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Prenatal β -catenin/Brn2/Tbr2 transcriptional cascade regulates adult social and stereotypic behaviors

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Abstract

Social interaction is a fundamental behavior in all animal species, but the developmental timing of the social neural circuit formation and the cellular and molecular mechanisms governing its formation are poorly understood. We generated a mouse model with mutations in two *Dishevelled* genes, *Dvl1* and *Dvl3*, that displays adult social and repetitive behavioral abnormalities associated with transient embryonic brain enlargement during deep layer cortical neuron formation. These phenotypes were mediated by the embryonic expansion of basal neural progenitor cells (NPCs) via

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Conflict of Interest

We declare that there are no competing financial interests in relation to the work described here.

deregulation of a β -catenin/Brn2/Tbr2 transcriptional cascade. Transient pharmacological activation of the canonical Wnt pathway during this period of early corticogenesis rescued the β -catenin/Brn2/Tbr2 transcriptional cascade and the embryonic brain phenotypes. Remarkably, this embryonic treatment prevented adult behavioral deficits and partially rescued abnormal brain structure in *Dvl* mutant mice. Our findings define a mechanism that links fetal brain development and adult behavior, demonstrating a fetal origin for social and repetitive behavior deficits seen in disorders such as autism.

Introduction

Social interaction is a fundamental behavior found in nearly all organisms. Uncovering the developmental mechanisms and genetic pathways governing social behavior is essential for understanding pathophysiological processes that may lead to the development of treatments for neuropsychiatric disorders that display social interaction abnormalities. Social behaviors result from extensive cognitive processes such as obtaining, retrieving, and processing information about the lives, relationships, and mental states of others. The cerebral cortex is responsible for integrating this complex information from multiple brain networks in order to respond to complex social behaviors¹. As social interaction behaviors become more complex, the related regions in the frontal cortex become more active, reflecting the transfer of information processing from automatic to cognitive processes^{1, 2}. Thus, it is likely that abnormal brain development can cause long-term alterations in brain circuitry that may later manifest in behavioral affective disorders in the adult³.

During cerebral cortical development, neuronal progenitor cells (NPCs) in the cerebral cortex undergo temporally and spatially coordinated proliferation, differentiation into neurons, and migration to their final destination, where they will form functional contacts with other neurons to form neuronal circuits (reviewed in⁴). The highly conserved Wnt/wingless signaling cascades regulate many of the processes involved in cortical development, including proliferation, differentiation and migration^{5, 6}. Wnt signals are relayed from the receptors to the downstream effectors by scaffold proteins of the Dvl family. Wnt signals can be transduced through multiple pathways, including canonical Wnt/ β -catenin, planar cell polarity (PCP), and Wnt/ Ca^{2+} cascades. The canonical WNT pathway plays a role in the development and patterning of the brain. Dvl can convert naive ectoderm into neural tissue in *Xenopus*⁷. In addition, WNT signaling via Dvl, posteriorizes neural ectoderm, generating an anteroposterior pattern in the central nervous system⁸. Dvl also appears to be important for later events in neural development, governing the morphology, differentiation and function of neurons in vertebrates⁹⁻¹¹.

Dvl1-null mice (*Dvl1*^{-/-}) exhibit abnormal social interaction behaviors and defects in prepulse inhibition of startle^{12, 13}. No gross pathological abnormalities were previously detected in the brains of adult *Dvl1*^{-/-} mice that could account for their behavioral phenotypes¹². It is possible that *Dvl1*^{-/-} mice have a subtle brain phenotype, due to the known functional redundancy among the *Dvl* genes¹⁴. Another possible explanation is that transient brain abnormalities are present during development that impact adult brain function. Thus, we hypothesized that the study of *Dvl* double mutants may unravel the subtle

developmental pathological abnormalities in the *Dvll1*^{-/-} mice. To test this hypothesis, we generated *Dvll1*^{-/-} *3*^{+/-} mutant mice and uncovered transient fetal brain growth abnormality that was associated with increase in NPCs proliferation and premature differentiation. These embryonic phenotypes were regulated by a novel transcriptional mechanism downstream of canonical Wnt signaling that links fetal brain development with adult social behavior and structural brain defects.

Material and Methods

Mice

All animal care and experiments were performed under protocols approved by UCSF Institutional Animal Care and Use Committee. We generated Wild-type (WT); *Dvl3*^{+/-}; *Dvll1*^{-/-} and *Dvll1*^{-/-} *3*^{+/-} mice on the 129S6 background by mating WT with *Dvl3*^{+/-} and *Dvll1*^{-/-} with *Dvll1*^{-/-}; *3*^{+/-} mice¹⁴. BAT-gal mice on the 129 background¹⁵ were crossed with the *Dvl* mutants to generate the BAT-gal *Dvl* mutant mice. Genotyping for all the mouse strains used were previously described¹⁴. *Brn2* mutant mice¹⁶ were maintained on C57BL/6 mice background as heterozygotes and genotyping for the *Brn2* mutants were previously described¹⁶. For rescue experiments WT and *Dvll1*^{-/-} and *Dvll1*^{-/-} *3*^{+/-} dams were IP injected daily from E9.5 to E14.5 with 4mg/kg CHIR99021, a potent and highly selective inhibitor of glycogen synthase kinase 3¹⁷ (Tocris), which was dissolved according to manufacturer's instructions (4mM DMSO in PBS). The CHIR99021 delivery regime was chosen to maximize effectiveness of the compound taking into consideration diffusion to the target and induction of the normalization of the transcriptional cascade that requires two steps of transcriptional regulation. For BrdU incorporation experiments WT and *Dvll1*^{-/-} and *Dvll1*^{-/-} *3*^{+/-} dams were IP injected 60 min prior embryos collection with 40µg of BrdU per g of mouse weight. The experimental data from each genotype was generated from at least 3 litters. Mice were housed in standard cages and maintained on a 12:12 h light/dark cycle at 22°C. All testing occurred during the light portion of the cycle.

Behavioral analysis

Wild-type, *Dvll1*^{-/-} and *Dvll1*^{-/-} *3*^{+/-} mice (129S6 background) were subjected to a behavioral phenotyping battery¹⁸. Briefly, several behavioral and physical features of the mice were assessed. Next, with the use of a cotton-tipped applicator, each mouse was assessed for several sensorimotor reflexes and its response to an approaching object. Finally, several postural reflexes were also assessed.

A separate cohort of wild-type, *Dvll1*^{-/-} and *Dvll1*^{-/-} *3*^{+/-} mice, and a third cohort of CHIR99021 and mock treated mice were subjected to: nest building, open field, three chamber social approach and marble burying. Mice were 6 weeks of age at the start of the home cage nesting behavioral testing. Mice were transferred to the Gladstone behavioral facility and allowed one week to habituate before behavioral testing began. The open field task was conducted first followed by three-chambered social approach task then finally marble burying task was performed. Time between the two tasks ranged from 2 days to 6 days. Behavioral testing was conducted in dedicated behavioral testing rooms during the

standard light phase, usually between 1000 h and 1500 h. All procedures were approved by the Gladstone Institute and the UCSF Animal Care and Use committee.

Nesting—Nest building activity was assessed in their home cages, 3–4 mice per cage. On the day of the testing a clean cage with equal amounts of bedding and a 5×5cm piece of commercial nesting pad (Ancare, Bellmore, NY) was placed in the cage. Assessments were made at 20, 40, 60 min and 24hours. At each time point the activity of the mice and an estimate of percentage of chewed nestlet were recorded. The depth of the thickest part of the nest was measured.

Open field—General exploratory locomotion in a novel open field environment was assayed as previously described¹⁸. Briefly, mice were transferred to the testing room and acclimated for at least one hour. Individual mice were placed in a standard Photobeam Activity System (PAS) open field apparatus (San Diego Instruments, San Diego, CA, USA). Test chambers consisted of clear Plexiglas sides and floor, approximately 40 × 40 × 30.5 cm. Mice were placed in the center of the open field at the initiation of the testing session. Photocells at standard heights for recording activity were aligned 16 to a side detecting horizontal and vertical activity. Total distance, horizontal activity, vertical activity, and center time were automatically collected using PAS software. Test chambers were cleaned with 30% ethanol between test subjects. At least five minutes between cleaning and the start of the next session was allowed for ethanol evaporation and odor dissipation.

Social approach—Social approach was assayed in a three-chambered apparatus using methods previously described¹⁹. Briefly, the apparatus was a rectangular, three-chambered box made of opaque white polycarbonate (60L × 40W × 25D, in cm). Mice were transferred to the testing room and acclimated for at least one hour. The test session began with a 10 min habituation session in the center chamber only, followed by a 10 min habituation to all three empty chambers. Following the second habituation phase, a clean novel object (wire cup) was placed in one of the side chambers and a novel mouse was placed in an identical wire cup located in the other side chamber. After both stimuli were positioned, the subject mouse was allowed access to all three chambers for 10 min. Trials were video recorded and time spent sniffing the novel object and time spent sniffing the novel mouse were later scored by a trained observer that was blinded to the experimental groups and treatments. Mice that did not explore both chambers with the mouse and the object were excluded from the test. On 129S6 genetic background, 10–30% of the cohorts tested were excluded. This behavior was not associated with any specific genotype or treatment. However, very few (<5%) mice were excluded when they were on the B6 genetic background (WT and Brn2^{+/-} mutants).

Acoustic startle and prepulse inhibition of the acoustic startle—Acoustic startle and prepulse inhibition of the acoustic startle responses were measured using four SR-Lab Systems (San Diego Instruments, San Diego, CA). A test session was begun by placing a subject in the Plexiglas cylinder where it was left undisturbed for 5 min. A test session consisted of seven trial types. One trial type was a 40 ms, 120 dB white noise sound burst presented alone as the startle eliciting stimulus. There were five different acoustic prepulse

plus acoustic startle stimulus trials. The prepulse sound was presented 100 ms before the startle stimulus, and was 20 ms in duration. Prepulse sounds used were 74, 78, 82, 86, or 90 dB in each of the five respective trial types. Finally, there were test trials where no stimulus was presented. These were used to measure baseline movement in the cylinders. Six blocks of the seven trial types were presented in pseudorandom order such that each trial type was presented once within each block of seven trials. The average inter-trial interval was 15 sec (ranged from 10 to 20 sec). The startle response was recorded for 65 ms (measuring the response every 1 ms) beginning at the onset of the startle stimulus. The background white noise level in each chamber was 70 dB. The maximum startle amplitude recorded during the 65-ms sampling window was used as the dependent variable. Percent prepulse inhibition of a startle response was calculated as: $100 - [(startle\ response\ on\ acoustic\ prepulse\ and\ startle\ stimulus\ trials / startle\ response\ alone\ trials) \times 100]$.

A separate startle response curve session was conducted using procedures identical to that described above, except that startle stimulus only trials were presented at dB levels ranging from 0 to 48 dB above the 70 dB baseline. The different startle stimuli were presented in a random order, with five presentations at each level. This procedure provided a gross indication of the hearing and startle.

Marble burying—Marble burying repetitive pattern of behavior was assessed as previously described²⁰. Clean cages (31 × 17 × 14 cm) were filled with 4.5 cm SANI-CHIP bedding, and mice were tested for 20 min. The number of marbles buried (>50% marble covered by bedding material) was recorded.

Histology and immunohistochemistry on whole brains

Embryonic brain samples were dissected without perfusion, and included forebrain through hindbrain. Brain weight was measured and brains were immediately fixed overnight in 4% paraformaldehyde (PFA). Embryo tails were collected for PCR analysis to determine the genotype of the embryo. Samples were transferred to a 30% sucrose solution for 24 hours and then embedded and flash frozen in O.T.C. compound (Tissue-Tek, Torrance, CA). Frozen coronal sections (25 μm) were collected serially every 250 μm, mounted on superfrost slides and stored at -80°C until use. Sections were treated identically and staining was performed on the same day for all genotypes. Cresyl-violet/Nissl staining was performed by standard protocols. The cortical wall and subregions of the cortical wall were measured at 3–4 locations mediolaterally across the neocortex and across the rostral caudal axis every 200 μm, using imagePro software. The Atlas of the Prenatal Mouse Brain²¹ was used as guides for the frontal neocortex, specifically coronal sections #1–4 at E12.5; #1–4 at E14.5; #4–9 at E16.5 and #7–11 for E18.5 were used for all the histological analysis.

For immunohistochemistry, sections were air dried for 1 hr and then rehydrated in PBS followed by incubation with blocking solution (5% donkey serum and 0.5% Triton X-100 in PBS) for 1 hr at room temperature (RT). For Tbr2 staining, antigen retrieval was performed by placing slides in Sodium Citrate Buffer (10mM Sodium Citrate, 0.05% Tween 20, pH 6.0) at 100°C for 30 min and the slides were then incubated in blocking solution. Sections were incubated with primary antibodies diluted in blocking solution for 24h at 4°C, followed

by the appropriate secondary antibody conjugated with Alexa Fluor 488, 568, or 647 (Invitrogen; 1:1000). Primary antibodies used were anti-Sox2 (Santa Cruz, sc17320; 1:200); anti-Tbr2 (Millipore, AB15894; 1:1000); anti-Tbr2 (gift from Dr. R. Hevner, Seattle Children's Research Institute; 1:500); anti-5-bromo-2'-deoxyuridine (BrdU) (Sigma, B8434 1:500); anti-phospho-Histone H3 (Sigma, H0412; 1:500); anti-Ki67 (Dako, M7249; 1:200); anti-Ctip2 (Abcam, ab18465; 1:2000); anti-Cux1 (Santa-cruz, sc-13024; 1:200); anti-NeuN (Millipore, MAB377; 1:500); anti-FoxP2 (Abcam, ab1307; 1:2000); Anti-Brn2 (Santa Cruz, cs6029; 1:400) anti-Brn2 (2R2; 1:2000)²², anti-Tbr1 (Abcam, 31940; 1:200). After immunohistochemistry, slides were mounted with DAPI counter-stain (Invitrogen, P36930).

For Lac-Z enzymatic staining (β -gal), sections were air dried for 1 hr and washed three times in PBS for 5 min. The sections were transferred to β -Gal staining solution [1 mg/ml of 5-bromo-4-chloro-3-indolyl β -D-galactoside (X-Gal), 2 mM MgCl₂, 5 mM potassium ferrocyanide, 5 mM potassium ferricyanide, 0.01% sodium deoxycholate, and 0.02% Nonidet-P40] at 37°C overnight. The samples were washed in distilled water three times, counterstained by Neutral Red, dehydrated in increasing concentrations of ethanol, cleared in xylene, and mounted. Sections were treated identically and the enzymatic reaction was performed on the same day for all genotypes (n=3 for each genotype, derived from at least 3 litters). Staining reaction was timed to ensure uniform exposure and slides from all the mice were randomized.

Neural progenitor cell culture

Neurospheres—Cerebral cortices were isolated from E14.5 mouse embryos, and dissociated by papain-containing dispersion solutions (Sumitomo Bakelite Co. MBX-9901) according to the manufacturer's protocol. At the final step, cells were filtered, added to uncoated tissue culture six-well plates containing N2 supplement with DMEM, 10 ng/ml human recombinant epidermal growth factor, and 10 ng/ml basic fibroblast growth factor (NPC media), then incubated at 37°C in 95% air and 5% CO₂ in a humidified chamber. Half of the media was replaced every other day, allowing cells to grow as floating neurospheres. When culture media began to turn yellow or after 10 days cells were passaged by incubation in 0.0025% Trypsin/EDTA solution (Gibco) for 5 min at 37°C, trypsin inhibitor 1mg/ml (Sigma, T6522) for 5 min at room temperature, collected by centrifugation and re-suspended in fresh NPC media. Doubling time was calculated by counting the number of NPCs at plating and at each passage using a hemocytometer (<http://www.doubling-time.com/compute.php>). Each embryo derived line of NPCs was passaged 3 times. Experimental results were obtained from secondary and tertiary neurospheres. In preparation for immunostaining, neurospheres were fixed in 4% PFA for 16 hours at 4°C, samples were transferred to a 30% sucrose solution for 24 hours at 4°C, embedded and flash frozen in O.T.C. compound (Tissue-Tek, Torrance, CA). Frozen coronal sections (10 μ m) were then cut on a cryostat and mounted on super-frost slides and stored at -80°C until use.

Monolayer adherent NPC cultures—Tertiary neurospheres were dissociated and plated as single cell suspension on poly-ornithine (Sigma, P3655, 10 μ g/ml) and laminin (Sigma, L2020, 5 μ g/ml) coated plates. Cells were grown in DMEM/F-12 Glutamax (Gibco), 1 \times B27 without vitamin A, 1 \times N2 supplement, 10 ng/ml hEGF, and 10 ng/ml bFGF and plated at the

density of 1×10^6 cells/cm². Cells were passaged when they reached ~90% confluence (usually within 3–6 days) by accutase (Millipore) treatment. For LiCl experiments, 5M LiCl was dissolved in water and added to the growth media at a final concentration of 5mM. The cells were checked daily for their growth and proliferation. Dose response curves using (0, 1, 2.5, 5 and 10mM LiCl) demonstrated that consistent and reliable inhibition of growth rate was achieved with 5mM LiCl within 3 days. In preparation for immunostaining, cells were cultured on 12mm cover glass and fixed in 4% PFA then washed with PBS three times and stored at 4°C until use.

Immunofluorescence—Slides or cells were washed once with PBS and then permeabilized with 0.5% Triton X-100 in PBS for 5 minutes. After washing with PBS and blocking with 10% Donkey serum, 0.5% Triton X-100 in PBS (blocking solution), cells were incubated with the primary antibodies diluted in blocking solution for 16 hours at 4°C. After PBS washes, cells were incubated with labeled secondary antibodies for 60 minutes. Nuclei were stained after incubation with 1µg/ml DAPI for 5 minutes. The slides were mounted using prolong-gold (Invitrogen, P36930).

Cell cycle analysis

Monolayer adherent NPCs were harvested, washed with PBS, resuspended in PBS and fixed for 1 hour in 4°C in 100% ethanol (–20°C). Cells were collected, resuspended in PBS containing 100 µg/ml RNase A for 30 min in 37°C, and stained with 1mg/ml propidium iodide solution (10mg/ 1×10^6 cells stained) for 2 h at 4°C. For each sample, 50000 cells were collected in the appropriate gate for analysis on FACSCalibur (BD Biosciences, Mountain View, CA) using FlowJo software (Tree Star, Ashland, OR).

Western Blotting

E14.5 cortex or monolayer cultured NPCs were washed with cold PBS. Protein was extracted with 50mM Tris pH 7.6, 500 mM NaCl, 1 mM EDTA, 1% NP-40, 1% deoxycholate, 0.1% SDS and protease inhibitor cocktail (Roche, # 1 836 153). Lysates were centrifuged for 25 min at 15000 rpm at 4°C, the resulting supernatant was collected and the total protein content of the samples was determined by the BCA Protein Assay Kit (Pierce Biotechnology, #23225). SDS gel electrophoresis was performed utilizing 15µg protein per lane. Gels were transferred to nitrocellulose membranes (BioRad, 9004-70-0). Proteins were detected with anti-Brn2 (2R2 1:2000²²); anti-Tbr2 (Milipore, AB15894; 1:2000); Anti-GAPDH (Acris, ACR001P; 1:10000). All membranes were visualized using ECL (Thermo scientific, 34080) and exposure to autoradiography film (Denville scientific inc. E3018).

Chromatin Immunoprecipitation Assays

ChIP assays²³ were performed on samples from mouse E14.5 cerebral cortex. For each ChIP, 20–30 mg of chromatin was used. For immunoprecipitation, rabbit anti-Brn2 (2R2²²) or rabbit anti β-catenin (Cell signaling, #8480) and normal rabbit IgG as control were used (Santa-cruz, sc-2027). Primer sequences used for qPCR amplification were:

NestinInt2 5′-CAGCCTGAGAATTCCTT-3′ 5′-
GAGAGGGAGGCCGATTCT-3′

mTbr2_905 5'-AGCTGGAGCAAAGGAGCTTA-3' 5'-
CTCTCTGCGGCACAATACAG -3'

mTbr2_95 5'- AAGAAACACCAAACCAGCAA-3' 5'-
TGCTATTGGCTGTTAGAGGTGA-3'

Axin T3 5'- GCATACCTCCCTTCCAGGAC -3' 5'-
GGCGCTTCCAACAAAAACT -3'

Axin T7 5'- CGGAAAAAGTGTGTGTGGAG -3' 5'-
ATCAAGCAACCCAGCTATCC -3'

Brn2 5'- AAGCTCTCTCCCGCTCTCTT -3' 5'- GGGTACAGCTCTGCACCAAT
-3'

qPCR was carried out using SsoFast EvaGreen Supermix (Bio-Rad) and run on the Eppendorf Mastercycler EP Realplex 2 thermal cycler. ChIP-qPCR signals were standardized to the nonspecific rabbit IgG signal. For the identification of putative binding sites for β -catenin and Brn2 binding sites at Brn2 and Tbr2 promoters respectively, we used five different prediction tools (TFBIND (TFBIND.hgc.jp); TESS (www.cbil.upenn.edu); TFSEARCH ver.1.3 (www.cbrj.jp/research/db/TFSEARCH.html), UniPROBE (the_brain.bwh.harvard.edu) and TRANSFAC²⁴). A putative binding site was tested only if two or more tools identified that site. For the identification of putative β -catenin sites, we used predicted TCF binding sites.

shRNA and RT-PCR

HEK293T cells were used to produce lentiviruses expressing GFP (Addgene, 11579, pSicoR) or shRNA viruses against Brn2 (Thermo, TRCN0000075429). HEK293T cells were cotransfected with pLKO.1-Brn2-C5/pSicoR, pCMV- R8.2, and pVSVg vectors using TransIT-LT1 (Mirus, MIR2300) according to the manufacturer's protocol. After 12 hr, the media was replaced with DMEM/F12 GlutaMAXTM (Invitrogen, 10565) and after an additional 24 hr, the viral supernatant was collected, filtered and supplemented with NPC media supplements. Monolayer adherent NPCs were infected with the viral supernatant by centrifugation for 1 hr at 32–35°C at 1000 rpm. Following centrifugation, the viral supernatant was replaced with fresh NPC media. Total cellular RNA was extracted with TRIzol (Invitrogen) and purified on a column using the RNeasy Mini Kit. DNAase treatment was performed on the column. Extracted RNA (1 μ g) samples were reverse transcribed into cDNA according to the manufacturer's protocol for RT-PCR (BioRad, 170-8891). RT-qPCR was performed by using SsoFast EvaGreen Supermix (Biorad) and run on the Eppendorf Mastercycler EP Realplex 2 thermal cycler. The specific primers used for RT-qPCR were:

Brn2_5' 5'- GGTGAGCAGGCTGTAGTGGT -3' 5'-
AGAGCCCAAGGCAGAAAAGT -3'

Brn2_M 5'- CTCACCACCTCCTTCTCCAG -3' 5'-
CCATTTCTCAAATGCCCTA -3'

Brn2_3' 5'- CCATCAGCAITGTCTGTGGTC -3' 5'-
GCCCCAAACAGATTCTACCA -3'

Brn2_3'B 5' - TGAACCAACCTTCCTTTCCA -3' 5'-
TTCATGGAAGTGTGCTCTGG -3'

Tbr2_2 5' - AAACACGGATATCACCCAGC -3' 5'-
GACCTCCAGGGACAATCTGA -3'

B2M_2 5' - CCTGGTCTTTCTGGTGCTTG -3' 5' - TATGTTTCGGCTTCCCATTCT
-3'

Due to the fact that Brn2 is a one exon gene, we designed three primer sets within the mRNA (Brn2_5', Brn2_M, Brn2_3') and as a negative control one primer set for the boundary region of the 3' end (Brn2_3'B).

NPC transfection

pCAG-GFP (Addgene 11150) or pCAG-mBrn2 (Addgene 19711) plasmids were transiently transfected into NPCs (750ng/200000cells one chamber of a four chambered slide) using Lipofectamine LTX+ (Invitrogen) according to the manufacturer's instruction. Transfection was performed in NPC media, which was replaced after 24 hr following transfection. Transfection efficiency was measured by GFP expression and was ~45%. Super Top-Flashx8 (Addgene 12456; 50ng), super Fop-Flashx8 (Addgene 12457; 50ng) and Renilla (10ng) plasmids were transiently transfected into NPCs (40000 cells per well of a 96 well plate) using Lipofectamine LTX+ in NPC media, which was replaced 24 hr following transfection. After 48 hr, cells were lysed in the 96 well plates with passive lysis buffer (Promega). Firefly luciferase and *Renilla* luciferase activity in the lysates were measured on a Synergy 2 microplate reader (BioTek Instruments, Winooski, VT) in replicates of 3, using the Dual-Luciferase reporter assay system (Promega). The ratios for firefly luciferase:*Renilla* luciferase were determined and expressed as relative luciferase activity.

Image Acquisition and Analysis

The immunofluorescent slides were viewed and photographed with 40× or 60× objectives using a Nikon C1si microscope. Analysis of the thickness and/or cell counts of immunohistochemical staining were captured and quantified across the entire image taken by the Image-Pro Plus system (version 5.1; Media Cybernetics).

For Sox2 and Tbr2 staining, the number of DAPI⁺Sox2⁺, DAPI⁺Tbr2⁺ and DAPI⁺Sox2⁺Tbr2⁺ cells was quantified from 2–3 sequential serial coronal sections (250µm apart) and normalized to the number of DAPI⁺ nuclei of the entire cortical wall (ventricle to pia). The data for each developmental time point was normalized to the WT of the indicated time point in order to present all the developmental stages in one plot.

For pHH3 analysis the number of DAPI⁺pHH3⁺ cells were counted across 500µm of the mediolateral plane at the level of the VZ and the SVZ, based on the DAPI staining, which was used to define the morphological compartments.

For cortical plate markers (Ctip2, Cux1, Brn2 and FoxP2) a rectangle of 150µm width was placed on the image in 3 locations and the number and width of the stained neurons was quantified. The data for each developmental time point was normalized to the WT of the

indicated time point in order to present all the developmental stages in one plot or as the raw data in case one developmental stage is presented.

Magnetic resonance imaging

Following embryonic mock or CHIR99021 treatment, 18 week old WT and *Dvll1*^{-/-} mice were anesthetized with ketamine/xylazine and intracardially perfused with 30mL of 0.1M PBS containing 10U/mL heparin (Sigma) and 2mM ProHance (a Gadolinium contrast agent, Bracco Diagnostics) followed by 30mL of 4% paraformaldehyde (PFA) containing 2mM ProHance²⁵. After perfusion, mice were decapitated and the skin, lower jaw, ears, and the cartilaginous nose tip were removed. The brain and remaining skull structures were incubated in 4% PFA + 2mM ProHance overnight at 4°C then transferred to 0.1M PBS containing 2mM ProHance and 0.02% sodium azide for at least 7 days prior to MRI scanning.

Magnetic resonance Imaging—A multi-channel 7.0 Tesla MRI scanner (Agilent Inc., Palo Alto, CA) was used to image the brains within skulls. Sixteen custom-built solenoid coils were used to image the brains in parallel²⁶.

For the anatomical MRI scans a T2- weighted, 3-D fast spin-echo sequence was used, with a cylindrical acquisition of k-space²⁷, and with a TR of 350 ms, and TEs of 12 ms per echo for 6 echoes, field-of-view of 20 × 20 × 25 mm³ and matrix size = 504 × 504 × 630 giving an image with 0.040 mm isotropic voxels. Total imaging time for this sequence is currently ~14 h.

Diffusion Tensor Imaging—For DTI imaging, a 3-D diffusion weighted fast spin-echo sequence was used with an echo train length of 6, with a TR of 270 ms, first TE of 30 ms, and a TE of 10 ms for the remaining 5 echoes, field-of-view 14 × 14 × 25 mm³ and a matrix size of 180 × 180 × 324 yielding an image with 0.078 mm isotropic voxels. Five b=0 s/mm² images and 30 high b-value images (b=1917 s/mm²) in 30 different directions were acquired, using the Jones30 scheme²⁸. Total imaging time for the DTI sequence is currently ~12 h.

To visualize and compare any changes in the mouse brains the anatomical images, as well as the b=0 s/mm² diffusion images (in a separate registration), are linearly (6 parameter followed by a 12 parameter) and nonlinearly registered. All scans are then resampled with the appropriate transform and averaged to create a population atlas representing the average anatomy of the study sample. The result of the registration is to have all scans deformed into exact alignment with each other in an unbiased fashion^{29, 30}.

For the volume measurements, this allows for the analysis of the deformations needed to take each individual mouse's anatomy into this final atlas space, the goal being to model how the deformation fields relate to genotype^{29, 30}. The jacobian determinants of the deformation fields are then calculated as measures of volume at each voxel. Significant volume changes can then be calculated by warping a pre-existing classified MRI atlas onto the population atlas, which allows for the volume of 159 segmented structures encompassing cortical lobes, large white matter structures (i.e. corpus callosum), ventricles, cerebellum,

brain stem, and olfactory bulbs^{31–33} to be assessed in all brains. Further, these measurements can be examined on a voxel-wise basis in order to localize the differences found within regions or across the brain.

For the diffusion measurements this registration allows direct comparisons of intensity differences between groups for all diffusion measures. Fractional Anisotropy, Mean Diffusivity, Axial Diffusivity, and Radial Diffusivity Maps were created using the FSL software package (www.fmrib.ox.ac.uk/fsl) and then resampled using the exact transform that was required to register the $b=0$ s/mm² images. Similar to the volume measurements differences between groups can be assessed in two different ways. 1) Mean values of these parameters can be calculated for specific regions throughout the brain using the exact same atlas used in the volume analysis, and 2) differences in these parameters can be examined on a voxel-wise basis in order to localize the differences found within regions or across the brain.

Statistical Analyses

The majority of the statistical analyses were performed using SPSS version 12. The four groups (WT, *Dvl3*^{+/-}, *Dvl1*^{-/-} and *Dvl1*^{-/-} *3*^{+/-}) were compared by one-way ANOVA or two-way ANOVA for the LiCl and CHIR99021 treatment experiments. The experimental variation was similar between groups and following treatments. Experimental sample size was chosen based on previous data, which was obtained in a similar experimental design using other mouse models. Where appropriate, two-tailed Student's t-test post-hoc analysis was performed with Bonferroni correction for multiple comparisons and the p-values presented are those of the corrected values. The investigator was blinded for the behavioral analysis but was not blinded to the group allocation during the experiment and/or when assessing the outcome of the histological and biochemical experiment. The statistical analysis for the social approach task was a paired t-test that compares the time sniffing the mouse and the object as paired data for each mouse¹⁹.

For the MRI analyses statistical analysis was performed in the R statistical environment (www.rproject.org). Multiple comparisons for the MRI analysis were controlled for using the False Discovery Rate (FDR)³⁴.

Results

Dvl1^{-/-} *3*^{+/-} mice display social deficits

Dvl2 or *Dvl3* single null mutants as well as *Dvl1*^{-/-} *2*^{-/-} and *Dvl2*^{-/-} *3*^{-/-} mutants do not survive past embryonic or neonatal stages, precluding behavioral examination^{14, 35, 36}. However, *Dvl1*^{-/-} *3*^{+/-} mice were viable and fertile. In addition, behavioral testing of *Dvl1*^{-/-} *2*^{+/-} mice did not reveal any additional phenotype compared with *Dvl1*^{-/-} mice. Therefore, we generated cohorts of *Dvl1*^{-/-}, *Dvl1*^{-/-} *3*^{+/-} and wild-type (WT) mice to assess them in a battery of behavioral tasks. Similar to *Dvl1*^{-/-} mutants¹², *Dvl1*^{-/-} *3*^{+/-} mice display social behavior abnormalities, including defects in social barbering, nesting and the three chamber social approach task (Figure 1a–c). These social behavioral tasks we used are qualitative measures of social interaction that determine whether or not a mouse genotype is

social, but they are not able to determine quantitative differences between mouse strains in social behavioral phenotypes¹⁹. However, a marble-burying task, which examines repetitive patterns of behavior, revealed that *Dv11*^{-/-} *3*^{+/-} mice buried more marbles than WT and *Dv11*^{-/-} mice (Figure 1d). No differences between *Dv11*^{-/-}, *Dv11*^{-/-} *3*^{+/-} mutant and WT mice were observed in a behavioral test battery, including normal gross physical appearance and sensorimotor reflexes (Supplementary Table 1). In addition, the total distance traveled and the time spent in the center of the chamber in the open field task was not different between *Dv11*^{-/-}, *Dv11*^{-/-} *3*^{+/-} and WT mice (Supplementary Figure 1a–b). We previously found that *Dv11*^{-/-} mice display inconsistent deficits in prepulse inhibition of startle (PPI,^{12, 13}). The additional reduction in *Dv1* allele in *Dv11*^{-/-} *3*^{+/-} mice did not display reduced PPI and startle response compared to *Dv11*^{-/-} and WT mice (Supplementary Figure 1–d). Thus, we concluded that *Dv11*^{-/-} *3*^{+/-} mice displayed abnormal social behaviors, similar to the *Dv11*^{-/-} mutants, and novel and repetitive behaviors not displayed by *Dv11*^{-/-} mutants, but were otherwise neurologically intact.

***Dv11*^{-/-} *3*^{+/-} mice display transient embryonic brain enlargement**

To determine whether *Dv11* and *Dv13* play a role in brain development, we analyzed the brain size and weight of WT, *Dv13*^{+/-}, *Dv11*^{-/-} and *Dv11*^{-/-} *3*^{+/-} embryos between embryonic days 12.5 (E12.5) and 21 (P0). The brain size (Supplementary Figure 1e) and weight (Figure 1e and Supplementary Table 2) were greater in *Dv11*^{-/-} *3*^{+/-} embryos than in WT or single mutants specifically at E14.5, but not at the other time points examined. Consistent with the transient increase in the overall brain size, *Dv11*^{-/-} *3*^{+/-} embryos displayed a dorsoventral thickening of cerebral cortex wall in all the histological sections (coronal sections #1–4 at E12.5; #1–4 at E14.5; #4–9 at E16.5 and #7–11 for E18.5²¹) of the frontal cortex, with a specific increase in cortical plate (CP) thickness at E14.5 only (Figure 1f–I, Supplementary Figure 1f and Supplementary Table 2).

***Dv11*^{-/-} *3*^{+/-} embryos display early differentiation of apical to basal progenitors**

Expansion of the CP in *Dv11*^{-/-} *3*^{+/-} mutants may be due to altered proliferation or differentiation of cortical neural progenitor cells (NPCs) as a result of reduced β -catenin transcriptional activity. There are two major classes of cortical NPCs in mice: Sox2⁺ apical radial glial NPCs and Tbr2⁺ basal NPCs⁴. We performed immunohistochemistry (IHC) for Sox2 and Tbr2 at various stages of embryonic development (Figure 2a). The area of Sox2 expressing cells was broader, and Tbr2⁺ cells were positioned more basally in *Dv11*^{-/-} *3*^{+/-} embryonic brains at E14.5 and E16.5 than in other genotypes (Figure 2a and Supplementary Figure 2a). Immunohistochemistry (IHC) for phospho-histone-H3, a mitotic marker, revealed that both apical and basal progenitor regions of *Dv11*^{-/-} *3*^{+/-} brains displayed more apical and basal mitotically active cells at E14.5 than WT (Supplementary Figure 2b–c). Combined Ki67 IHC and BrdU incorporation experiments revealed an increased number of Ki67⁺ (cycling cells) and BrdU⁺Ki67⁺ cells in the *Dv1* mutant cortex (Supplementary Figure 2d–e), indicating a significant increase in the proliferation in *Dv11*^{-/-} and *Dv11*^{-/-} *3*^{+/-} mutants. To determine the specific type of NPC that was expanded, we measured the change of each NPC subclass relative to WT. The percentage of Sox2⁺Tbr2⁻ NPCs was reduced in the *Dv11*^{-/-} *3*^{+/-} embryos compared with all other genotypes at E14.5 (Figure 2b). The percentage of Sox2⁺Tbr2⁺ NPCs were unaffected by genetic loss of *Dv1s* (Figure 2c). In

contrast, the percentage of the minor cell population of basally positioned and doubly labeled Sox2⁺Tbr2⁺ NPCs was slightly increased at E13.5 in all *Dvl* mutants compared to WT, reaching statistical significance at E14.5 only (Figure 2d), suggesting that the increase cortical thickening starts at E13.5 and persists to E14.5. The percentage ratios of Sox2⁺:Tbr2⁺:Sox2⁺Tbr2⁺ NPCs are 40%:17%:5% for WT and 29%:22%:20% for *Dvl1*^{-/-}*3*^{+/-} mice, respectively. These findings were recapitulated in adherent NPC monolayer cultures from E14.5 brains (Supplementary Figure 3ab). There was a significant increase of Sox2⁺Tbr2⁺ NPCs in monolayers from *Dvl3*^{+/-} and *Dvl1*^{-/-}*3*^{+/-} compared to WT cells both in neurosphere (Supplementary Figure 3c–e) and monolayer cultures (Supplementary Figure 3f–g). *Dvl1*^{-/-} NPCs were enriched for Sox2⁻Tbr2⁺ cells, but not for Sox2⁺Tbr2⁺ NPCs (Supplementary Figure 3g). The population doubling time was significantly decreased in *Dvl1*^{-/-} and *Dvl1*^{-/-}*3*^{+/-} NPCs, while there was a non-significant trend toward decreased doubling time in *Dvl3*^{+/-} NPCs (Supplementary Figure 3h). One interpretation of these findings is that elevated expression of Tbr2 either in Sox2⁺ or Sox2⁻ NPCs may result in increased NPC proliferation in the *Dvl* mutants.

***Dvl1*^{-/-}*3*^{+/-} embryos display early expansion of deep layer neurons**

To determine the relationship between the early differentiation of apical to basal progenitors and the early and transient thickening of the CP in brains from *Dvl* mutant embryos, we examined cortical lamination by performing IHC for Ctip2 (layer 5 and 6 marker) and Cux1 (layer 2–4 marker). The number of deep layer Ctip2⁺ cells in the CP was increased in *Dvl1*^{-/-}*3*^{+/-} mutants compared to WT at E14.5, then decreased by E18.5 in the mutants (Figure 3a–b). By contrast, superficial neuronal layers, marked by Cux1, were similar in numbers in all genotypes starting at E16.5 when this marker is normally expressed³⁷ and thereafter (Figure 3a and Supplementary Figure 4a). The expanded Ctip2⁺ cells in the CP at E14.5 were indeed neurons, since the NeuN⁺ cell layer was significantly increased in *Dvl1*^{-/-}*3*^{+/-} brains (Figure 3c and Supplementary Figure 4b). Thus, there was a transient expansion of deep layer Ctip2⁺ neurons in *Dvl1*^{-/-}*3*^{+/-} mice at E14.5 that did not persist throughout brain development.

Next, we examined the expression of FoxP2 (layer 6 marker, expressed only after E18.5,³⁸) and Brn2 (marking layers 2–3 and 5). Similar to Ctip2⁺ cells, FoxP2⁺ and Brn2⁺ deep layer neurons were reduced at E18.5 in all the *Dvl* mutants compared to WT embryos (Figure 3d–f). The number of Brn2⁺ neurons in layers 2–3 of *Dvl1*^{-/-}*3*^{+/-} mutants were also reduced at E18.5 (Figure 3e and 3g), even though Cux1⁺ cells were unchanged (Figure 3a and Supplementary Figure 4a). Layer 2–3 Brn2⁻DAPI⁺ nuclei were observed in *Dvl1*^{-/-}*3*^{+/-} mice, suggesting that these cells were not expressing Brn2 (Figure 3e and Supplementary Figure 4c). Thus, Brn2 levels in layer 2–3 neurons were reduced in *Dvl1*^{-/-}*3*^{+/-} brains, while the number of superficial layer neurons remained unchanged.

β-catenin transcriptional activity regulates Brn2 levels in NPCs

Brn2 levels increased in the entire cortical wall of WT embryos from E14.5 to E18.5 and were consistently reduced in *Dvl1*^{-/-}*3*^{+/-} embryos (Figure 3e, 4a and Supplementary Figure 4c). Western blot analysis at E14.5 of cortical lysates revealed reduced expression of Brn2 in *Dvl1*^{-/-} and *Dvl1*^{-/-}*3*^{+/-} mutants (Supplementary Figure 4–e). Consistent with these *in vivo*

observations, the number of Brn2⁺ cells was reduced by 50% in *Dvl1*^{-/-} *3*^{+/-} NPCs compared to WT NPCs (Figure 4b and 4c). These findings suggest that the loss of *Dvl* genes results in the reduction of Brn2 expression in the developing neocortex.

Previous studies demonstrated that β -catenin activates Brn2 expression in cancer cells³⁹. The reduced levels of Brn2 in *Dvl* deficient VZ/SVZ and cultured NPCs led us to hypothesize that β -catenin may directly regulate Brn2 in NPCs. To test this hypothesis, we examined Wnt/ β -catenin transcriptional activity using TOP-flash assays in WT and *Dvl1*^{-/-} *3*^{+/-} NPCs. TOP-flash encodes seven copies of LEF/TCF binding sites linked to firefly luciferase and reflects Wnt/ β -catenin transcriptional activity. TOP flash, β -catenin transcriptional activity was reduced in untreated *Dvl1*^{-/-} *3*^{+/-} NPCs or upon activation of the canonical Wnt signaling with Wnt3A compared with WT NPCs (Figure 4d). To validate the reduced canonical Wnt signaling in vivo, we crossed WT and *Dvl* mutants with the BAT-gal reporter mice¹⁵ that express β -galactosidase (β -gal) in the presence of active β -catenin transcription. β -gal staining was visually reduced in cortical sections of E14.5 *Dvl1*^{-/-} *3*^{+/-} embryos compared to the WT cortex, specifically in the VZ and the CP, although staining was variable (Supplementary Figure 5a) and thus was not quantified. β -gal staining was also reduced in *Dvl1*^{-/-} and *Dvl3*^{+/-} embryos but to a lesser extent (Supplementary Figure 5b). In addition, β -gal staining in WT mice was most pronounced in the frontal cortex at E14.5, but this was diminished by E16.5 and restricted to the cortical hem region (Supplementary Figure 5c). These results suggest that the transient increase in proliferation and early expansion of deep layer neurons in the *Dvl1*^{-/-} *3*^{+/-} embryos at E14.5 was associated with reduced β -catenin transcriptional activity and reduced canonical Wnt signaling in the VZ and CP.

Lithium Chloride (LiCl) treatment, which prevents β -catenin degradation downstream of *Dvl* in the canonical Wnt pathway, elevated β -catenin transcriptional activity to similar levels in the WT and *Dvl1*^{-/-} *3*^{+/-} NPCs (Figure 4d), decreased the proliferation of NPCs from both genotypes (Supplementary Figure 5d), and restored Brn2 to similar levels in *Dvl1*^{-/-} *3*^{+/-} as in WT NPCs (Figure 4e and Supplementary Figure 5e). Double labeling for Brn2 and an activated form of β -catenin (ABC) revealed that the nuclear staining of Brn2 and ABC were spatially and quantitatively correlated (Supplementary Figure 5f). Finally, chromatin immunoprecipitation (ChIP) with a β -catenin antibody demonstrated specific enrichment for β -catenin interaction with a putative TCF binding site on the Brn2 promoter region in WT E14.5 cortex, but not with the known β -catenin interaction sites⁴⁰ on Axin2 (Figure 4f–g), while this enrichment was reduced in *Dvl1*^{-/-} *3*^{+/-} cortex. These findings demonstrate that β -catenin binds to the Brn2 promoter and suggest that down-regulation of β -catenin transcriptional activity in *Dvl* mutants leads to direct reduction of Brn2 levels and increased proliferation in *Dvl1*^{-/-} *3*^{+/-} NPCs.

Brn2 regulates *Tbr2* expression in NPCs

We observed an inverse correlation between *Tbr2* and Brn2 levels in *Dvl1*^{-/-} *3*^{+/-} embryonic brains and NPCs. The VZ/SVZ region in the mutant brains displayed elevated expression of *Tbr2* but reduced expression of Brn2 (Figure 2a, 3e and 3h). The levels of Brn2 were reduced in *Dvl1*^{-/-} *3*^{+/-} compared to WT NPCs by Western blot analysis, consistent with the

IHC results, while LiCl treatment resulted in similar Brn2 levels in both WT and *Dvl1*^{-/-} *3*^{+/-} NPCs (Supplementary Figure 6a and 6b). In contrast, Tbr2 levels were higher in *Dvl1*^{-/-} *3*^{+/-} NPCs than WT, and decreased upon LiCl treatment (Supplementary Figure 6a and 6c). These results suggest that Wnt signaling pathway regulates both Brn2 and Tbr2. We further hypothesized that Brn2 may directly regulate Tbr2 expression. ChIP analysis with a Brn2 antibody²² for two predicted Brn2 binding sites on the Tbr2 promoter (Figure 4h) demonstrated that these sites were significantly enriched for Brn2 binding in the WT but not in the *Dvl1*^{-/-} *3*^{+/-} cortex (Figure 4i). In addition, a known binding site of Brn2 on the nestin promoter⁴¹ was also significantly enriched for Brn2 binding in the WT but not in the *Dvl1*^{-/-} *3*^{+/-} cortex (Figure 4i)

To further support the direct regulation of Tbr2 by Brn2, we knocked down and overexpressed Brn2 in NPCs. We used lentiviral-based shRNA to reduce Brn2 levels in WT NPCs. Tbr2 mRNA and protein levels increased as Brn2 levels decreased over time (Supplementary Figure 6e and 6f), suggesting that Brn2 represses Tbr2 expression. We next transfected CAG-Brn2 into WT and *Dvl1*^{-/-} *3*^{+/-} NPCs, and found that the rapid growth rates of *Dvl1*^{-/-} *3*^{+/-} NPCs was reduced to WT levels after Brn2 overexpression (Figure 4j and 4k). Importantly, the expression pattern of Brn2 and Tbr2 in the Brn2 transfected *Dvl1*^{-/-} *3*^{+/-} NPCs resembled that of WT cells (Figure 4l), providing further support for the direct regulation of Tbr2 by Brn2. Collectively, these findings demonstrate that Brn2 directly represses Tbr2 expression, and suggest that reduced Brn2 levels in *Dvl1*^{-/-} *3*^{+/-} mice resulted in upregulation of Tbr2 and increased proliferation.

Brn2 regulates adult social behavior

If both embryonic brain enlargement and social behavior phenotypes in *Dvl1* and *Dvl3* mutants are mediated by Brn2 downregulation, then the phenotype of *Brn2* null mice²² should be similar to the *Dvl* mutant phenotypes. Indeed, at E14.5, the thickness of the cortical wall was significantly greater in *Brn2*^{-/-} embryos than in WT embryos (Figure Supplementary Figure 6g), due to expansion of the VZ and CP (Supplementary Figure 6h and Supplementary Table 3). There was an increase in Tbr2⁺ and Sox2⁺Tbr2⁺ cells in the VZ (Figure 4m and 4n) and early differentiation of Ctip2⁺ neurons in the CP (Figure 4m), similar to the *Dvl1*^{-/-} *3*^{+/-} embryos. Pools of both Sox2⁻Tbr2⁺ and Sox2⁺Tbr2⁺ cells are increased in *Brn2*^{-/-} embryos compared to WT. It is important to note that Brn2 levels are completely reduced in *Brn2*^{-/-} mice while in *Dvl* mutants Brn2 levels are reduced by about 50% (Figure 4c and Supplementary Figure 6a–b). This might explain the broader effect on the Tbr2⁺ cells in the *Brn2*^{-/-} null embryos than *Dvl1*^{-/-} *3*^{+/-} mutants. *Brn2* null mice die between P0 and P10, which precludes behavior testing of adults. Therefore, we examined *Brn2*^{+/-} embryos and viable adults. We found that *Brn2*^{+/-} embryos displayed VZ and CP expansion that was intermediate to the phenotypes of WT and *Brn2*^{-/-} embryos (Figure 4n and Supplementary Figure 6h). *Brn2*^{+/-} 6-week-old mice displayed reduced nest depth compared with WT mice (Figure 4o), and at 8–10 weeks, *Brn2*^{+/-} mice did not display social preference, unlike their WT littermate controls (Figure 4p). In addition, at 12 weeks of age, *Brn2*^{+/-} mice displayed increased marble burying compared with WT mice (Figure 4q). The similar embryonic progenitor and neuronal expansion phenotypes and adult behavior deficits found in *Brn2* and *Dvl* mutant mice provide strong genetic support for the notion

that the embryonic brain enlargement of $Dvl1^{-/-}3^{+/-}$ mice is mediated by deregulation of the β -catenin/Brn2/Tbr2 transcriptional cascade, and that this prenatal deregulation causes the social behavior deficits observed in the adult mutant mice.

Prenatal canonical Wnt signaling regulates adult social behavior

We predicted that if the transient embryonic brain enlargement in $Dvl1^{-/-}3^{+/-}$ mice, due to reduced prenatal β -catenin transcriptional activity, was the cause of the social behavior deficits observed in the adult $Dvl1^{-/-}$ and $Dvl1^{-/-}3^{+/-}$ mice, then normalization of β -catenin specifically at the time of embryonic brain enlargement would rescue the adult social behavior phenotype. To test this hypothesis, we treated pregnant dams between E9.5–E14.5 with daily IP injections of 4 mg/kg CHIR99021 (Supplementary Figure 7a), a specific GSK3 β inhibitor that activates intracellular canonical Wnt signaling⁴². At the end of the treatment (E14.5), we examined embryonic brain enlargement and the associated cellular phenotypes. CHIR99021 treatment between E9.5–E14.5 normalized brain weight (Supplementary Figure 7b), cerebral cortical wall thickness (Figure 5a and 5b), and CP thickness of $Dvl1^{-/-}3^{+/-}$ E14.5 embryonic brains (Figure 5a and Figure 5d). In addition, CHIR99021 treatment between E9.5–E14.5 normalized the percentages of the doubly labeled Sox2⁺Tbr2⁺ NPCs in the E14.5 $Dvl1^{-/-}$ and $Dvl1^{-/-}3^{+/-}$ brains (Figure 5e and 5h), as well as the number and width of Ctip2⁺ and Ctip2⁺Tbr1⁺ neurons, which mark the deep layer neurons in the CP (Figure 5i–l). Importantly, CHIR99021 treatment normalized Brn2 and β -catenin levels in $Dvl1^{-/-}$ and $Dvl1^{-/-}3^{+/-}$ embryonic brains and increased Brn2 levels in $Dvl1^{-/-}$ brains (Supplementary Figure 8a–d). These findings demonstrate that CHIR99021 treatment rescued the brain enlargement and all of the associated cellular phenotypes in $Dvl1^{-/-}$ and $Dvl1^{-/-}3^{+/-}$ embryos. CHIR99021 treatment had no effect on VZ thickness in any of the genotypes, as expected (Figure 5a and 5c). However, in WT mice, CHIR99021 treatment resulted in an increase in the percentages of the Sox2⁺Tbr2⁻ cells (Figure 5e and 5f) and a decrease in the percentages of the Sox2⁻Tbr2⁺ cells (Figure 5e and 5g), providing additional support for the role of canonical Wnt signaling in regulating Tbr2 expression.

To test whether deficits in social behavior of adult $Dvl1^{-/-}$ and $Dvl1^{-/-}3^{+/-}$ mutant mice originated from the developmental defects, we produced a cohort of adult mice treated *in utero* with CHIR99021 or mock treated (4mM DMSO in PBS) between E9.5–14.5 in an identical regimen that corrected the embryonic brain phenotypes. These adult mice were tested in the social behavior, marble burying, and open field tasks. At 6 weeks of age, mock treated $Dvl1^{-/-}$ and $Dvl1^{-/-}3^{+/-}$ mutant mice displayed reduced nest depths compared with WT mice, as we had found previously, while the treatment with CHIR99021 during embryonic development normalized this behavior in $Dvl1^{-/-}$ and $Dvl1^{-/-}3^{+/-}$ mutant mice (Figure 6a and Supplementary Figure 8e). At 8–10 weeks, WT mock treated males displayed social preference, and $Dvl1^{-/-}$ mock treated males did not, as expected (Figure 6b). By contrast, CHIR99021 treatment of $Dvl1^{-/-}$ mice rescued the lack of social preference of the $Dvl1^{-/-}$ mock treated mice (Figure 6b). Surprisingly, WT male mice treated with CHIR99021 as embryos did not display social preference, perhaps due to the effect of CHIR99021 treatment on Sox2⁺ (Figure 5f) and Tbr2⁺ (Figure 5g) cells. Females from the 129S6 strain do not display social preference in the three chamber social approach task⁴³.

Consistent with this, we found that mock and CHIR99021 treated females displayed no social preference regardless of genotype (Supplementary Figure 8f), and thus females were not informative in this task. In addition, out of the 7 litters that were treated with CHIR99021, no adult male $Dvll^{-/-} \mathcal{I}^{+/-}$ mice were found ($Dvll^{-/-}:Dvll^{-/-} \mathcal{I}^{+/-}$ male ratios were 14:0 and female ratios were 5:8), as the CHIR99021 treated $Dvll^{-/-} \mathcal{I}^{+/-}$ neonates were born but died postnatally of unknown causes. Thus, $Dvll^{-/-} \mathcal{I}^{+/-}$ males could not be tested in the social approach task. At 12 weeks of age, mock treated $Dvll^{-/-} \mathcal{I}^{+/-}$ male and female mice displayed increased marble burying compared with WT mice, as we had found previously, while the treatment with CHIR99021 during embryonic development normalized this behavior in $Dvll^{-/-} \mathcal{I}^{+/-}$ female mice (Figure 6c). WT male mice treated with CHIR99021 as embryos displayed increased marble burying compared with mock treated WT mice, providing additional support for the role of canonical Wnt signaling in regulating stereotypic behavior. There were no significant effects of genotype or treatment in the open field task (Figure 6d and 6e). Thus, CHIR99021 treatment displayed a degree of specificity for the development of social behavior and stereotypic behavior.

These findings demonstrate that the rescue of the canonical Wnt pathway in $Dvll^{-/-}$ and $Dvll^{-/-} \mathcal{I}^{+/-}$ mutant embryos specifically during E9.5–14.5 rescued the embryonic cellular and molecular phenotypes of the developing cortex, as well as the adult social behavior deficits and the increase in stereotypic behavior. These data indicate that precise tuning of the Wnt canonical pathway during this critical developmental period is essential for the establishment of normal social and stereotypic behavior in the adult.

Prenatal canonical Wnt signaling rescues *Dvl* mutant adult brain architecture defects

Our previous work was unable to detect any gross histopathological abnormalities in $Dvll^{-/-}$ mice¹². To test whether adult $Dvll^{-/-}$ mice (18 weeks old) present with brain morphological and structural deficits that may have originated from developmental defects, we produced a third cohort of adult mice treated *in utero* with CHIR99021 or mock treated (4mM DMSO in PBS) between E9.5–14.5 that corrected the embryonic and behavioral deficits of the *Dvl* mutants. Using a T2-weighted magnetic resonance (MR) imaging sequence and advanced registration techniques, we performed a volumetric analysis of the entire brain and 159 independent regions throughout the brain including but not limited to the several cortical delineations, large white matter structures (i.e. corpus callosum), ventricles, cerebellar structures, brain stem, and olfactory bulbs^{31–33}. $Dvll^{-/-}$ mock treated brains were smaller than WT brains (94% of WT volume), and that effect did not change following embryonic CHIR treatment in either WT or $Dvll^{-/-}$ mice (Figure 6f). Thus, we assessed the neuroanatomy of the independent regions as relative volume to account for the brain volume difference between groups. We found that 21 out of the 159 independent regions displayed significantly different relative volumes between $Dvll^{-/-}$ mock compared to WT mock brains (FDR<0.05), and these regions were often increased (Figure 6g and Supplementary Table 4). A bilateral increase in relative volume of the dentate gyrus and the dorsal raphe nuclei were of particular interest as they were also seen as some of the most affected regions in a previous report that examined 26 mouse models relevant to autism⁴⁴. Diffusion Tensor Imaging (DTI) and subsequent analyses revealed that 68 out of the 159 independent regions

were significantly different ($FDR < 0.10$) in fractional anisotropy (FA), which was predominantly a decreased FA (Figure 6h and Supplementary Table 5).

CHIR99021 embryonic treatment did not affect either T2 or DTI imaging in WT mice. However, volumetric analysis revealed significant differences between mock and CHIR99021 embryonically treated *Dvl1*^{-/-} mice. The olfactory bulbs and the lateral olfactory tract displayed a significant interaction between genotype and treatment, the most substantial of which was also found voxel-wise in the olfactory bulb (Supplementary Figure 9a–c). The second area where an interaction was found, also voxel-wise, was the tip of the cerebral peduncle seen in both an axial and coronal slice (Supplementary Figure 9b). We also examined the proportion of the voxels that displayed evidence of an improvement with treatment. We found that 74.7% and 81.2% of the voxels that were significantly different by volumetric analysis (Figure 6i) and FA analysis (Supplementary Figure 9a, Supplementary Table 4 and 5), respectively, in the *Dvl1*^{-/-} mock vs. the WT mock trended towards recovery in the *Dvl1*^{-/-} CHIR treated mice. Lastly, analysis of the relative volume of *Dvl1*^{-/-} CHIR99021 compared to WT mock brains revealed that 6 of the 21 regions that were significantly different in *Dvl1*^{-/-} mock vs. WT brains were not significantly different in the *Dvl1*^{-/-} CHIR99021 treated brains, demonstrating a regional-specific normalization of brain structure. These regions were the fundus of the striatum, olfactory bulb, copula white matter frontal association cortex, primary motor cortex and the primary somatosensory cortex (Supplementary Table 4 and 6). Thus, CHIR99021 treatment displayed a degree of specificity for the development of specific brain structures, including several regions of the cerebral cortex.

Discussion

A β -catenin/Brn2/Tbr2 pathway regulates fetal brain growth and development, as well as adult social and repetitive behavior

Social interaction is a fundamental behavior found in nearly all organisms, but the molecular and cellular mechanisms governing this behavior are poorly understood. We generated *Dvl1*^{-/-} *3*^{+/-} mice, which allowed us to identify a β -catenin/Brn2/Tbr2 transcriptional pathway that regulates embryonic brain size and deep layer neurogenesis that is essential for normal social and stereotypic behaviors. We propose a model in which Dvl1 and Dvl3 facilitate canonical Wnt signaling via β -catenin to directly promote expression of the Brn2 transcription factor (Supplementary Figure 10a and 10b). Brn2 in turn acts as a direct repressor on the Tbr2 promoter to inhibit expression of this transcription factor (Supplementary Figure 10a and 10b) and the expansion of basal progenitors during deep layer neurogenesis (Supplementary Figure 10c and 10d).

Multiple lines of evidence support our proposed mechanism. The genetic loss of *Dvl* genes leads to an early enlargement of the embryonic brain, which is the result of a transient over-proliferation of neural progenitors in the cerebral cortex and premature differentiation of deep layer cortical neurons (Supplementary Figure 10c and 10d). During cortical development in WT mice (Supplementary Figure 10a), activation of the canonical Wnt pathway in the VZ results in the activation of β -catenin transcriptional activity, increased Brn2 expression, and suppression of Tbr2 in neural progenitors to promote accurate

temporal proliferation and neurogenesis. In *Dvl* mutant mice (Supplementary Figure 10b), there is less activation of the canonical Wnt pathway, reduced β -catenin transcriptional activity in the VZ, decreased *Brn2* expression, and premature expression of *Tbr2*. *In vitro*, both β -catenin transcriptional activity and *Brn2* levels were decreased while *Tbr2* levels were increased in cortical NPCs from *Dvl1^{-/-}3^{+/-}* versus WT embryos, associated with increased proliferation. Activation of β -catenin transcriptional activity in *Dvl1^{-/-}3^{+/-}* NPCs by LiCl reversed both the decrease in *Brn2* levels and the increase in *Tbr2* levels, resulting in reduced proliferation of the *Dvl* mutant NPCs. Knock-down of *Brn2* in WT NPCs resulted in increased *Tbr2*, while forced expression of *Brn2* rescued the proliferative phenotype of *Dvl1^{-/-}3^{+/-}* NPCs. *Brn2* null mice phenocopied the embryonic cortical phenotype of the *Dvl* mutant mice, and *Brn2* heterozygotes display similar adult behavior deficits (Figure 4m–q). Finally, the activation of the canonical Wnt pathway by CHIR99021 specifically during this critical embryonic period (E9.5–14.5) rescued the embryonic dysregulation of the β -catenin/*Brn2*/*Tbr2* transcriptional cascade in the developing brain, the transient over-proliferation of neural progenitors and the premature differentiation of deep layer cortical neurons. Importantly, this treatment also rescued the social and stereotypic behavioral deficits that were observed in adult *Dvl* mutant mice. Finally, restoration of the canonical Wnt pathway in *Dvl1^{-/-}* mutant embryos specifically during E9.5–14.5 resulted in the recovery of specific adult brain morphological and structural deficits, including areas of the cerebral cortex. The ability to reverse an adult phenotype by transient embryonic normalization of the Wnt pathway provides strong evidence for our model.

The β -catenin/*Brn2*/*Tbr2* transcriptional cascade and its role in the regulation of NPC number and brain size during development have not been previously described, although features of this cascade are supported by previous studies. Early in neurogenesis, NPCs respond to Wnt stimulation by proliferating symmetrically, whereas later in development Wnts also induce neuronal differentiation by promoting terminal neurogenesis of intermediate progenitors (IPs)^{6, 45, 46}. These studies highlight the complex temporal and context-dependent control of Wnt signaling in distinct progenitor types during brain development. It is likely that some of the temporal and context-dependent effects of β -catenin on NPCs are mediated by transcriptional regulation of *Brn2*. In melanoma cells, β -catenin directly regulates *Brn2* expression and controls proliferation⁴⁷, which implies that the regulation of *Brn2* expression by canonical Wnt signaling is not unique to NPCs. In the brain, *Brn2* is a positive regulator of genes that are expressed in apical NPCs such as *Nestin*⁴¹ and *Sox2*⁴⁸, while *Brn2* co-regulates neurogenesis with *Mash-1*⁴⁹. *Brn2* is one of three factors necessary for the direct reprogramming of fibroblasts to induced neurons by contributing long-term transcription of selected genes⁵⁰. Similar to *Dvl* mutants, initial examination of *Brn2* knockouts failed to reveal significant cortical brain abnormalities¹⁶, but our more detailed examination revealed that *Brn2* mutant mice display similar embryonic brain enlargement and social behavior phenotypes as *Dvl* mutants, strongly supporting the linkage of these phenotypes and the molecular pathway. *Dvl* and *Brn2* mutant mice are maintained on 129S6 and C57Bl6/J inbred genetic backgrounds, demonstrating that the embryonic brain development and the behavioral phenotypes are strain independent. *Tbr2* null mice are severely affected and cortical layers are reduced in thickness, confirming that *Tbr2*⁺ IPs are necessary to expand the pool of the neurons of each cortical layer⁵¹. It was

previously shown that sustained β -catenin activity in the developing cortex delays the onset of Tbr2 expression⁵², which is consistent with our finding that reduction of β -catenin activity due to the loss of *Dvl* genes resulted in an increase in Tbr2 expression, while CHIR99021 treatment of WT embryos resulted in reduce Tbr2 expression. In contrast to our findings, it was recently reported that Brn2 promotes the expression of Tbr2⁵³. This study employed the N2a cell line and the Engrailed dominant negative system to eliminate Brn2 function, which may result in off target effects on closely related POU-domain transcription factors such Brn1 and Oct6. Our studies used *Brn2* genetic mutants as well as Brn2 knock-down or overexpression in primary cells with findings consistent with our model.

The initial expansion of deep layer neurons (Figure 3a–b) in *Dvll1^{-/-}3^{+/-}* mice was due to the deregulation of Brn2 levels, but the mechanism for the rapid decrease in the deep layer neurons at about E16.5 is not clear. We were unable to find evidence for several potential mechanisms that could account for the decrease of deep layer neurons. Premature NPCs depletion is unlikely to account for the deep layer neuronal decrease, since the later born superficial layer neurons are unaffected in the *Dvll1^{-/-}3^{+/-}* mice. Apoptosis of the increased numbers of deep layer neurons could potentially account for the deep layer neuronal decrease, but the number of apoptotic nuclei was not different between *Dvll1^{-/-}3^{+/-}* and WT mice at E16.5 (Supplementary Figure 11a). Interneurons invade the CP after E15.5⁵⁴, however, the number of GABA positive neurons in the cortex of E18.5 *Dvll1^{-/-}3^{+/-}* and WT mice are similar (Supplementary Figure 11b). Lastly, it was reported that microglia (Iba1⁺ cells) could limit the production of cortical neurons by phagocytizing neural precursor cells⁵⁵. However, the number of Iba1⁺Tbr2⁺ cells was not different between *Dvll1^{-/-}3^{+/-}* and WT mice at E16.5 (Supplementary Figure 11c). Thus, we speculate that the rate of neurogenesis at early stages of cortical development in *Dvll1^{-/-}3^{+/-}* mice is rapid due to β -catenin transcriptional activity that drives the induction of basal identity of the progenitor pool, and then decreases at E16.5 due to the lack of β -catenin transcriptional activity (Supplementary Figure 5) and Brn2, a proneuronal transcription factor (Supplementary Figure 10e).

Redundancy of *Dvl* genes in development and social behavior

Our study was aimed at unraveling the molecular mechanism that controls abnormal social interaction behaviors in *Dvll1* mutant mice. Adult *Dvll1^{-/-}* mice display social interaction abnormalities, but detectable brain gross pathological abnormalities were not found that could account for their behavioral phenotypes¹². One reason for the lack of detectable abnormalities in adult *Dvll1^{-/-}* mutants is that the relevant phenotypes occurred during embryonic development, since *Dvll1^{-/-}3^{+/-}* embryos displayed brain enlargement mediated by expansion of basal progenitors during the production of deep layer neurons (E14.5). In addition, the known functional redundancy of *Dvl2* and *Dvl3* resulted in a subtle, difficult to detect brain phenotype in *Dvll1^{-/-}* mice that was revealed only by after examination of *Dvll1^{-/-}3^{+/-}* mutants. Once embryonic brain defects were uncovered in *Dvll1^{-/-}3^{+/-}* mutants, some of these phenotypes were also found in *Dvll1^{-/-}* or *Dvl3^{+/-}* mutants. Functional redundancy among the *Dvl* genes results from their overlapping expression patterns and their high degree of conservation, and this is highlighted by their ability when overexpressed to cross-rescue phenotypes resulting from loss-of-function of other *Dvl* genes 14. Some of

the embryonic brain phenotypes (Sox2⁺Tbr2⁺ cells at E14.5 and reduced Brn2 in layer 5 at E18.5) were found in all *Dvl* mutants, other phenotypes (cortical thickness) were present only in *Dvl1*^{-/-} and *Dvl1*^{-/3^{+/+}} mutants, while all phenotypes were found in *Dvl1*^{-/3^{+/+}} mutants. In addition, while *Dvl1*^{-/-} and *Dvl1*^{-/3^{+/+}} mutants share the social behavior deficits, *Dvl1*^{-/3^{+/+}} mutants display additional behavioral deficits, such as marble burying (Figure 1d). These observations are consistent with complex functional redundancy of *Dvl1* and *Dvl3*, affecting the penetrance and expressivity of an individual embryonic brain or adult social behavior phenotype in *Dvl3*^{+/+}, *Dvl1*^{-/-} and *Dvl1*^{-/3^{+/+}} mutants, as is the complete penetrance of all phenotypes in the *Dvl1*^{-/3^{+/+}} mutants. It does not appear that *Dvl2* plays an important role in social behavior, since preliminary studies of *Dvl2*^{+/+} and *Dvl1*^{+/+}2^{+/+} mutants did not reveal social interaction abnormalities (J.L., A.W.B., unpublished). This suggests that functional redundancy among *Dvl* genes is incomplete, which is supported by *in vitro* analysis of *Dvl* genes⁵⁶.

Fetal development and autism

The embryonic brain enlargement and developmental defects displayed by *Dvl* mutant mice appear to be responsible for the adult social and stereotypic behavioral deficits displayed by *Dvl* mutants, supporting an embryonic origin for the social interaction phenotypes in the mouse. The embryonic brain phenotypes are mediated by expansion of basal progenitors during the production of deep layer neurons (E14.5) due to the dysregulation of a β -catenin/Brn2/Tbr2 transcriptional cascade. Premature neuronal differentiation of deep layer neurons was observed in *Dvl* mutant mice without affecting superficial layers (Figure 3). The initial enlargement was due to the deregulation of Brn2 levels. We speculate that the rate of neurogenesis at early stages of cortical development in *Dvl1* mutant mice is rapid due to induction of basal identity of the progenitor pool, and then decreases due to the lack of β -catenin transcriptional activity (Supplementary Figure 5) and Brn2, a proneuronal transcription factor (Supplementary Figure 10e). The linkage of embryonic brain phenotypes and adult social interaction deficits in both *Brn2* and *Dvl1*^{-/-}3^{+/+} mutants further supports the embryonic origin of the social behavioral deficits. Finally, the successful rescue of the embryonic brain defects in the *Dvl* mutants by the specific and restricted activation of the canonical Wnt pathway between E9.5 and E14.5 also rescued the adult social behavior deficits that were observed in *Dvl1*^{-/-} and *Dvl1*^{-/3^{+/+}} mutants. Surprisingly, CHIR99021 treatment resulted in lack of sociability and increase of stereotypic behavior in WT mice (Figure 6b and 6c), suggesting that both activation and inhibition of β -catenin canonical pathway during deep layer neurogenesis are associated with these behavioral abnormalities. Such a “bell-shaped” dose response relationship in canonical Wnt signaling was recently proposed for autism based on genetic studies in humans⁵⁷, as well as in syndromic mouse models of autism, such as *Tsc2*(+/-) and *Fmr1*(-/y) mice⁵⁸ and MECP2 gain- and loss-of-function mice⁵⁹. This provides additional support for a bell-shaped dose response relationship in ASD.

The phenotypes and molecular mechanisms responsible for the behavioral defects in the *Dvl* mutants may model some aspects of autism. *Dvl1*^{-/-}3^{+/+} mutants displays both social deficits and repetitive patterns of behavior, similar to autism. Postmortem brain gene expression^{60, 61} and neuronal counting in young autistic cases⁶² identified developmental

disturbances in molecular and genetic mechanisms that govern proliferation, cell cycle regulation, and apoptosis, and inferred that these events occurred prior to birth. Two recent independent studies of human ASD genes identified spatial and temporal convergence of these genes in human mid-fetal cortical development^{63, 64}. Interestingly, one study identified coexpression of ASD genes with deep layer 5/6 projection neurons^{63, 64}, while the second study identified superficial layer glutamatergic inter- and intra-cortical neurons^{63, 64}. In addition, two Wnt associated genes (Wnt10B and CTTNBP2) and Ctip2 (Bcl11b) were present in their coexpression networks with genes important for the development of autism⁶⁴, supporting a role for prenatal canonical Wnt signaling in the development of deep layer neurons in humans.

Embryonic neurogenesis has been identified as a critical temporal period in other environmental and genetic mouse models of autism. Several known environmental exposure models, including maternal infection and maternal valproate administration, have emphasized the importance of this embryonic period^{65, 66}. Knockdown of Tuberous sclerosis 1 (TSC1) from the thalamic circuitry at E12.5 alters neuronal physiology, resulting in both seizures and compulsive grooming in adult mice³. By contrast, only a subset of these phenotypes occurs when thalamic Tsc1 is deleted at a later embryonic stage.

Structural histopathological abnormalities in the adult *Dv11*^{-/-} mice that could account for their behavioral phenotypes have not been identified¹². By employing state-of-the-art T2 and DTI MR imaging, we found that the whole brain volume of *Dv11*^{-/-} mice was reduced (Figure 6f) and that multiple brain regions of the *Dv1* mutant brains displayed abnormal size (Figure 6g–i and Supplementary Table 4–6). A previous report investigated 26 different mouse lines with relevance to autism, and clustered these lines into three large groups that differed in both directional volumetric differences as well as localization of anatomical phenotypes⁴⁴. Based on this study, the *Dv11*^{-/-} mouse brain MRI phenotype corresponds to group 1, and closely and specifically resembles the *Fmr1* (FVB) and *Nrxn1a* models, with less resemblance to the *Shank3* and *En2* models. The common phenotype of mouse mutant within this group is increased volumes in large white matter structures and frontal and parieto-temporal lobes as well as decreased volumes in the cerebellar cortex. Consistent with these mutants, the frontal (FDR=0.15) and parieto-temporal lobes (FDR=0.14) were increased in relative volume in the *Dv11*^{-/-} mice.

T2 and DTI MR imaging also revealed that of the 21 brain regions that were abnormal in *Dv11*^{-/-} mutants in the adult brain, 6 were rescued by CHIR99021 treatment: the fundus of the striatum; olfactory bulb; copula white matter; frontal association cortex; primary motor cortex; and the primary somatosensory cortex (Supplementary Table 4 and 6). These regions were previously reported to be associated with the neurological manifestation in autistic patients. The striatum was previously implicated in the stereotypic pattern of behavior⁶⁷. The abnormalities in the olfactory bulb could be associated with the heightened olfaction that was reported in autistic patients⁶⁸. We have not behaviorally tested olfaction in *Dv1* mutant mice and future studies will be needed to address this in detail, both molecularly and behaviorally. The motor cortex and the copula (part of the cerebellum) structural abnormalities could participate in the motor impairments found in 80% of patients with autism⁶⁹, as well as the reduced somatosensory response and connectivity in the

somatosensory cortex found in autism^{70, 71}. Lastly, the increased volume of the frontal association cortex (FAC) in *Dvl1*^{-/-} mock treated mice was almost completely recovered by *in utero* CHIR99021 treatment (Figure 6i and Supplementary Figure 9c). Interestingly, the FAC was also increased, though not significantly, in WT mice that were *in utero* treated with CHIR99021 that also displayed abnormal behavior. The specific involvement of the FAC in the social behavioral deficits seen in *Dvl1*^{-/-} mock treated and WT CHIR99021 treated mice, provides further support for an important role of the embryonic canonical β -catenin signaling on the development of specific brain structures central for the neurological/behavioral abnormalities in autism.

In summary, our studies of the *Dvl* and *Brn2* mutant mice support a model in which abnormalities in embryonic brain development are responsible for abnormal adult brain structures and behavioral abnormalities. We identified a critical period during embryonic brain development for the establishment of normal social behavior in mice, when deep layer projection neurons are produced, which is regulated by a β -catenin/*Brn2*/*Tbr2* transcriptional cascade. In addition, we link this critical period with abnormalities in specific adult brain structures. These findings are consistent with a model that autism may be caused by increased production of neurons during cortical prenatal development⁶², specifically during the development of deep layer glutamatergic projection neurons^{63, 64}. Further studies will be required to determine the consequences of the abnormal development of deep layer neurons to the adult brain circuitry and function and possible therapeutic interventions.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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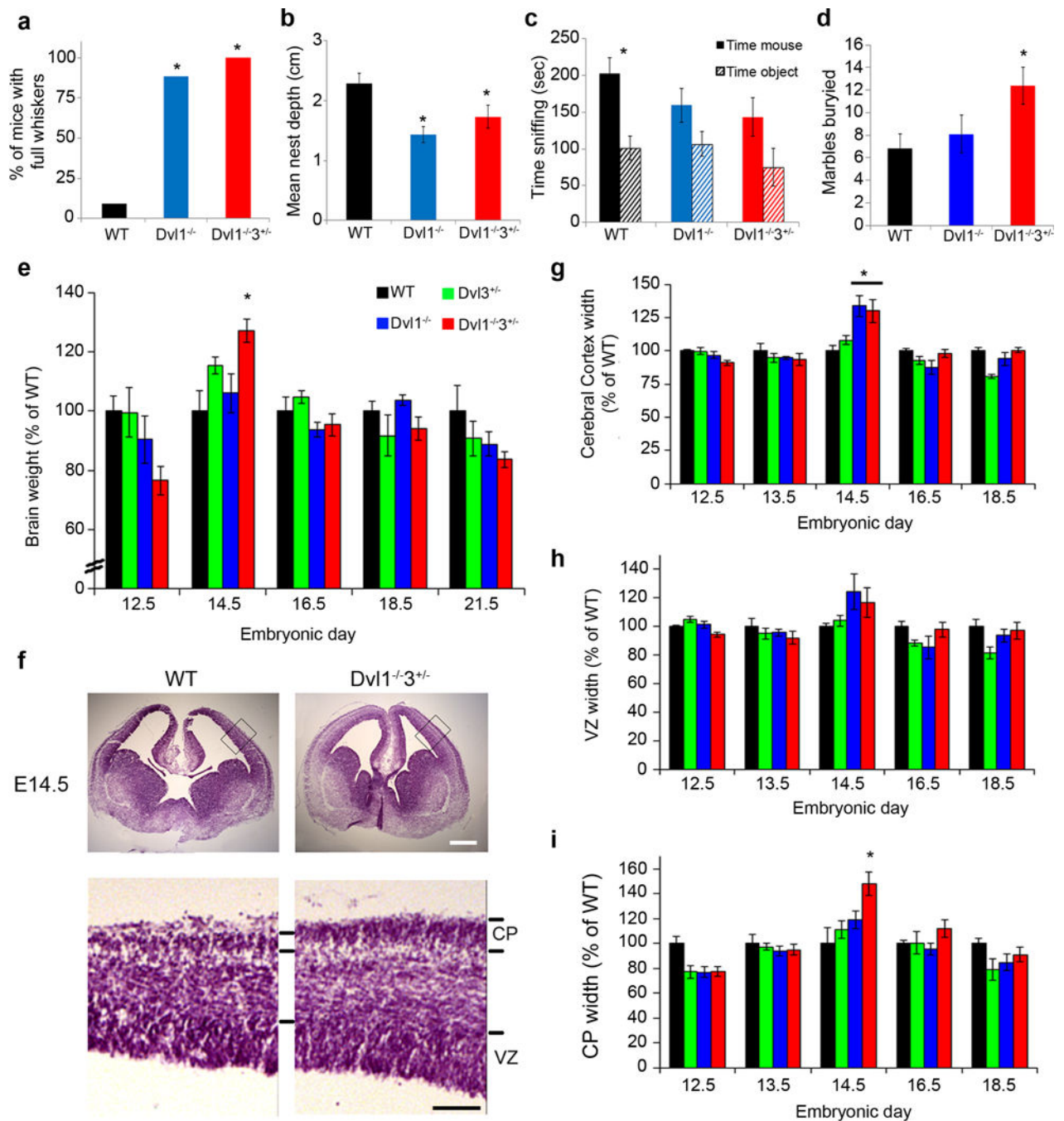


Figure 1. *Dvl1*^{-/-3+/-} mice display social deficits and embryonic brain enlargement

WT, *Dvl1*^{-/-} and *Dvl1*^{-/-3+/-} mice behavioral phenotypes were determined in: (a) Whisker trimming (n 11) (*p<0.0001), (b) nest building (n 5) (*p<0.04), (c) three chamber social approach (n 9) (*p<0.01) and (d) marble burying task (n 8) (*p<0.02). e, WT, *Dvl1*^{-/-} and *Dvl1*^{-/-3+/-} embryonic heads (E12.5–E13.5) or brains (E14.5–P0) were dissected, weighed and quantified as a percentage of the average WT brain weight for the indicated time point (mean±SEM, n 5, ANOVA<0.001; *p<0.04 for comparison of the *Dvl1*^{-/-3+/-} brains with those from WT). f, Coronal sections of WT, *Dvl1*^{-/-} and *Dvl1*^{-/-3+/-} were Nissl stained and

representative images of the neocortex are shown at E14.5 (scale bar: 250; 100 μ m). Nissl stained images were quantified as percentage of the average width of WT total neocortex (**g**), ventricular zone (**h**) and cortical plate (**i**) for the indicated time points. Results are presented as mean \pm SEM (n = 4, ANOVA<0.02; *p<0.02 for comparison of the brains from *Dvll1*^{-/-} *3*^{+/-} and *Dvll1*^{-/-} mutants with those of WT).

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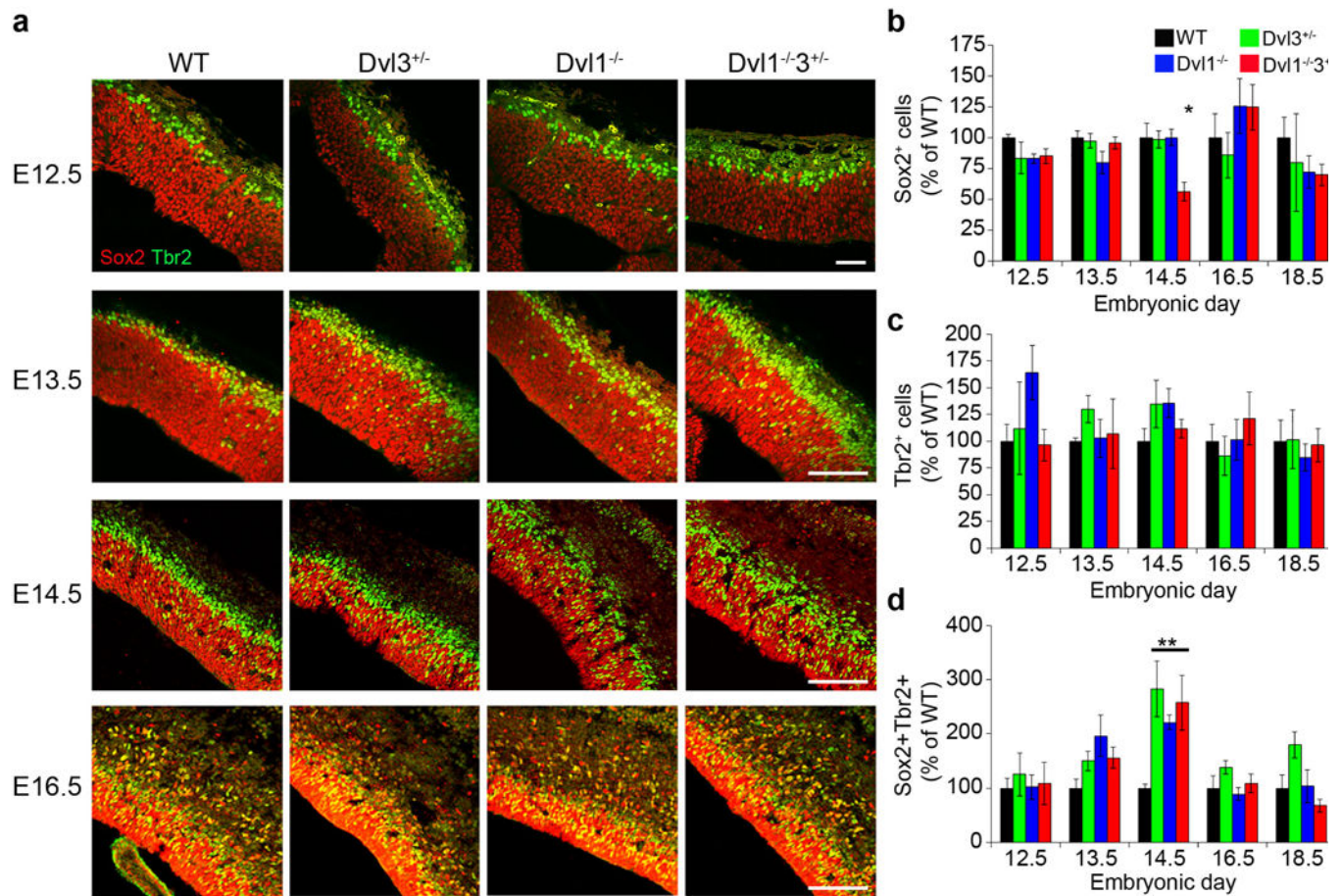


Figure 2. Early expansion of basal progenitors in *Dvl1*^{-/-3^{+/-}} embryos

a. Coronal sections from WT, *Dvl1*^{-/-} and *Dvl1*^{-/-3^{+/-}} embryonic brains were immunostained with anti-Sox2 (Red) and anti-Tbr2 (Green) (scale bar: 100 μ m). Quantification of the single and double labeled cells was performed and results are presented as mean \pm SEM (n = 4). **b.** Sox2 single labeled cells (ANOVA<0.01; *p<0.01 for comparing the results of the *Dvl1*^{-/-3^{+/-}} with those of all the mouse groups); **c.** Tbr2 single labeled cells and **d.** Sox2⁺Tbr2⁺ double labeled cells. (ANOVA<0.02; *p<0.002 for comparing the results of the *Dvl* mutants with those of WT mouse groups).

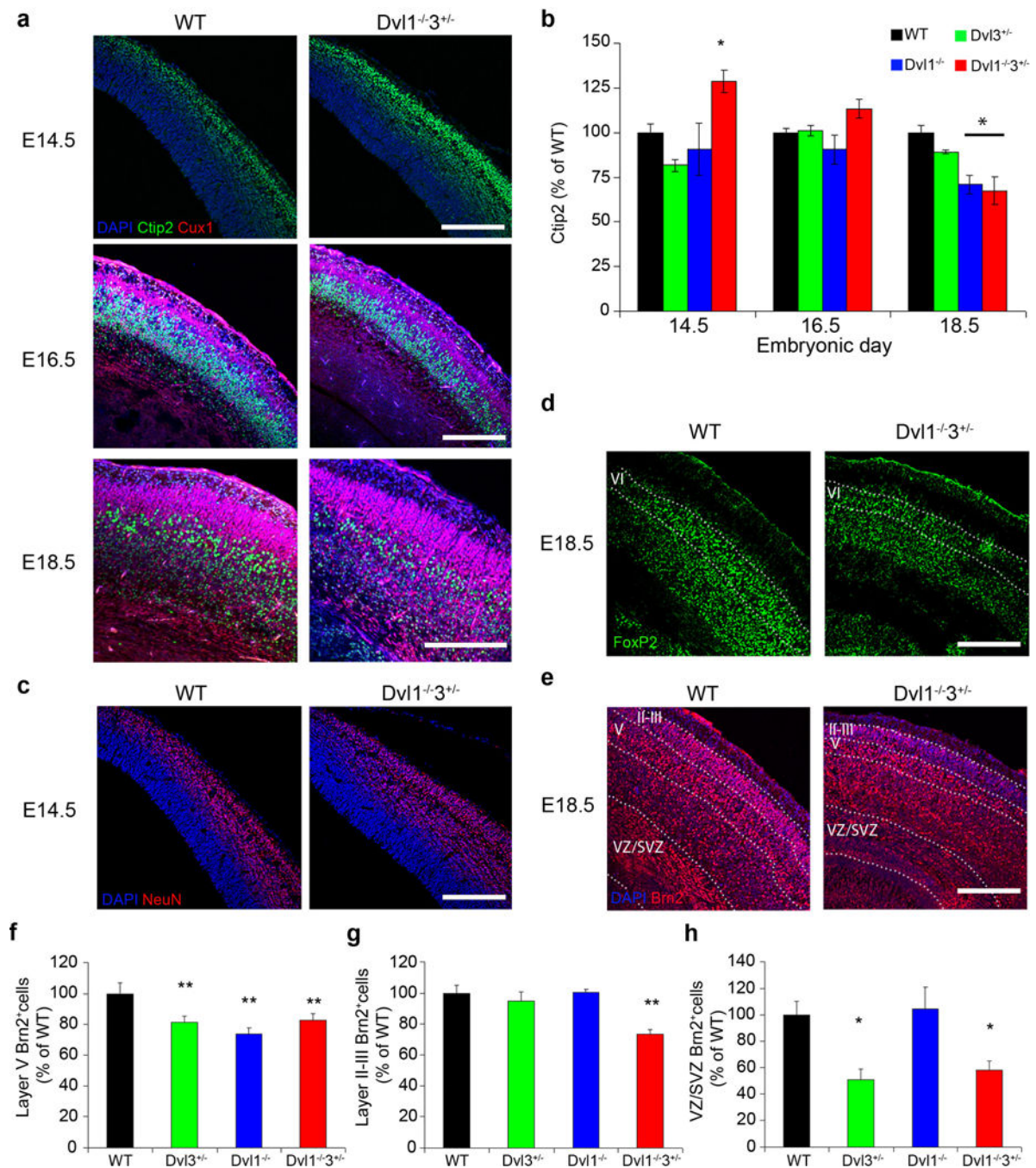


Figure 3. Early differentiation of deep layer neurons in *Dvl1*^{-/-3+/-} embryos

a, WT and *Dvl* mutant embryonic brain sections were immunostained with DAPI (Blue), anti-Ctip2 (Green) and anti-Cux1 (Red) (Scale bar: 200 μ m), Representative images of the staining are shown. **b**, Quantification of the percentage of Ctip2 labeled cells are presented as mean \pm SEM (n = 4, ANOVA<0.007; *p<0.01 for comparing the results of the *Dvl1*^{-/-3+/-} with those of the WT mice). **c**, WT and *Dvl* embryonic brain sections were immunostained with DAPI (Blue), anti-NeuN (Green) and representative images of the staining are shown. (Scale bar: 200 μ m). **d–e**, Coronal sections of WT and *Dvl* embryonic brains at E18.5 were

immunostained with DAPI (Blue), anti-FoxP2 (Green) (**d**) and anti-Brn2 (Red) (**e**). Representative images of the staining are shown. (Scale bar: 200 μ m). **f–h**, Quantification of the percentage of Brn2⁺ neuron in layer V (ANOVA<0.04) (**f**), layers II–III (ANOVA<0.003) (**g**) and VZ/SVZ (ANOVA<0.009) (**h**), results are presented as mean \pm SEM of n=3–5 per group (**p<0.01 and *p<0.05 for comparing the results of the *Dvl* mutants with those of the WT). Black, green, blue and red bars represent WT, *Dvl3*^{+/-}, *Dvl1*^{-/-} and *Dvl1*^{-/-} *3*^{+/-} genotypes, respectively.

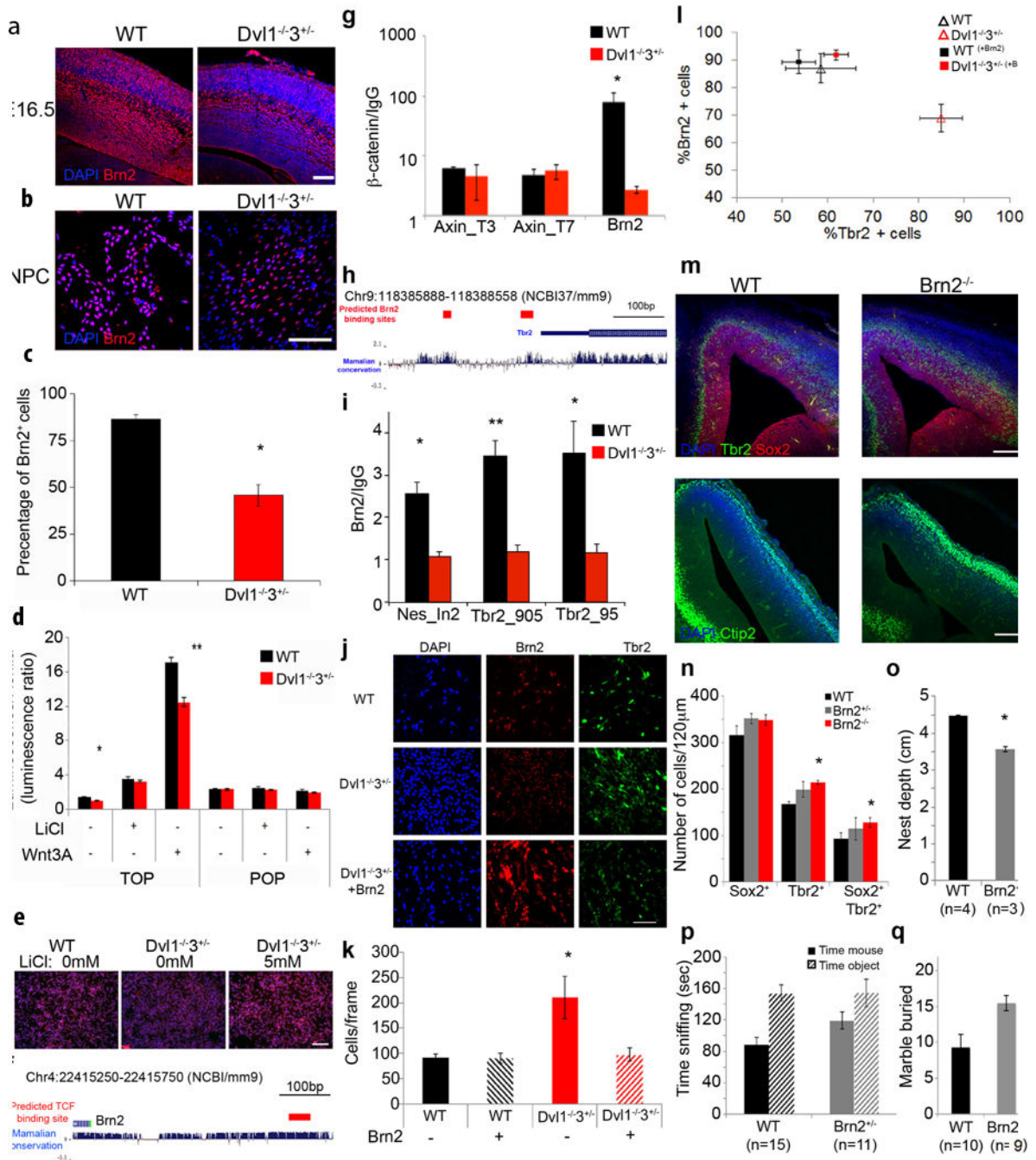


Figure 4. β -catenin/Brn2/Tbr2 transcriptional cascade directly regulates NPCs proliferation
WT and *Dvl1*^{-/-}3^{+/-} brains at E16.5 were double-labeled with DAPI (blue) and anti-Brn2 (red) and representative images are presented (Scale bar: 100μm) (a) and in WT and *Dvl1*^{-/-}3^{+/-} NPCs (Scale bar: 100μm) (b). c, Brn2 labeled NPCs were quantified as the percentage of total cells counted (DAPI). Results are presented as mean±SEM (n = 3, *p<0.01 for comparing the percentage of Brn2⁺ cells in the *Dvl1*^{-/-}3^{+/-} NPCs with those of the WT NPCs). d, WT and *Dvl1*^{-/-}3^{+/-} NPCs were transfected with TOP-Flash and fire-fly renilla reporters and treated with either 5mM LiCl or 100ng/ml Wnt3A. Results are

presented as mean±SEM (n = 3, ANOVA<0.001; *p<0.02 and **p<0.002 for comparing the luminescence ratio in the *Dv11^{-/-}3^{+/-}* NPCs with those of the WT NPCs). **e**, WT and *Dv11^{-/-}3^{+/-}* NPC treated with 5mM LiCl were immunostained for Brn2 and representative images are presented (Scale bar: 100µm). **f**, Schematic Image of the promoter region of Brn2, the predicted region for TCF binding is marked by red rectangle. **g**, At E14.5 WT and *Dv11^{-/-}3^{+/-}* cortices were isolated and β-catenin ChIP was performed followed by qPCR analysis for the Axin2 intron1 and the predicted region on the Brn2 promoter. (mean±SEM, n = 4, *p<0.04 for comparing the enrichment in the *Dv11^{-/-}3^{+/-}* cortical cells with those of the WT cortical cells). **h**, Schematic image of the promoter region of Tbr2, the red rectangle indicates the predicted regions for Brn2 binding. **i**, At E14.5 WT and *Dv11^{-/-}3^{+/-}* cortices were isolated and Brn2 ChIP was performed followed by qPCR analysis for the nestin intron2 and the predicted binding sites on the Tbr2 promoter. Results are presented as fold enrichment of the Brn2 signal above the IgG control (mean±SEM, n = 4, *p<0.04, **p<0.003, for comparing the enrichment in the *Dv11^{-/-}3^{+/-}* cortical cells with those of the WT cortical cells). **j**, WT and *Dv11^{-/-}3^{+/-}* NPCs grown as monolayer cultures were transfected with pCAG-Brn2. NPCs were immunostained for DAPI, Brn2 and Tbr2. Representative images are presented (Scale bar: 100µm). **k**, pCAG-Brn2 transfected WT and *Dv11^{-/-}3^{+/-}* NPCs were immunostained for DAPI and DAPI⁺ nuclei were counted. Results are presented as mean±SEM (n = 4, ANOVA<0.04 *p<0.05 for comparing the results of the *Dv11^{-/-}3^{+/-}* with those of the WT and the Brn2 transfected *Dv11^{-/-}3^{+/-}* NPCs groups). **l**, pCAG-Brn2 transfected WT and *Dv11^{-/-}3^{+/-}* NPCs were immunostained for DAPI, Brn2 and Tbr2. The percentage of Brn2⁺ and Tbr2⁺ cells was measured. Results are presented as mean±SEM (n = 4, ANOVA<0.04 *p<0.03, comparing the results of the GFP transfected *Dv11^{-/-}3^{+/-}* with those of the WT and the Brn2 transfected *Dv11^{-/-}3^{+/-}* NPCs). **m**, WT and *Brn2^{-/-}* brain coronal sections from E14.5 were immunostained with anti-Sox2 (Red) and anti-Tbr2 (Green) and representative images are presented (scale bar: 100µm). WT and *Brn2^{-/-}* brain coronal sections from E14.5 were immunostained with anti-Ctip2 (Green) and representative images are presented (scale bar: 100µm). **n**, The number of Sox2⁺, Tbr2⁺ and Sox2⁺Tbr2⁺ labeled cells were quantified and the results are presented as mean±SEM (n = 4, ANOVA<0.04; *p<0.001 for comparing the results of the Brn2 mutants with those of the WT embryos). **o**, Nest building activity in WT and *Brn2^{+/-}* cages was assessed and measurements of the nest depth are presented as mean±SEM (n = 4, *p<0.003 for comparing the results of the Brn2 mutants with those of the WT cages). **p**, WT and *Brn2^{+/-}* mice were assessed in the social approach task and scored, results are presented as mean±SEM (*p<0.002 for comparing the results of the time spent sniffing the mouse compared with the object). **q**, The marble burying task was scored and the results are presented as mean±SEM (*p<0.02 for comparing the results of the *Brn2^{+/-}* mutants with those of the WT mice).

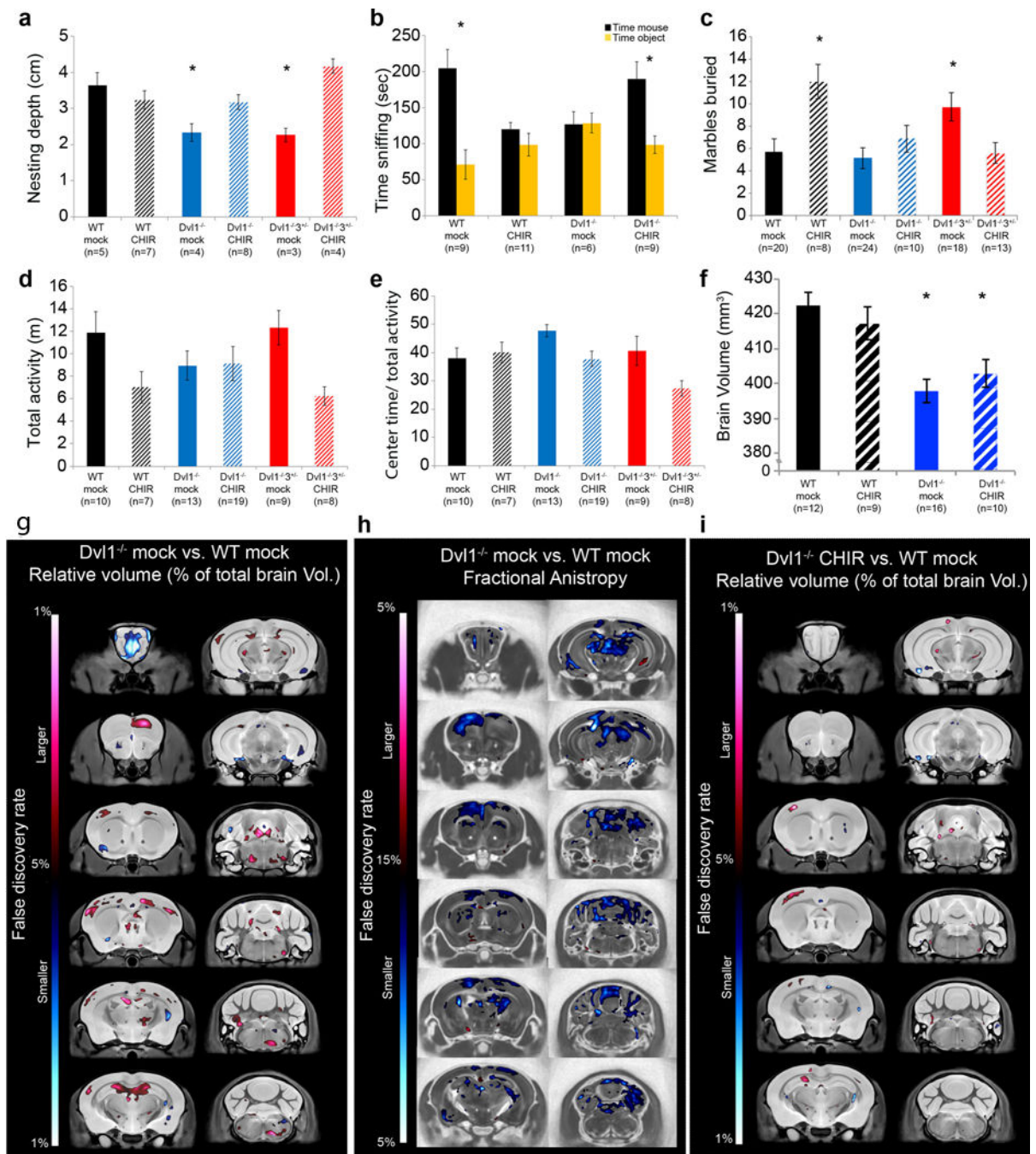


Figure 5. Embryonic activation of Wnt canonical pathway regulates *Dvl1*^{-/-}3^{+/-} embryonic brain phenotype

a, WT and *Dvl1* mutant brains at E14.5 following CHIR99021 treatment were Nissl stained and representative images are presented (scale bar: 100 μ m). Nissl stained images were quantified as a percentage of the average width of WT total neocortex (ANOVA<0.02) (**b**), ventricular zone (**c**) and cortical plate (ANOVA<0.001) (**d**). Results are presented as mean \pm SEM (n = 4, ANOVA<0.01; *p<0.02 for comparison of the brains from *Dvl1*^{-/-}3^{+/-} mock treated with those of WT mock treated embryos). **e**, Coronal sections of WT and *Dvl1* mutant

mock and CHIR99021 treated embryo brains at E14.5 were double label with anti-Sox2 (Red) and anti-Tbr2 (Green) and representative images are presented (scale bar: 100µm). Quantification presented as mean±SEM of Sox2 single labeled cells (**f**); Tbr2 single labeled cells (**g**) (n = 4, ANOVA<0.05 *p<0.03 for comparing the results of the WT mock and CHIR99021 treated embryos) and Sox2⁺Tbr2⁺ double labeled cells (**h**) (ANOVA<0.006; *p<0.01 for comparing the results of the *Dvl1*^{-/-} and *Dvl1*^{-/-} 3^{+/-} mock treated with those of the other embryo genotypes). (**i**) WT and *Dvl1*^{-/-} 3^{+/-} brains at E14.5 from mock or CHIR99021 treated embryos were immunostained with DAPI (Blue) and anti-Ctip2 (Green) and representative images are presented (Scale bar: 200µm). **j**, Quantification of Ctip2⁺ cells (mean±SEM) are presented as percentage of the average number of Ctip2⁺ cells from WT mock treated embryos (n = 4, ANOVA<0.02; *p<0.01 for comparing the results of the *Dvl1*^{-/-} 3^{+/-} mock treated with those of the *Dvl1*^{-/-} 3^{+/-} CHIR99021 treated and WT embryos).

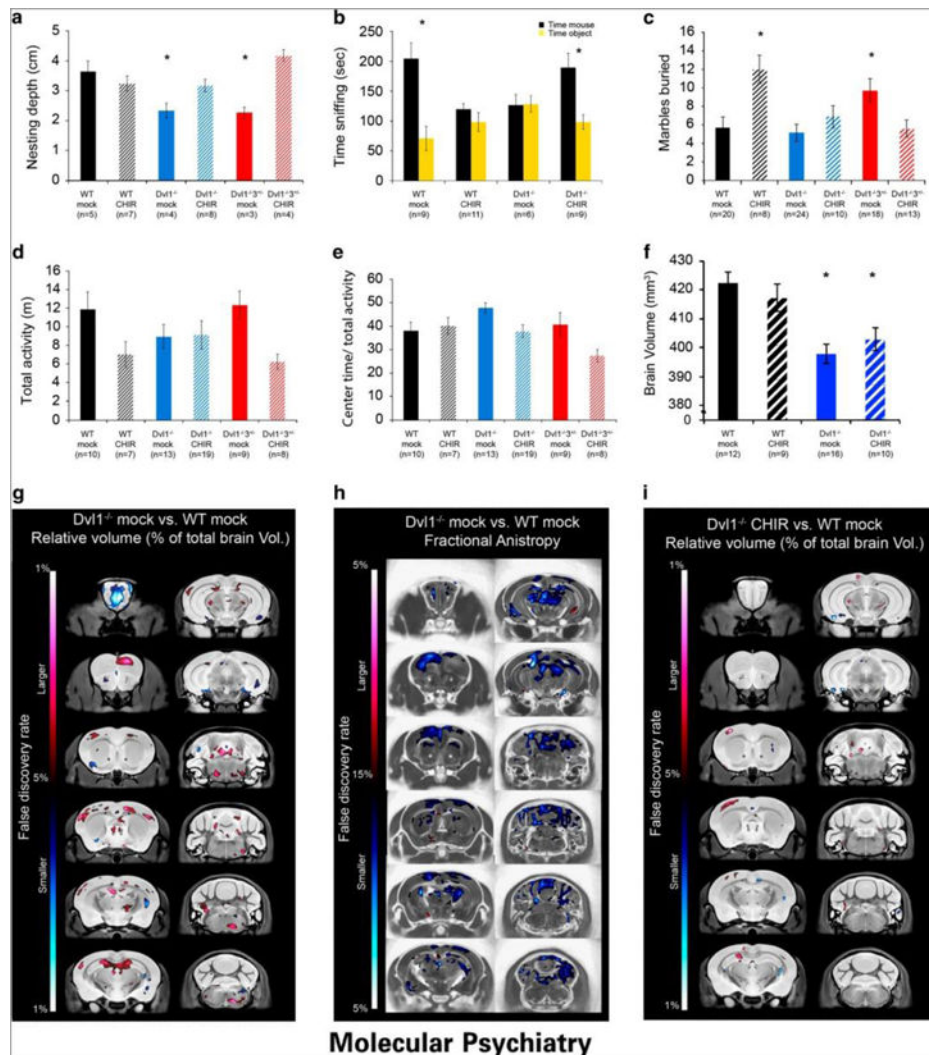


Figure 6. Prenatal Wnt canonical pathway regulates adult social behavior

WT and *Dvl* mutant (*Dvl1*^{-/-} and/or *Dvl1*^{-/-} 3^{+/-}) adult mice treated with CHIR99021 or mock treated only as embryos (E9.5–14.5) were tested in behaviors found to be abnormal in the *Dvl* mutants. **a**, Nest depth was measured after 24hr and the recorded depth are presented as mean±SEM (n = 3, ANOVA<0.002; *p<0.04 for comparing the results of the *Dvl1*^{-/-} and *Dvl1*^{-/-} 3^{+/-} mock treated mice with those of the other mouse groups). **b**, The social approach task was recorded, scored and the results are presented as mean±SEM (*p<0.02 for comparing the results of the time spent sniffing the mouse compared with the object). **c**, The marble burying task was scored and the results are presented as mean±SEM (ANOVA<0.001; *p<0.03 for comparing the results of the *Dvl1*^{-/-} 3^{+/-} mock treated and WT CHIR99021 treated mice with those of the other mouse). Total activity in the open field task (in meters distance traveled, **d**) the time spent in the center of the arena relative to their total activity (**e**) in the open field task are presented as mean±SEM. There were no significant genotypic differences in these tasks. WT and *Dvl1*^{-/-} mock and CHIR mice treated *in utero* between E9.5–14.5 (n = 9) were imaged using an MRI separately, with both a T2-weighted anatomical scan and a subsequent Diffusion Tensor Imaging (DTI) scan. (**f**) Total brain

volume (mm^3) for the four groups are presented as $\text{mean} \pm \text{SEM}$ (* $p < 0.0004$ for comparison of the results from $Dv11^{-/-}$ with those of WT brains). **g**, Voxel-wise analysis highlighting significant relative volume differences (false discovery rate, FDR) throughout the brain between the mock treated $Dv11^{-/-}$ and WT mice (**h**) Fractional anisotropy (FA) analysis of significant differences (FDR) throughout the brain between the mock treated $Dv11^{-/-}$ and WT mice. **i**, Voxel-wise analysis of significant relative volume differences (FDR) throughout the brain between the $Dv11^{-/-}$ CHIR and WT mock treated mice.