(61) Oshiyama, T.; Satoh, T.; Okano, K.; Tokuyama, H. Total Synthesis of Makaluvamine A/D, Damirone B, Batzelline C, Makaluvone, and Isobatzelline C Featuring One-Pot Benzyne-Mediated Cyclization–Functionalization. *Tetrahedron* **2012**, *68*, 9376–9383.

(62) Oshiyama, T.; Satoh, T.; Okano, K.; Tokuyama, H. Total Synthesis of Batzelline C and Isobatzelline C. *RSC Adv.* **2012**, *2*, 5147.

(63) Yamashita, Y.; Poignant, L.; Sakata, J.; Tokuyama, H. Divergent Total Syntheses of Isobatzellines A/B and Batzelline A. *Org. Lett.* **2020**, *22*, 6239–6243.

(64) Noro, T.; Sakata, J.; Tokuyama, H. Synthetic Studies on Discorhabdin V: Construction of the A–F Hexacyclic Framework. *Tetrahedron Lett.* **2021**, *81*, No. 153333.

(65) Yu, H.; Sercel, Z. P.; Rezgui, S. P.; Farhi, J.; Virgil, S. C.; Stoltz, B. M. Total Synthesis of Aleutianamine. *J. Am. Chem. Soc.* **2023**, *145*, 25533–25537.

(66) Rezgui, S. P.; Farhi, J.; Yu, H.; Sercel, Z. P.; Virgil, S. C.; Stoltz, B. M. Divergent Total Syntheses of Pyrroloiminoquinone Alkaloids Enabled by the Development of a Larock/Buchwald–Hartwig Annulation/Cyclization. *Chem. Sci.* **2024**, *15*, 12284–12290.

(67) Kita, Y.; Egi, M.; Okajima, A.; Ohtsubo, M.; Takada, T.; Tohma, H. Hypervalent Iodine(III) Induced Intramolecular Cyclization of Substituted Phenol Ethers Bearing an Alkyl Azido Sidechain–a Novel Synthesis of Quinone Imine Ketals. *Chem. Commun.* **1996**, *13*, 1491–1492.

(68) Kita, Y.; Egi, M.; Ohtsubo, M.; Saiki, T.; Okajima, A.; Takada, T.; Tohma, H. Hypervalent Iodine(III)-Induced Intramolecular Cyclization Reaction of Substituted Phenol Ethers with an Alkyl Azido Side-Chain: A Novel and Efficient Synthesis of Quinone Imine Derivatives. *Chem. Pharm. Bull.* **1999**, 47, 241–245.

(69) Backenköhler, J.; Spindler, S.; Spiteller, P. Total Synthesis of Damirone C, Makaluvamine O, Makaluvone, Batzelline C and Batzelline D. *ChemistrySelect* **2017**, *2*, 2589–2592.

(70) Smith, M. W.; Falk, I. D.; Ikemoto, H.; Burns, N. Z. A Convenient C-H Functionalization Platform for Pyrroloiminoquinone Alkaloid Synthesis. *Tetrahedron* **2019**, *75*, 3366–3370.

(71) Shimomura, M.; Ide, K.; Sakata, J.; Tokuyama, H. Unified Divergent Total Synthesis of Discorhabdin B, H, K, and Aleutianamine via the Late-Stage Oxidative N,S-Acetal Formation. *J. Am. Chem. Soc.* **2023**, *145*, 18233–18239.

(72) An, J.; Jackson, R. K. I.; Tuccinardi, J. P.; Wood, J. L. Pyrroloiminoquinone Alkaloids: Total Synthesis of Makaluvamines A and K. Org. Lett. **2023**, *25*, 1868–1871.

(73) Derstine, B. C.; Cook, A. J.; Collings, J. D.; Gair, J.; Saurí, J.; Kwan, E. E.; Burns, N. Z. Total Synthesis of (+)-Discorhabdin V. *Angew. Chem., Int. Ed.* **2024**, 63, No. e202315284.

(74) Lam, C. F. C.; Cadelis, M. M.; Copp, B. R. Exploration of the Influence of Spiro-Dienone Moiety on Biological Activity of the Cytotoxic Marine Alkaloid Discorhabdin P. *Tetrahedron* **2017**, *73*, 4779–4785.

(75) Lam, C. F. C.; Cadelis, M. M.; Copp, B. R. Exploration of the Electrophilic Reactivity of the Cytotoxic Marine Alkaloid Discorhabdin C and Subsequent Discovery of a New Dimeric C-1/N-13-Linked Discorhabdin Natural Product. *Mar. Drugs* **2020**, *18*, 404.

(76) Dolušić, E.; Larrieu, P.; Meinguet, C.; Colette, D.; Rives, A.; Blanc, S.; Ferain, T.; Pilotte, L.; Stroobant, V.; Wouters, J.; Van den Eynde, B.; Masereel, B.; Delfourne, E.; Frédérick, R. Indoleamine 2,3-Dioxygenase Inhibitory Activity of Derivatives of Marine Alkaloid Tsitsikammamine A. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 47–54.

(77) Legentil, L.; Benel, L.; Bertrand, V.; Lesur, B.; Delfourne, E. Synthesis and Antitumor Characterization of Pyrazolic Analogues of the Marine Pyrroloquinoline Alkaloids: Wakayin and Tsitsikammamines. *J. Med. Chem.* **2006**, *49*, 2979–2988.

(78) Rives, A.; Le Calvé, B.; Delaine, T.; Legentil, L.; Kiss, R.; Delfourne, E. Synthesis and Antitumor Evaluation of Analogues of the Marine Pyrroloiminoquinone Tsitsikammamines. *Eur. J. Med. Chem.* **2010**, *45*, 343–351.

(79) Levy, T.; Marchand, L.; Stroobant, V.; Pilotte, L.; Van den Eynde, B.; Rodriguez, F.; Delfourne, E. IDO1 and TDO Inhibitory Evaluation of Analogues of the Marine Pyrroloiminoquinone Alkaloids: Wakayin and Tsitsikammamines. *Bioorg. Med. Chem. Lett.* **2021**, *40*, No. 127910.

(80) Shinkre, B. A.; Raisch, K. P.; Fan, L.; Velu, S. E. Synthesis and Antiproliferative Activity of Benzyl and Phenethyl Analogs of Makaluvamines. *Bioorg. Med. Chem.* **2008**, *16*, 2541–2549.

(81) Shinkre, B. A.; Raisch, K. P.; Fan, L.; Velu, S. E. Analogs of the Marine Alkaloid Makaluvamines: Synthesis, Topoisomerase II Inhibition, and Anticancer Activity. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 2890–2893.

(82) Wang, W.; Rayburn, E. R.; Velu, S. E.; Nadkarni, D. H.; Murugesan, S.; Zhang, R. In Vitro and In Vivo Anticancer Activity of Novel Synthetic Makaluvamine Analogues. *Clin. Cancer Res.* **2009**, *15*, 3511–3518.

(83) Kita, Y.; Egi, M.; Takada, T.; Tohma, H. Development of Novel Reactions Using Hypervalent Iodine(III) Reagents: Total Synthesis of Sulfur-Containing Pyrroloiminoquinone Marine Product, (\pm) -Makaluvamine F. Synthesis **1999**, 1999, 885–897.

(84) For recent examples of the Larock indole synthesis applied in complex molecule synthesis, see this and the following reference: Lin, Y.-C.; Schneider, F.; Eberle, K. J.; Chiodi, D.; Nakamura, H.; Reisberg, S. H.; Chen, J.; Saito, M.; Baran, P. S. Atroposelective Total Synthesis of Darobactin A. J. Am. Chem. Soc. **2022**, 144, 14458–14462.

(85) Nesic, M.; Ryffel, D. B.; Maturano, J.; Shevlin, M.; Pollack, S. R.; Gauthier, D. R., Jr.; Trigo-Mouriño, P.; Zhang, L.-K.; Schultz, D. M.; McCabe Dunn, J. M.; Campeau, L.-C.; Patel, N. R.; Petrone, D. A.; Sarlah, D. Total Synthesis of Darobactin A. *J. Am. Chem. Soc.* **2022**, *144*, 14026–14030.

(86) Larock, R. C.; Yum, E. K. Synthesis of Indoles via Palladium-Catalyzed Heteroannulation of Internal Alkynes. *J. Am. Chem. Soc.* **1991**, *113*, 6689–6690.

(87) Mejía-Oneto, J. M.; Padwa, A. Application of the Rh(II) Cyclization/Cycloaddition Cascade for the Total Synthesis of (\pm) -Aspidophytine. *Org. Lett.* **2006**, *8*, 3275–3278.

(88) Batail, N.; Bendjeriou, A.; Lomberget, T.; Barret, R.; Dufaud, V.; Djakovitch, L. First Heterogeneous Ligand- and Salt-Free Larock Indole Synthesis. *Adv. Synth. Catal.* **2009**, *351*, 2055–2062.

(89) White, J. D.; Yager, K. M.; Yakura, T. Synthetic Studies of the Pyrroloquinoline Nucleus of the Makaluvamine Alkaloids. Synthesis of the Topoisomerase II Inhibitor Makaluvamine D. J. Am. Chem. Soc. **1994**, *116*, 1831–1838.

(90) Kita, Y.; Watanabe, H.; Egi, M.; Saiki, T.; Fukuoka, Y.; Tohma, H. Novel and Efficient Synthesis of Pyrroloiminoquinones Using a Hypervalent Iodine(III) Reagent. J. Chem. Soc. Perkin 1 **1998**, 4, 635–636.

(91) The use of CD_3OD as an NMR solvent resulted in slow substitution of labile methoxy groups in compounds **31**, **41**, **42**, and **43**. In addition, hydrogen deuterium exchange at C6 was observed in many PIQs (see Supporting Information for details). Very recently deuteromethylation of the PIQ framework at C2 has been reported in $(CD_3)_2SO$; see Orfanoudaki, M.; Akee, R. K.; Martínez-Fructuoso, L.; Wang, D.; Kelley, J. A.; Smith, E. A.; Henrich, C. J.; Schnermann, M. J.; O'Keefe, B. R.; Grkovic, T. J. Nat. Prod. **2024**, 87, 415–423.

(92) Kotoku and coworkers very recently adapted Burns's core synthesis strategy to make the damirone (*o*-quinone) core, which was imine N-alkylated. These were converted to makaluvamine J and a handful of unnatural analogues for anticancer selectivity studies: Kiichi, Y.; Fukuoka, K.; Kitano, A.; Ishino, K.; Kotoku, N. Unified Synthesis and Biological Evaluation of Makaluvamine J and Its Analogs. *Molecules* **2024**, *29*, 1389.

(93) We have included comparisons of step counts for the syntheses of these alkaloids in the Supporting Information, as well as summaries

of some of the major approaches used by others to make PIQ compounds.

(94) Singh, P.; Vydyam, P.; Fang, T.; Estrada, K.; Gonzalez, L. M.; Grande, R.; Kumar, M.; Chakravarty, S.; Berry, V.; Ranwez, V.; Carcy, B.; Depoix, D.; Sánchez, S.; Cornillot, E.; Abel, S.; Ciampossin, L.; Lenz, T.; Harb, O.; Sanchez-Flores, A.; Montero, E.; Le Roch, K. G.; Lonardi, S.; Ben Mamoun, C. Insights Into the Evolution, Virulence, and Speciation of *Babesia M01* and *Babesia divergens* Through Multiomics Analyses. *Emerging Microbes Infect.* **2024**, *13*, No. 2386136.

(95) Singh, P.; Lonardi, S.; Liang, Q.; Vydyam, P.; Khabirova, E.; Fang, T.; Gihaz, S.; Thekkiniath, J.; Munshi, M.; Abel, S.; Ciampossin, L.; Batugedara, G.; Gupta, M.; Lu, X. M.; Lenz, T.; Chakravarty, S.; Cornillot, E.; Hu, Y.; Ma, W.; Gonzalez, L. M.; Sánchez, S.; Estrada, K.; Sánchez-Flores, A.; Montero, E.; Harb, O. S.; Le Roch, K. G.; Ben Mamoun, C. *Babesia duncani* Multi-omics Identifies Virulence Factors and Drug Targets. *Nat. Microbiol.* **2023**, *8*, 845–859.

(96) Chahine, Z.; Abel, S.; Hollin, T.; Barnes, G. L.; Chung, J. H.; Daub, M. E.; Renard, I.; Choi, J. Y.; Vydyam, P.; Pal, A.; Alba-Argomaniz, M.; Banks, C. A. S.; Kirkwood, J.; Saraf, A.; Camino, I.; Castaneda, P.; Cuevas, M. C.; De Mercado-Arnanz, J.; Fernandez-Alvaro, E.; Garcia-Perez, A.; Ibarz, N.; Viera-Morilla, S.; Prudhomme, J.; Joyner, C. J.; Bei, A. K.; Florens, L.; Ben Mamoun, C.; Vanderwal, C. D.; Le Roch, K. G. A Kalihinol Analogue Disrupts Apicoplast Function and Vesicular Trafficking in P. falciparum Malaria. *Science* **2024**, 385, eadm7966.