

UCSF

UC San Francisco Previously Published Works

Title

Targeting tauopathies for therapeutic translation

Permalink

<https://escholarship.org/uc/item/0d55s3kd>

Journal

Nature Reviews Neurology, 12(2)

ISSN

1759-4758

Authors

Rojas, Julio C
Boxer, Adam L

Publication Date

2016-02-01

DOI

10.1038/nrneurol.2016.5

Peer reviewed

Targeting tauopathies for therapeutic translation

Julio C. Rojas and Adam L. Boxer

Tau protein abnormalities are key pathogenic features of Alzheimer disease and other neurodegenerative diseases. In 2015, new studies of the less common tauopathies, including progressive supranuclear palsy, chronic traumatic encephalopathy and frontotemporal lobar degeneration, have identified *in vivo* biomarkers and mechanisms that initiate tau pathology.

The microtubule-associated protein tau (encoded by the *MAPT* gene) regulates microtubule structure and function in neurons¹. In 2015, research rapidly advanced the prospect of tau-based therapeutics for neurodegenerative disorders (FIG. 1).

Tau-dependent cytotoxic mechanisms are pervasive in neurodegenerative disorders, occurring not only in Alzheimer disease (AD), but also in conditions such as chronic traumatic encephalopathy (CTE) and parkinsonian disorders¹. These mechanisms involve post-translational modifications of tau, including hyperphosphorylation, cleavage and aggregation.

Tauopathies are diseases in which insoluble deposits of tau protein aggregate in neurons and glia, as shown by autopsy studies of human patients. When animal or cell culture models are inoculated with brain tissue from patients with tau pathology, tau can self-template and spread transcellularly in a prion-like fashion². This finding suggests that the tau protein is an excellent target for therapeutic molecules that reduce its levels, alter its post-translational modification, or block its spread.

The mechanisms that initiate tau pathology are a major unresolved question. AD is the most common tauopathy, but translational studies designed to elucidate mechanisms of tau-dependent neurodegeneration are complicated by the presence of co-pathologies that influence the clinical phenotype. In 2015, Nag *et al.* highlighted this challenge by showing that the TAR DNA binding protein 43 (TDP-43) and hippocampal sclerosis make major contributions to the typical amnesic AD phenotype³.

Autopsy specimens from 636 patients³ showed that hippocampal sclerosis and TDP-43 were important contributors to cognitive impairment. A previous study from the same authors had demonstrated that comorbid synuclein and vascular pathologies are common in AD, and also contribute to the clinical phenotype⁴. Together, these comorbid pathologies could influence clinical assessments, making correlations with tau levels problematic. Initial translational efforts might, therefore, be more successful in disorders with more homogeneous underlying pathology, such as the primary tauopathies, progressive supranuclear palsy (PSP) or autosomal dominant frontotemporal lobar degeneration (FTLD) associated with *MAPT* mutations; in all of these disorders, abnormal tau is the main culprit.

Zhao *et al.*⁵ demonstrated the advantages of focusing on PSP for understanding molecular mechanisms of tau-mediated neurodegeneration. Previous studies have identified strong genetic risk factors for PSP, particularly polymorphisms in *MAPT* itself⁶, but the molecular mechanisms underlying these risk factors were poorly understood. Zhao and colleagues confirmed that a genetic variant that increases the risk of PSP is associated with upregulated appoptosin expression and subsequent high levels of cleaved tau⁵. In cell culture models, cleaved tau had a high tendency to dissociate from microtubules and aggregate⁵. Appoptosin also promoted tau cleavage via caspase-3 activation⁵ and its overexpression promoted synaptic abnormalities and prion-like transcellular spread of cleaved tau⁵. When mice received injections of an appoptosin-expressing virus into the globus pallidus, they developed bradykinesia and tau pathology similar to that seen in patients with PSP⁵. Overexpression of appoptosin was also observed in the brains of patients with AD or FTL⁵, providing support for links between the molecular mechanisms of tau-dependent neurodegeneration in PSP, AD and FTL⁵.

Another molecule that regulates tau structure and function is the peptidyl-prolyl *cis-trans* isomerase NIMA-interacting 1 (Pin1). *PIN1* mutations have previously been implicated as a genetic risk factor for AD. Pin1 exerts a neuroprotective effect by modulating the chirality of tau phosphorylated at residue 231 (p231 tau), maintaining it in a *trans*

Key advances

- Cistauosis is a conformational change in phosphorylated tau, observed in chronic traumatic encephalopathy and Alzheimer disease, that contributes to microtubule dysfunction, prion-like spread of abnormal tau and apoptosis⁷
- In cell culture and animal models, the neuropathological effects of cistauosis can be blocked by a therapeutic monoclonal antibody against the *cis*-phospho-tau epitope⁷
- Overexpression of the mitochondrial carrier protein appoptosin, associated with a strong genetic risk factor for progressive supranuclear palsy (PSP), promotes caspase-mediated tau cleavage, motor dysfunction and tau neuropathology⁵
- In asymptomatic carriers of mutations linked with frontotemporal dementia, cognitive and structural MRI changes are detected years before expected disease onset, suggesting the possibility of clinical prevention trials in *MAPT* mutation carriers⁹
- Elevated tau levels are measurable in the peripheral blood of individuals with a history of repetitive traumatic brain injury, and correlate with the severity of clinical symptoms⁸
- Pure tauopathies, such as PSP, constitute ideal human patient models for translational studies of tau, including clinical trials

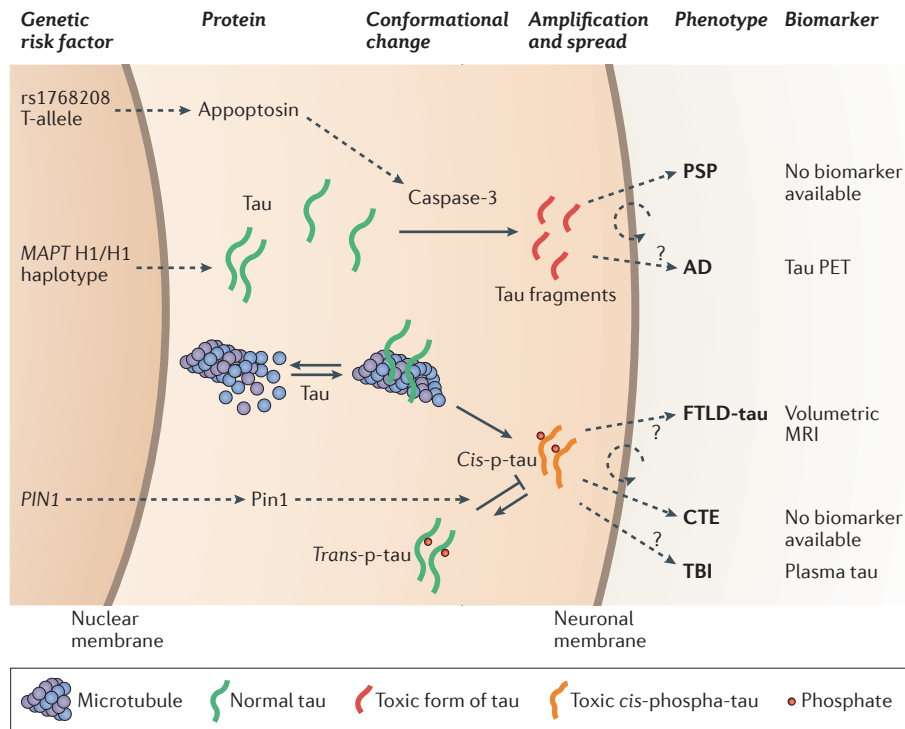


Figure 1 | Advances in tau research in 2015. The rs1768208 polymorphism, linked to progressive supranuclear palsy (PSP), elevates the level of apoptosin and promotes caspase-3-mediated tau cleavage. In PSP and Alzheimer disease (AD), the cleaved tau is thought to spread trans-synaptically, resulting in the disease⁵. In AD, tau-PET signal is elevated¹⁰. One of the functions of tau, encoded by the *MAPT* gene, is to stabilize microtubules. The *MAPT* H1/H1 haplotype is linked to PSP⁶, and MRI can reveal brain atrophy in asymptomatic *MAPT* mutation carriers⁹. *PIN1* polymorphisms promote toxic *cis*-p-tau formation. In animal models, traumatic brain injury (TBI) induces *cis*-p-tau formation that leads to neuronal dysfunction. In human patients with TBI, tau pathology is reflected by elevated blood levels of tau⁸. CTE, chronic traumatic encephalopathy; FTLD, frontotemporal lobar degeneration.

conformation and preventing the formation of toxic *cis* p231 tau (*cis* p-tau), a process that Kondo *et al.* termed ‘cistausis’ (REF. 7). Kondo and co-workers discovered that *cis* p-tau was prominent in autopsy specimens from humans with CTE⁷, in line with the previous reports that have described fulminant tau pathology in CTE. The researchers were also able to induce cistausis in mouse models of single or repetitive traumatic brain injury (TBI), and in cultured neurons by inducing neuronal stress⁷. In these models, cistausis led to disruption of microtubule assembly, impaired axonal transport, and spread of *cis* p-tau to contiguous neurons via a prion-like mechanism with resulting induction of apoptosis. Remarkably, this pathogenic cascade was prevented by monoclonal antibodies to *cis* p-tau, which decreased cellular neurotoxicity, histopathological changes and behavioural deficits in rodent models of TBI⁷. Similar to apoptosin-induced tau cleavage, cistausis occurred before insoluble tau deposition, and within days of the induction of TBI⁷. The findings suggest that monoclonal antibodies that neutralize toxic forms of tau, such as *cis* p-tau, might be effective therapies.

An intriguing study by Olivera *et al.*⁸ suggests that tau could spread into the periphery in individuals with TBI. In military personnel who reported having three or more TBIs, plasma tau concentrations were higher than in individuals with a single documented TBI, who in turn had higher plasma tau concentrations than controls without TBI. Plasma tau concentrations correlated with the severity of post-concussive symptoms. These findings complement the cistausis model and suggest the possibility of monitoring tau-related neurodegeneration with a blood test.

If tau cleavage and cistausis are initiating factors for tau-mediated neurodegeneration, intervention with anti-tau antibodies or other therapies early in the disease course (ideally before the onset of symptoms) is paramount. The Genetic FTD Initiative (GENFI) is a multi-centre natural history study of carriers of autosomal dominant FTLD mutations, including asymptomatic individuals with *MAPT* mutations recruited in the UK, EU and Canada. In the first report of GENFI results⁹, neuropsychological abnormalities could be detected up to 5 years and atrophy at least 10 years prior

to the estimated time of symptom onset. A pattern of sequential atrophy was demonstrated, with insular and temporal cortices affected first, followed by frontal and subcortical areas and, around the time of symptom onset, parietal and cingulate cortices. These results suggest that clinical trials of interventions to prevent the onset of tau-related neurodegeneration in asymptomatic *MAPT* carriers are feasible. In North America, the ARTEL and LEFFTDS projects, which are similar to GENFI, will further enable such studies.

Advances in understanding the pathophysiology of tau-dependent neurodegeneration have sharpened the rationale for new therapies aimed at the tau protein itself. Insights from PSP and CTE have suggested mechanisms that initiate tau pathology, new targets for tau directed therapeutics and potential biomarkers to assess therapeutic effects in humans.

Julio C. Rojas and Adam L. Boxer are at the Clinical Trials Program, Memory and Aging Center, Department of Neurology, University of California, San Francisco, 675 Nelson Rising Lane, Suite 190, MC 1207, San Francisco, California 94143, USA.

Correspondence to A.L.B. adam.boxer@memory.ucsf.edu

doi:10.1038/nrneuro.2016.5
Published online 22 Jan 2016

1. Wang, Y. & Mandelkow, E. Tau in physiology and pathology. *Nat. Rev. Neurosci.* **17**, 22–35 (2015).
2. Goedert, M. Alzheimer’s and Parkinson’s diseases: the prion concept in relation to assembled A β , tau, and α -synuclein. *Science* **349**, 1255–1255 (2015).
3. Nag, S. *et al.* Hippocampal sclerosis and TDP-43 pathology in aging and Alzheimer disease. *Ann. Neurol.* **77**, 942–952 (2015).
4. Schneider, J. A., Arvanitakis, Z., Leurgans, S. E. & Bennett, D. A. The neuropathology of probable Alzheimer disease and mild cognitive impairment. *Ann. Neurol.* **66**, 200–208 (2009).
5. Zhao, Y. *et al.* Apoptosin-mediated caspase cleavage of tau contributes to progressive supranuclear palsy pathogenesis. *Neuron* **87**, 963–975 (2015).
6. Hoglinger, G. U. *et al.* Identification of common variants influencing risk of the tauopathy progressive supranuclear palsy. *Nat. Genet.* **43**, 699–705 (2011).
7. Kondo, A. *et al.* Antibody against early driver of neurodegeneration *cis* P-tau blocks brain injury and tauopathy. *Nature* **523**, 431–436 (2015).
8. Olivera, A. *et al.* Peripheral total tau in military personnel who sustain traumatic brain injuries during deployment. *JAMA Neurol.* **72**, 1109–1116 (2015).
9. Rohrer, J. D. *et al.* Presymptomatic cognitive and neuroanatomical changes in genetic frontotemporal dementia in the Genetic Frontotemporal dementia Initiative (GENFI) study: a cross-sectional analysis. *Lancet Neurol.* **14**, 253–262 (2015).
10. Johnson, K. A. *et al.* Tau positron emission tomographic imaging in aging and early Alzheimer disease. *Ann. Neurol.* <http://dx.doi.org/10.1002/ana.24546> (2015).

Acknowledgements

J.C.R. and A.L.B. are supported by NIH (grants U54NS092089 and R01AG038791) and the Tau Consortium.

Competing interests statement

A.L.B. has received research support from Avid, Biogen, Bristol Myers Squibb, C2N Diagnostics, Cortice Biosciences, Eli Lilly, Forum Pharmaceuticals, Genentech and TauRx. He has served as a consultant for Asceneuron, Ipietron, Ionis (formerly Isis) Pharmaceuticals, Janssen and Merck. He serves on a Data and Safety Monitoring Board for Neurogenetics Pharmaceuticals. He has stock and/or options in Alector and Delos. J.C.R. declares no competing interests.