

UC Merced

UC Merced Previously Published Works

Title

Purinergic signaling during Porphyromonas gingivalis infection

Permalink

<https://escholarship.org/uc/item/0736q62c>

Journal

Biomedical Journal, 39(4)

ISSN

2319-4170

Authors

Almeida-da-Silva, Cássio Luiz Coutinho
Morandini, Ana Carolina
Ulrich, Henning
[et al.](#)

Publication Date

2016-08-01

DOI

10.1016/j.bj.2016.08.003

Peer reviewed

Available online at www.sciencedirect.com

ScienceDirect

Biomedical Journal

journal homepage: www.elsevier.com/locate/bj

Review Article: Special Edition

Purinergic signaling during *Porphyromonas gingivalis* infection



Cássio Luiz Coutinho Almeida-da-Silva^a, Ana Carolina Morandini^{b,d},
Henning Ulrich^c, David M. Ojcius^{a,d,1}, Robson Coutinho-Silva^{a,*,1}

^a Immunobiology Program, Biophysics Institute of the Federal University of Rio de Janeiro, Brazil

^b Department of Oral Health Sciences, College of Dental Medicine, Medical University of South Carolina, Charleston, SC, USA

^c Department of Biochemistry, Institute of Chemistry, University of São Paulo, São Paulo, Brazil

^d Department of Biomedical Sciences, University of the Pacific, San Francisco, USA



Prof. Robson
Coutinho-Silva

ARTICLE INFO

Article history:

Received 11 April 2016

Accepted 2 August 2016

Available online 24 September 2016

Keywords:

Purinergic receptors

Innate immunity

Porphyromonas gingivalis

P2X7 receptor

Oral microbes

Inflammasome

ABSTRACT

Despite recent advances unraveling mechanisms of host–pathogen interactions in innate immunity, the participation of purinergic signaling in infection-driven inflammation remains an emerging research field with many unanswered questions. As one of the most-studied oral pathogens, *Porphyromonas gingivalis* is considered as a keystone pathogen with a central role in development of periodontal disease. This pathogen needs to evade immune-mediated defense mechanisms and tolerate inflammation in order to survive in the host. In this review, we summarize evidence showing that purinergic signaling modulates *P. gingivalis* survival and cellular immune responses, and discuss the role played by inflammasome activation and cell death during *P. gingivalis* infection.

Innate immunity and oral microbes

The host organism is always ready to respond to foreign stimuli, such as infection by pathogens. The first response of the host immune system is carried out by innate immunity. Unlike the adaptive response (which includes specific antibodies and lymphocytes), the repertoire of the innate

response is common among all normal and healthy individuals. This response involves cellular and humoral activities, as well as chemical (e.g. acidic stomach pH, saliva and tears) and anatomical barriers (e.g. epithelial cells throughout the body).

Cellular responses to pathogens depend on the recognition of evolutionarily conserved structures that are typically

* Corresponding author. Biophysics Institute of the Federal University of Rio de Janeiro, Health Sciences Center, G. Block, Av. Carlos Chagas Filho, 373, University City, Island of Fundão, Rio de Janeiro, RJ 21941-902, Brazil. Tel.: +55 21 3938 6565; fax: +55 21 2281 8193.

E-mail address: rscsilva@biof.ufrj.br (R. Coutinho-Silva).

Peer review under responsibility of Chang Gung University.

¹ DMO and RCS are joint senior authors on this work.

<http://dx.doi.org/10.1016/j.bj.2016.08.003>

2319-4170/© 2016 Chang Gung University. Publishing services by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

present in microbes but not in the host. These molecules, called “Pathogens Associated Molecular Patterns” (PAMPs), are recognized by innate immune cells by “Pattern Recognition Receptors” (PRRs). PRRs include Toll-like receptors (TLRs), nucleotide-binding and oligomerization domain (NOD)-like receptors (NLRs), retinoic-acid-inducible gene I (RIG-I)-like receptors (RLRs), and the C-type lectin receptors (CLRs), as well as DNA receptors (cytosolic sensors for DNA) [1–3]. Different PAMPs can be recognized by PRRs, whose ligation leads to activation of transcription factors, such as activator protein 1 (AP-1) and nuclear factor kappa B (NF- κ B), which in turn, modulate gene transcription of pro-inflammatory cytokines and chemokines [4]. This response is important for the host to control infections and prevent disease.

Knowledge about the human microbiota and its relationship with the host is also crucial for better understanding the mechanisms of immunity, since the microbiota can stimulate and modulate the immune system. The human body contains 10 times more prokaryotic than eukaryotic cells, and humans and microbes have evolved to gradually become dependent on one another [5]. The oral mucosa is associated with hundreds of different viruses and bacterial, archaeal, fungal, and protozoan species, many of which can interact to form biofilms conferring resistance to the microorganisms against mechanical and chemical stress [5,6]. In this context, bacterial communities found in the oral mucosa are highly complex (comprising around 1000 species) and are one of the most complex in the whole body [6]. Most of the oral microbes live as commensals within the host, but some species can become pathogenic in response to the host genotype, stress, diet or behavior (e.g. smoking) [7].

Not surprisingly, some oral microbes have been related to oral disorders, including periodontal disease (or periodontitis), which is commonly manifested as a chronic and inflammatory condition induced by biofilms and pathogens closely associated with the periodontium (the structures that protect and support teeth, such as gingiva, periodontal ligament and alveolar bone). When the periodontium is damaged, a process of uncontrolled bone resorption is initiated and can lead to tooth loss as a consequence [7,8]. Socransky and colleagues identified several bacteria present in subgingival biofilms from individuals with periodontitis and grouped these bacteria in complexes according to their association with disease [9]. Among these complexes, the one most strongly associated with periodontitis is the so-called “red complex” bacteria. The “red complex” consists of *Tannerella forsythia*, *Treponema denticola* and *Porphyromonas gingivalis*, which are related to human periodontitis because of their strong association with the diseased sites [9,10].

P. gingivalis is an anaerobic, asaccharolytic, black-pigmented, non-motile and non-spore forming Gram-negative bacterium, which exists as different strains with variable virulence [11–13]. *P. gingivalis* is mainly found during diseased states but it can be also found in healthy individuals, with a prevalence that ranges around 25% in healthy individuals and 79% in individuals with periodontitis [14]. Recent studies have suggested a new paradigm of periodontal pathogenesis, which assigns a larger role for some bacteria from the oral microbiota and host susceptibility in

development of disease. Usually, pathogenic bacteria lead to inflammation by direct infection and dysregulation of the commensal microbiota. *P. gingivalis* was described as a “keystone pathogen” due to its ability to induce dysbiosis and inflammation even in relatively low numbers. Using a murine model of experimental periodontitis, it was shown that, even in low-abundance after inoculation, *P. gingivalis* can infect the oral mucosa and promote changes in numbers and composition of the oral commensal microbiota, leading to a dysbiotic environment [15]. Dysbiosis is characterized by an imbalance in the relative abundance of species within the microbiota that is related to disease induction [7]. This study also showed that *P. gingivalis* inoculation in the oral cavity led to bone loss in specific pathogen free (SPF) mice, but *P. gingivalis* alone could infect but did not induce bone loss in germ-free mice. These data resulted in the “keystone pathogen hypothesis” of periodontal disease [15].

Besides being considered a keystone pathogen, this bacterium is also associated with increased risk of diverse systemic diseases. Oral health is important for overall health since it has been shown that periodontitis increases the patients’ risk for atherosclerosis, rheumatoid arthritis, and cancer [16–20]. In fact, during periodontal disease, bacteria reach blood vessels temporarily and this transient bacteremia is responsible for spreading bacteria to different sites of the organism, such as atherosclerotic plaques (as detected by PCR [17]), where they can accelerate pathogenesis.

P. gingivalis has evolved several virulence factors to evade innate and adaptive immunity and cause disease. There is a large body of evidence describing the ambivalent behavior of this microorganism, which on one hand needs to escape immune-mediated detection and on the other hand tolerates and perpetuates inflammation to survive in the host. It was shown that *P. gingivalis* fimbriae can highly activate human monocytes and poorly activate epithelial cells with regards to interleukin (IL)-6, IL-8, macrophage colony stimulating factor (M-CSF) and tumor necrosis factor (TNF)- α responses, thus reflecting different strategies used by this bacterium when interacting with distinct host cell types [21]. As another example, *P. gingivalis* can manipulate host neutrophils through complement C5a receptor and TLR2 pathways in a cooperative crosstalk involving downstream adaptor molecules such as MyD88 [22]. This study showed higher bacterial survival in infected MyD88-deficient mice compared with wild-type mice. *In vitro*, MyD88-depleted human periodontal fibroblasts showed decreased expression of major inflammatory cytokines such as IL-6 and IL-8 compared to control cells when stimulated with *P. gingivalis* LPS [23]. Moreover, cysteine proteinases called gingipains from *P. gingivalis* can influence the composition of polymicrobial biofilms [24]. This work revealed an interdependency between the gingipains of *P. gingivalis* and *T. forsythia* or *T. denticola*, thus suggesting supported survival and virulence of the biofilm community as a whole. The capacity of *P. gingivalis* to interact with other periodontal pathogens such as *Fusobacterium nucleatum* was also demonstrated in the case of *P. gingivalis*’ ability to suppress inflammasome activity [25], which will be discussed later in this review. Finally, *P. gingivalis* expresses hemagglutinins, as well as an atypical and less immunogenic LPS, which can be an antagonist of TLR4 ligation; a serine phosphatase B

(SerB) which inhibits IL-8 synthesis by gingival epithelial cells (GECs); fimbriae that are involved in bacterial adhesion; and a nucleoside-diphosphate-kinase (NDK), which hydrolyzes extracellular ATP (eATP) and will be discussed in more detail later in this review.

P. gingivalis can also evade different mechanisms of adaptive immunity. Human neutrophils, peripheral blood mononuclear cells (PBMCs) and GECs infected with *P. gingivalis* produced IL-1 β but not the T-cell chemokine CXCL10 [26]. Thus, *P. gingivalis* infection inhibited interferon (IFN)- γ -induced and *F. nucleatum*-induced CXCL10 secretion by epithelial cells. Moreover, *P. gingivalis* adhesion induced morphological changes, reactive-oxygen-species (ROS) production and increased intracellular Ca⁺² levels in T-cells [27]. This periodontal pathogen also inhibited AP-1 and NF- κ B activity, as well as IL-2 accumulation, by means of its gingipains. Another study demonstrated that gingipains of *P. gingivalis* cleave immunoglobulin G1 (IgG1) in gingival crevicular fluid of patients and may suppress antibody-dependent antibacterial activity *in vivo* [28]. Also, it was shown that, upon initial encounter with *P. gingivalis in vivo*, murine splenic T cells and CD11b⁺ cells produced IL-10, and this cytokine suppressed IFN- γ T cell responses [29]. Furthermore, it was demonstrated that supernatants of human immune cells infected with two different strains of *P. gingivalis*-induced T helper cell polarization to a Th17 profile instead of a Th1 profile [30]. Moreover, *P. gingivalis* favored the generation of Th17-related cytokines such as IL-1 β , IL-6 and IL-23 but not the Th1-related cytokine, IL-12. Interestingly, another laboratory showed that subcutaneous vaccination with formalin-killed *P. gingivalis* protected the mice from alveolar bone resorption and inflammation by downregulation of Th17 cells and IL-17A production, while promoting upregulation of regulatory T (Treg) cells, IL-10 and transforming growth factor- β 1 (TGF- β 1) [31].

Purinergic signaling in the context of infection and inflammation

ATP is traditionally associated with cellular energy metabolism in all prokaryotic and eukaryotic cell types, but it is also recognized that ATP and other nucleotides are released from cells following stress or injury [32,33]. It has been shown that controlled and uncontrolled mechanisms of ATP release to the extracellular space takes place during cellular stress, death or tissue injury. It has been demonstrated thus far that ATP is released from necrotic cells via pannexin channels, connexin hemichannels and also via the P2X7 receptor [34,35]. In the extracellular compartment, nucleotides can be recognized by the host immune system as danger signals and can promote several biological activities in different immune cell types. Examples include: maturation of immature dendritic cells, secretion of pro-inflammatory cytokines by macrophages, chemotaxis and IL-8 production by eosinophils, and costimulation for antigenic stimulation by T and B cells [35,36]. These molecules bind to purinergic receptors expressed on virtually all immune cell types [33].

Purinergic receptors are divided into two families: P1 and P2 receptors [32]. The G-protein coupled metabotropic P1 receptors recognize exclusively adenosine and can be

subdivided into A₁, A_{2A}, A_{2B} and A₃ receptors, all of which with different binding affinities for adenosine [32,37]. The receptors A₁ and A₃ are coupled to protein G_i, and A_{2A} and A_{2B} becoming associated with protein G_s [37,38]. The P2 receptors can be subdivided into two subtypes: non-selective ion-gated channel P2X receptors (that recognize ATP) and G-coupled P2Y receptors (that recognize ATP, ADP, UTP, UDP and UDP-glucose) [35,36]. To date, seven P2X receptors (from P2X1 to P2X7) with different affinities for ATP have been described. Among these receptors, the P2X7 receptor has a low affinity for ATP (requiring ≥ 100 μ M to be activated; while others can be activated at lower concentrations) and has been associated with immune responses and inflammation, such as inhibition of infection by intracellular pathogens and activation of the inflammasome, which will be discussed later in this review [4,39–41,41].

ATP ligation of the P2X7 receptor leads to opening of a transmembrane non-selective cationic channel that allows K⁺ efflux and Na⁺ and Ca²⁺ influx and promotes cytoplasmic membrane depolarization [35]. P2X7 receptor activation is associated with pore formation, which depends on the concentration and duration of ATP treatment [41]. Continuous stimulation of the P2X7 receptor with ATP can induce cell death either by necrosis or apoptosis [42], as well as lead to opening of a pore that allows the passage of molecules up to 900 Da [43].

Since recognition of high levels of eATP results in modulation of immune responses, the host has a sophisticated and sensitive mechanism to regulate the composition, duration, intensity and magnitude of purinergic signaling, as reviewed elsewhere [41]. Immune and non-immune cells utilize a group of nucleotide-hydrolyzing enzymes called ecto-nucleotidases to control exacerbated levels of nucleotides and maintain steady-state conditions. The ecto-nucleoside triphosphate diphosphohydrolases (E-NTPDases which degrade extracellular tri- and diphosphonucleosides to monophosphonucleosides), ecto-nucleotide pyrophosphatase/phosphodiesterases (E-NPPs that hydrolyze pyrophosphate and phosphodiester bonds in a wide range of substrates), and ecto-5'-nucleotidase (which degrades AMP) are the most relevant ecto-nucleotidases in the context of innate immunity [41]. These enzymes are responsible for maintaining healthy and stable levels of eATP [36] and generating adenosine, a metabolite of ATP breakdown [41].

While ATP exhibits pro-inflammatory and stimulatory effects in the immune system and physiology [35,44–51], adenosine has primarily anti-inflammatory and inhibitory effects [37,52–56]. Therefore, the balance between ATP versus adenosine levels is important in modulating cellular immune responses and pathogen survival [Fig. 1].

ATP effects in intracellular pathogen infection

Immune and epithelial cells activate microbicidal pathways and contribute to inflammation after ligation of purinergic P2 receptors by eATP [Fig. 1]. Facultative and obligate intracellular microbes survive inside the host cell, where they acquire nutrients needed for microbial replication and propagation of the infection [53,57]. For intracellular pathogens, it is

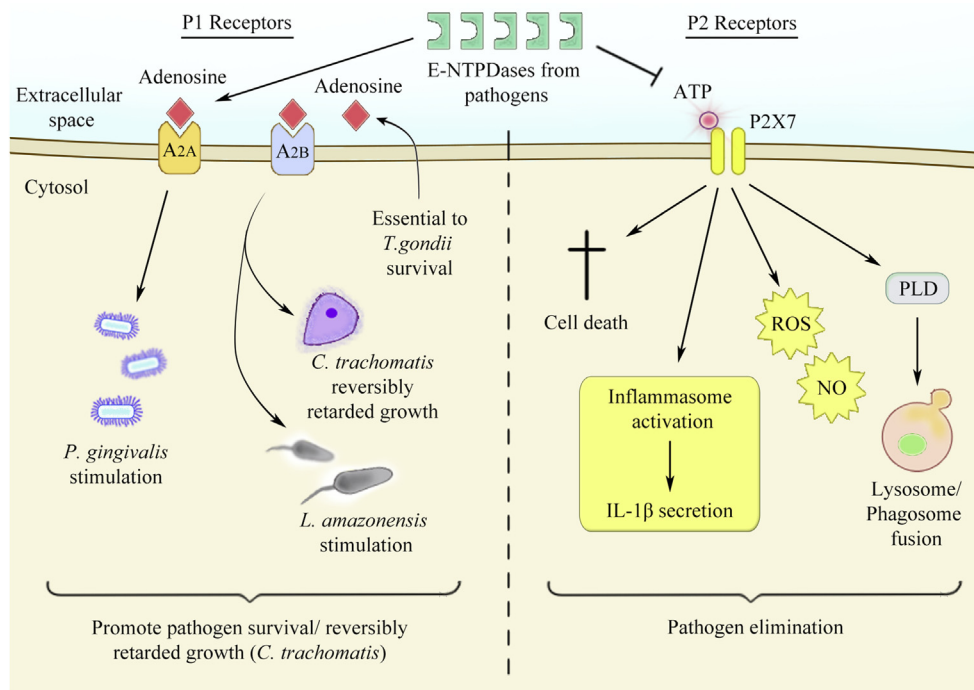


Fig. 1 – Schematic figure of the antagonistic effects of P1 and P2 receptors on infected cells. On the left, extracellular adenosine is recognized by P1 receptors (for example, A_{2A} and A_{2B}) and can promote pathogen survival or reversibly retard microbial growth. On the other hand, on the right, eATP released from stressed, dying or infected cells binds to P2 receptors (for example, P2X7) and leads to pathogen elimination through several pathways: (1) host cell death; (2) inflammasome activation and IL-1 β secretion; (3) ROS and NO production; (4) or phospholipase D activation, promoting lysosome and phagosome fusion. Importantly, ecto-nucleotidases (E-NTPDases) from several pathogens inhibit pathogen elimination by eATP cleavage and/or favor microbial survival by generating extracellular adenosine.

advantageous to be able to prevent or delay apoptosis of the host cell in order to promote survival and growth of the pathogen. In this regard, several pathogens evolved different mechanisms to promote their own growth inside the host cell, as for example *Mycobacterium tuberculosis*, *Chlamydia trachomatis*, *Leishmania* species, *Toxoplasma gondii* and also *P. gingivalis*.

Unlike activation of surface CD95 (Fas receptor), leading to host cell apoptosis, or complement-mediated host cell lysis, which do not induce mycobacterial death [50], eATP treatment of macrophages enhances their antimicrobial properties in a P2X7 receptor-dependent manner. We and others showed that eATP-related killing of *M. tuberculosis* and *C. trachomatis* within human and murine macrophages is mediated by phospholipase D, which is associated with mobilization of intracellular Ca^{+2} and consequently lysosomal fusion and acidification of the phagosomes containing the pathogen [44,48,49]. In another study, it was shown that adenine nucleotides (adenosine, AMP and ATP) can inhibit *C. trachomatis* growth in epithelial cells [58]. Moreover, millimolar eATP concentrations inhibit chlamydial infection via P2X7 receptor in macrophages [48], while micromolar eATP concentrations reversibly inhibit chlamydial infection via the P2X4 receptor in epithelial cells [58].

Infection with the protozoan parasite *Leishmania amazonensis* is also controlled by eATP treatment. Murine

macrophages infected with *L. amazonensis* showed enhanced P2X7 receptor expression *in vitro* and were more responsive to eATP activation *in vitro* and *in vivo*, where cells from established cutaneous lesions were more sensitive to eATP than cells from uninfected mice [46]. Additionally, elimination of *Leishmania* via eATP ligation of the P2X7 receptor is involved with leukotriene B4 production in a 5-lipoxygenase dependent-manner [45,46]. Another work showed that UTP, but not UDP, inhibits *L. amazonensis* infection in murine macrophages, inducing morphological damage in the intracellular parasite, promoting apoptosis of macrophages and production of oxygen and nitrogen reactive species, and increasing intracellular Ca^{+2} concentrations [59]. These effects are believed to occur via P2Y2 and P2Y4 receptors following their upregulation during *L. amazonensis* infection. Periodate-oxidized ATP also induces morphological changes directly in the parasite, dampening the attachment and entry of the protozoa in murine macrophages [60].

Another intracellular protozoan parasite affected by purinergic signaling is *T. gondii*. eATP treatment in infected macrophages promotes *T. gondii* elimination via P2X7 receptor through acidification of the parasitophorous vacuole and ROS production [47,51]. Additionally, our group recently demonstrated that UTP and UDP treatment in murine macrophages infected with *T. gondii* promotes 90% elimination of the parasite, without inducing NO, ROS or apoptosis in the host cell

[61]. Interestingly, UTP and UDP induced parasite egress from the host cell via P2Y2, P2Y4 and P2Y6, thus compromising infectivity and replication of the egressed parasites.

The expression of P2X2, P2X4, P2X5, P2X6 and P2X7 receptors was also reported in GECs, and eATP, unlike other extracellular nucleotides such as UTP and ADP, induced apoptosis of these cells [62]. These studies also demonstrated that *P. gingivalis* infection inhibits eATP-induced apoptosis in GECs through a mechanism that depends on a homolog of nucleoside-diphosphate-kinase (NDK), which is secreted by *P. gingivalis* [62,63]. NDK is an ubiquitous enzyme that is highly conserved in prokaryotes and eukaryotes, including plants [64,65]. The main role of NDK is to catalyze the transfer of terminal phosphate groups from 5'-triphosphate- to 5'-diphosphate-nucleotides [64–66]. Thus NDK has the ability to hydrolyze (and also synthesize) any NTP/dNTP [65,67]. In this context, the NDK from *P. gingivalis* can hydrolyze eATP after infection in human GECs. The enzyme can thus (1) diminish eATP-induced apoptosis [62]; (2) inhibit eATP-induced ROS via P2X7/NADPH oxidase signaling [63]; and (3) attenuate eATP-

induced inflammasome activation, therefore impairing IL-1 β release [68]. In contrast, the presence of NDK from *Pseudomonas aeruginosa* [69] and *P. gingivalis* during infection in murine macrophages contributes to pro-IL-1 β production/stability and induces IL-1 β secretion [unpublished data – Fig. 2], thus demonstrating differences in the role of NDK from *P. gingivalis* after infection in different cell lines.

Adenosine effects on *P. gingivalis* infection

Adenosine is also considered as a danger signal, which can be released from stressed, necrotic or dying cells, or can be generated via dephosphorylation of eATP by the ecto-nucleotidases CD39 and CD73, as reviewed elsewhere [41,70]. In favor of the role of adenosine in downregulating immune responses and inflammation [37,52,53,55,56] [Fig. 1], a recent study has found that agonist of adenosine receptors, when added exogenously in vitro, stimulates the growth of *P. gingivalis* in GECs [54]. This work demonstrated that GECs express all the

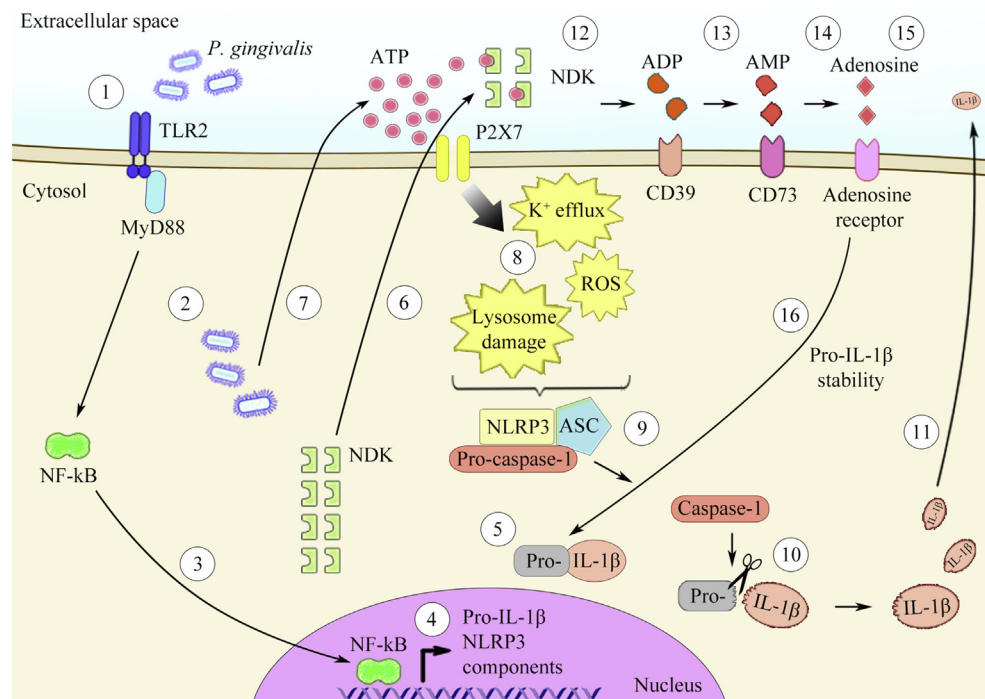


Fig. 2 – Schematic figure of the effects of P1 receptors, P2 receptors, and ecto-nucleotidases on *P. gingivalis*-induced inflammasome activation. (1) As the first signal required for inflammasome activation, *P. gingivalis* is recognized by the host cell through TLR2, and (2) reaches the cytosolic compartment. (3) Recognition of the bacterium via TLR2 promotes the translocation of the transcriptional factor NF- κ B to the nucleus. (4) Once in the nucleus, NF- κ B induces pro-inflammatory cytokines and transcription of inflammasome components (5) and promotes pro-IL-1 β synthesis. (6) Concomitantly, *P. gingivalis* secretes NDK, which may be released to the extracellular compartment. (7) In addition, after infection, this oral bacterium induces ATP release from the host cell to the extracellular space. As a second signal for inflammasome activation, (8) ATP ligation to the P2X7 receptor can promote K⁺ efflux, ROS generation and/or lysosome damage, (9) which can activate the NLRP3 inflammasome. The NLRP3 inflammasome activates procaspase-1 to the mature caspase-1, and (10) this enzyme, in turn, proteolytically processes pro-IL-1 β into IL-1 β , (11) which is released from the cell. (12) NDK from *P. gingivalis* hydrolyzes eATP, generating its own metabolites, such as ADP, (13) which are recognized and cleaved via ecto-nucleotidases from the host, such as CD39, generating AMP. (14) AMP binds to CD73 on the host cell, and this enzyme can generate adenosine. (15) Adenosine interaction via adenosine receptor (16) can promote pro-IL-1 β stability and thus, supports IL-1 β secretion.

adenosine receptors, and stimulation of the A_{2A} receptor via CGS21680 specific-agonist could enhance proliferation of *P. gingivalis*. On the other hand, a high-affinity adenosine receptor agonist NECA, or adenosine by itself, could reversibly inhibit growth of the intracellular bacterium *C. trachomatis* via A_{2B} receptors [71]. Adenosine is also important in survival of protozoan parasites, such as the obligate intracellular pathogen *T. gondii*. CD73-generated extracellular adenosine is important for *T. gondii* survival because this pathogen can not to make its own [53]. Thus, CD73-deficient mice infected with *T. gondii* have lower parasite levels and are protected from chronic infection when compared with wild-type mice. Since mice lacking adenosine receptors have no effect in cyst formation, CD73 expression is thought to promote *T. gondii* differentiation and cyst formation by a mechanism dependent on adenosine generation, but independent of adenosine receptor signaling [53], suggesting that the parasite utilizes adenosine as a substrate for its own metabolism. Thus, adenosine effects on cells infected with intracellular pathogens depends on the specific type of pathogen and may depend on which adenosine receptor is stimulated [Fig. 1].

Because of the role of adenosine in facilitating survival of some microorganisms, some pathogens have evolved mechanisms to stimulate extracellular adenosine generation independently of the host. *Staphylococcus aureus* produces adenosine synthase A (AdsA) as a virulence factor, a cell wall-anchored enzyme which allows the bacteria to escape from phagocytic clearance and favoring the formation of organ abscesses [52]. Moreover, the same study showed that other prominent bacteria from the oral cavity including *Enterococcus faecalis* and *Streptococcus mutans* possess uncharacterized homologs of adenosine synthase.

Furthermore, some pathogens have evolved extracellular nucleotide-hydrolyzing enzymes that mimic the ecto-nucleotidases expressed in the host [Fig. 1]. For example, the surface of *Trypanosoma cruzi* expresses a Mg^{2+} -dependent ecto-ATPase activity, whose function is 20 times greater in trypomastigotes than in epimastigotes, suggesting a role for this enzyme in promoting infection in the vertebrate host [72]. A comparison of the Mg^{2+} -ecto-ATPase activities of the three forms of *T. cruzi* showed that the noninfective epimastigotes are less efficient at hydrolyzing eATP than the infective trypomastigote and amastigote stages [73]. Another parasite with Mg^{2+} -dependent ecto-ATPase is *L. amazonensis*, where the avirulent promastigotes are less efficient than the virulent promastigotes in hydrolyzing eATP, suggesting that virulent strains acquire adenosine and utilize it in its favor [74].

Interestingly, some bacteria secrete ATP during their growth [75], which may play a role in bacterial physiology. In the oral cavity, it was found that *Aggregatibacter actinomycetemcomitans* but not *P. gingivalis*, *Prevotella intermedia*, or *F. nucleatum* secrete ATP into the culture supernatant during its growth [76]. Infection in general can induce release of ATP from the host cell [77], and *P. gingivalis* infection stimulates ATP liberation after infection in GECs [63], THP-1 macrophages [78], and murine macrophages (unpublished data). It is tempting to speculate that eATP may be secreted during infection by some pathogens during their growth, such as the common bacteria *Escherichia coli*, *Staphylococcus*, *Acinetobacter*, and others; while supporting the survival of other pathogens

during infection – for example during infection by *L. amazonensis*, *T. gondii* and *T. cruzi*, due to their ability to hydrolyze eATP generating adenosine, which is necessary for their survival.

Inflammasomes and purinergic signaling associated with *P. gingivalis* infection

Inflammasomes, discovered in 2002, are multi-protein complexes assembled in the host cell in response to infection or cellular stress, leading to a type of cell death called pyroptosis and/or maturation and secretion of pro-inflammatory cytokines, such as IL-1 β and IL-18 [79–81]. Pyroptosis is a non-homeostatic and lytic cell death dependent on caspase-1 and/or caspase-11 activation [80,82,82]. P2X7 receptor was shown to activate NLRP3 inflammasomes [83]; and recently, caspase-11-induced pyroptosis was shown to require pannexin-1 channels and the P2X7 receptor [84]. This kind of cell death presents similarities to necrosis such as an increase in the cytoplasmic volume and rupture of the cellular plasma membrane. In the context of inflammasome activation, pro-inflammatory cytokines are important for eliminating pathogens [4,82]. Pyroptosis is important because of the cytokines, chemokines and DAMPs which are released to the extracellular compartment, and also because this type of cell death exposes intracellular bacteria to extracellular immune surveillance, thus allowing their destruction by antimicrobial peptides, immunoglobulins, and the complement system, and their uptake by immune cells [79]. IL-1 β affects virtually all cells and organs of the body and is one of the most important cytokines that mediate autoimmunity, infections and degenerative diseases [85]. This cytokine has a role in the central nervous system as an endogenous pyrogenic agent, and it can also induce inflammation, leukocyte recruitment, and Th17 profile immune responses [85,86]. Inflammasomes are usually studied in immune cells, but they can also be activated in several types of epithelial cells, including GECs [87].

Canonical inflammasomes convert procaspase-1 into the catalytically active enzyme, caspase-1, whereas a still completely undefined non-canonical inflammasome promotes activation of procaspase-11 [82,88]. Several canonical inflammasome complexes have been identified depending on the receptor that recognizes the PAMPs (for example, NLRP1, NLRP3, AIM2, NLRC4), while the non-canonical inflammasome can be activated by cytosolic LPS derived from Gram-negative bacteria. One of the best characterized inflammasomes contains the NLR member, NLRP3, the adaptor protein, apoptosis-associated speck-like protein containing a CARD (ASC), and the protease, caspase-1. The NLRP3 inflammasome can be activated by different stimuli, such as bacterial, viral, and fungal pathogens, pore-forming toxins, crystals, silica and DAMPs (for example, eATP) [82,89]. The activation of the canonical NLRP3 inflammasome typically requires two signals: (1) a PAMP, such as LPS, leading to transcription of NF- κ B, upregulating genes encoding pro-inflammatory cytokines, chemokines and proteins involved in the inflammasome platform; and (2) a DAMP, such as eATP, which induces inflammasome activation after ligation to the P2X7 receptor [39]. Once activated, these complexes promote activation of

the protease caspase-1, which cleaves pro-IL-1 β and pro-IL-18 into their active forms: IL-1 β and IL-18.

As reviewed elsewhere [79], inflammasome activation occurs in response to microbial invasion and is important for controlling infections. The murine NLRP1 inflammasome recognizes the cytosolic *Bacillus anthracis* lethal toxin, and mutations in the *Nlrp1b* gene confer susceptibility to anthrax lethal toxin-induced macrophage death. A defective NLRP3 inflammasome leads to mice susceptibility to *Candida albicans* and *Aspergillus fumigatus* infections [79]. Besides stimulating secretion of IL-1 β and IL-18, the induction of pyroptosis is a critical *in vivo* mechanism by which the NLRC4 inflammasome clears flagellin-expressing bacteria, such as *Legionella pneumophila* and *Burkholderia thailandensis*. Moreover, *Francisella tularensis* activates the AIM2 inflammasome, as shown by the increased susceptibility of caspase-1-deficient mice to infection with this pathogen [79].

IL-1 β regulates innate immune responses and is critical for host defense against bacterial infection. However, excessive IL-1 β production or inflammasome components [78], as well as increased P2X7 receptor and NLRP3 mRNA levels [90,91], are linked to periodontal disease in human gingival tissues. Recently, our group demonstrated that in murine macrophages, eATP-induced IL-1 β secretion is impaired by *P. gingivalis* fimbriae in a P2X7-dependent manner [92]. In human macrophages, *P. gingivalis* induces IL-1 β secretion and inflammatory cell death via caspase-1 activation (pyroptosis). Moreover, IL-1 β secretion and pyroptotic cell death requires both NLRP3 and AIM2 inflammasome activation by this oral pathogen. *P. gingivalis* infection induces ATP release from macrophages, which is mediated by NLRP3 inflammasome activation via P2X7 receptor stimulation and lysosomal damage [78]. In addition, GECs express the inflammasome components, NLRP3, NLRC4 and NLRP1 [93]. *P. gingivalis* stimulated expression of IL-1 β mRNA and intracellular accumulation of pro-IL-1 β , although IL-1 β secretion required on the addition of eATP *in vitro*. In GECs, eATP, but not *P. gingivalis* alone, induced caspase-1 activation [93]. Thus, *P. gingivalis* infection can provide the signals necessary for synthesis of pro-IL-1 β , but an exogenous danger signal, such as eATP, must activate the inflammasome, allowing the infected cell to secrete mature IL-1 β .

eATP induces ROS production through a complex consisting of the P2X4, P2X7 receptor and pannexin-1, and the P2X7-mediated ROS production can activate the NLRP3 inflammasome and caspase-1 [94]. Interestingly, *P. gingivalis* infection in GECs induces partially reduction of NLRP3 mRNA levels compared with uninfected GECs [93], suggesting that *P. gingivalis* can inhibit inflammasome components to promote its own survival. Consistent with the idea that *P. gingivalis* suppresses immune responses, *P. gingivalis* also suppresses inflammasome activation due to infection by another oral bacterium, *F. nucleatum*. This repression affects IL-1 β and IL-18 processing and cell death, in both human and murine macrophages. *F. nucleatum* activates IL-1 β secretion via the NLRP3 inflammasome, but when macrophages are co-infected with *F. nucleatum* and *P. gingivalis*, activation of the inflammasome and caspase-1, as well as IL-1 β secretion, are inhibited by *P. gingivalis* [25]. Since inflammasome activation is important for controlling infection, and *P. gingivalis*-induced inflammasome activation is linked to induction of periodontitis, it is still

unclear if deficiencies in inflammasome activation could favor *P. gingivalis* infection or, instead, impair periodontitis and alveolar bone loss induced by this periodontal pathogen.

Concluding remarks

Purinergic signaling can up- or down-modulate immune responses depending on the danger signals present, the purinergic receptor that is activated, and the cell type involved in the process [Fig. 1]. A large body of evidence demonstrates that purinergic signaling affects clearance or persistence of infection by *P. gingivalis* and others pathogens. P1 as well as P2 receptors modulate *P. gingivalis* infection as a function of the danger signal involved. Moreover, ecto-enzymes from the host cell or from the pathogens can modulate the course of infection by influencing the availability of nucleotides in the microenvironment. Finally, P2X7 receptor is involved in the activation of inflammasomes and its activation can control different infections. Because of the purinergic signaling can modulate different intracellular infections including those with the oral pathogen *P. gingivalis*, this field represents an important focus for future research regarding survival and elimination of different pathogens.

Conflicts of interest

All authors have declared that are no conflicts of interest.

Acknowledgments

This work was supported by funds from Conselho Nacional de Desenvolvimento Científico e Tecnológico do Brasil (CNPq - 311362/2014-1; 448152/2014-2), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior and Science Without Borders (CAPES - 038/2012), Programa de Núcleos de Excelência (PRONEX), Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro (FAPERJ - E-26/010.002985/2014). This study was approved and partially funded by Science Without Borders Federal Government Program to RCS and DMO and with a post-doctoral scholarship to ACM (2013/16113-9 and 2015/20954-4).

REFERENCES

- [1] Medzhitov R. Recognition of microorganisms and activation of the immune response. *Nature* 2007;449:819–26.
- [2] Kumar H, Kawai T, Akira S. Pathogen recognition by the innate immune system. *Int Rev Immunol* 2011;30:16–34.
- [3] Takeda K, Akira S. Toll-like receptors. *Curr Protoc Immunol* 2015;109:14.
- [4] Said-Sadier N, Ojcius DM. Alarmins, inflammasomes and immunity. *Biomed J* 2012;35:437–49.
- [5] Avila M, Ojcius DM, Yilmaz O. The oral microbiota: living with a permanent guest. *DNA Cell Biol* 2009;28:405–11.
- [6] Wade WG. The oral microbiome in health and disease. *Pharmacol Res* 2013;69:137–43.

- [7] Hajishengallis G. Periodontitis: from microbial immune subversion to systemic inflammation. *Nat Rev Immunol* 2015;15:30–44.
- [8] Hajishengallis G. Immunomicrobial pathogenesis of periodontitis: keystones, pathobionts, and host response. *Trends Immunol* 2014;35:3–11.
- [9] Socransky SS, Haffajee AD, Cugini MA, Smith C, Kent Jr RL. Microbial complexes in subgingival plaque. *J Clin Periodontol* 1998;25:134–44.
- [10] Socransky SS, Haffajee AD. Dental biofilms: difficult therapeutic targets. *Periodontol* 2000 2002;28:12–55.
- [11] Zambon JJ, Reynolds HS, Slots J. Black-pigmented *Bacteroides* spp. in the human oral cavity. *Infect Immun* 1981;32:198–203.
- [12] Griffen AL, Lyons SR, Becker MR, Moeschberger ML, Leys EJ. *Porphyromonas gingivalis* strain variability and periodontitis. *J Clin Microbiol* 1999;37:4028–33.
- [13] Igboin CO, Griffen AL, Leys EJ. *Porphyromonas gingivalis* strain diversity. *J Clin Microbiol* 2009;47:3073–81.
- [14] Griffen AL, Becker MR, Lyons SR, Moeschberger ML, Leys EJ. Prevalence of *Porphyromonas gingivalis* and periodontal health status. *J Clin Microbiol* 1998;36:3239–42.
- [15] Hajishengallis G, Liang S, Payne MA, Hashim A, Jotwani R, Eskin MA, et al. Low-abundance biofilm species orchestrates inflammatory periodontal disease through the commensal microbiota and complement. *Cell Host Microbe* 2011;10:497–506.
- [16] Kholy KE, Genco RJ, Van Dyke TE. Oral infections and cardiovascular disease. *Trends Endocrinol Metab* 2015;26:315–21.
- [17] Haraszthy VI, Zambon JJ, Trevisan M, Zeid M, Genco RJ. Identification of periodontal pathogens in atheromatous plaques. *J Periodontol* 2000;71:1554–60.
- [18] Lundberg K, Wegner N, Yucel-Lindberg T, Venables PJ. Periodontitis in RA—the citrullinated enolase connection. *Nat Rev Rheumatol* 2010;6:727–30.
- [19] de Aquino SG, Abdollahi-Roodsaz S, Koenders MI, van de Loo FA, Pruijn GJ, Marijnissen RJ, et al. Periodontal pathogens directly promote autoimmune experimental arthritis by inducing a TLR2- and IL-1-driven Th17 response. *J Immunol* 2014;192:4103–11.
- [20] Whitmore SE, Lamont RJ. Oral bacteria and cancer. *PLoS Pathog* 2014;10:e1003933.
- [21] Eskin MA, Hajishengallis G, Kinane DF. Differential activation of human gingival epithelial cells and monocytes by *Porphyromonas gingivalis* fimbriae. *Infect Immun* 2007;75:892–8.
- [22] Maekawa T, Krauss JL, Abe T, Jotwani R, Triantafidou M, Triantafidou K, et al. *Porphyromonas gingivalis* manipulates complement and TLR signaling to uncouple bacterial clearance from inflammation and promote dysbiosis. *Cell Host Microbe* 2014;15:768–78.
- [23] Morandini AC, Chaves Souza PP, Ramos-Junior ES, Souza Costa CA, Santos CF. MyD88 or TRAM knockdown regulates interleukin (IL)-6, IL-8, and CXCL12 mRNA expression in human gingival and periodontal ligament fibroblasts. *J Periodontol* 2013;84:1353–60.
- [24] Bao K, Belibasakis GN, Thurnheer T, Aduse-Opoku J, Curtis MA, Bostanci N. Role of *Porphyromonas gingivalis* gingipains in multi-species biofilm formation. *BMC Microbiol* 2014;14:258.
- [25] Taxman DJ, Swanson KV, Broglie PM, Wen H, Holley-Guthrie E, Huang MT, et al. *Porphyromonas gingivalis* mediates inflammasome repression in polymicrobial cultures through a novel mechanism involving reduced endocytosis. *J Biol Chem* 2012;287:32791–9.
- [26] Jauregui CE, Wang Q, Wright CJ, Takeuchi H, Uriarte SM, Lamont RJ. Suppression of T-cell chemokines by *Porphyromonas gingivalis*. *Infect Immun* 2013;81:2288–95.
- [27] Khalaf H, Bengtsson T. Altered T-cell responses by the periodontal pathogen *Porphyromonas gingivalis*. *PLoS One* 2012;7:e45192.
- [28] Guentsch A, Hirsch C, Pfister W, Vincents B, Abrahamson M, Sroka A, et al. Cleavage of IgG1 in gingival crevicular fluid is associated with the presence of *Porphyromonas gingivalis*. *J Periodontol Res* 2013;48:458–65.
- [29] Gaddis DE, Maynard CL, Weaver CT, Michalek SM, Katz J. Role of TLR2-dependent IL-10 production in the inhibition of the initial IFN-gamma T cell response to *Porphyromonas gingivalis*. *J Leukoc Biol* 2013;93:21–31.
- [30] Moutsopoulos NM, Kling HM, Angelov N, Jin W, Palmer RJ, Nares S, et al. *Porphyromonas gingivalis* promotes Th17 inducing pathways in chronic periodontitis. *J Autoimmun* 2012;39:294–303.
- [31] Wang L, Guan N, Jin Y, Lin X, Gao H. Subcutaneous vaccination with *Porphyromonas gingivalis* ameliorates periodontitis by modulating Th17/Treg imbalance in a murine model. *Int Immunopharmacol* 2015;25:65–73.
- [32] Burnstock G. Purine and pyrimidine receptors. *Cell Mol Life Sci* 2007;64:1471–83.
- [33] Burnstock G, Boeynaems JM. Purinergic signalling and immune cells. *Purinergic Signal* 2014;10:529–64.
- [34] Pellegatti P, Falzoni S, Pinton P, Rizzuto R, Di VF. A novel recombinant plasma membrane-targeted luciferase reveals a new pathway for ATP secretion. *Mol Biol Cell* 2005;16:3659–65.
- [35] Idzko M, Ferrari D, Eltzschig HK. Nucleotide signalling during inflammation. *Nature* 2014;509:310–7.
- [36] Trautmann A. Extracellular ATP in the immune system: more than just a “danger signal”. *Sci Signal* 2009;2:e6.
- [37] Hasko G, Linden J, Cronstein B, Pacher P. Adenosine receptors: therapeutic aspects for inflammatory and immune diseases. *Nat Rev Drug Discov* 2008;7:759–70.
- [38] Jacobson KA, Gao ZG. Adenosine receptors as therapeutic targets. *Nat Rev Drug Discov* 2006;5:247–64.
- [39] Coutinho-Silva R, Ojcius DM. Role of extracellular nucleotides in the immune response against intracellular bacteria and protozoan parasites. *Microbes Infect* 2012;14:1271–7.
- [40] Coutinho-Silva R, Monteiro da CC, Persechini PM, Ojcius DM. The role of P2 receptors in controlling infections by intracellular pathogens. *Purinergic Signal* 2007;3:83–90.
- [41] Morandini AC, Savio LE, Coutinho-Silva R. The role of P2X7 receptor in infectious inflammatory diseases and the influence of ectonucleotidases. *Biomed J* 2014;37:169–77.
- [42] Solle M, Labasi J, Perregaux DG, Stam E, Petrushova N, Koller BH, et al. Altered cytokine production in mice lacking P2X(7) receptors. *J Biol Chem* 2001;276:125–32.
- [43] Coutinho-Silva R, Persechini PM. P2Z purinoceptor-associated pores induced by extracellular ATP in macrophages and J774 cells. *Am J Physiol* 1997;273:C1793–800.
- [44] Kusner DJ, Adams J. ATP-induced killing of virulent *Mycobacterium tuberculosis* within human macrophages requires phospholipase D. *J Immunol* 2000;164:379–88.
- [45] Chaves MM, Marques-da-Silva C, Monteiro AP, Canetti C, Coutinho-Silva R. Leukotriene B4 modulates P2X7 receptor-mediated *Leishmania amazonensis* elimination in murine macrophages. *J Immunol* 2014;192:4765–73.
- [46] Chaves SP, Torres-Santos EC, Marques C, Figliuolo VR, Persechini PM, Coutinho-Silva R, et al. Modulation of P2X(7) purinergic receptor in macrophages by *Leishmania amazonensis* and its role in parasite elimination. *Microbes Infect* 2009;11:842–9.
- [47] Correa G, Marques da SC, de Abreu Moreira-Souza AC, Vommaro RC, Coutinho-Silva R. Activation of the P2X(7) receptor triggers the elimination of *Toxoplasma gondii*

- tachyzoites from infected macrophages. *Microbes Infect* 2010;12:497–504.
- [48] Coutinho-Silva R, Stahl L, Raymond MN, Jungas T, Verbeke P, Burnstock G, et al. Inhibition of chlamydial infectious activity due to P2X7R-dependent phospholipase D activation. *Immunity* 2003;19:403–22.
- [49] Kusner DJ, Barton JA. ATP stimulates human macrophages to kill intracellular virulent *Mycobacterium tuberculosis* via calcium-dependent phagosome-lysosome fusion. *J Immunol* 2001;167:3308–15.
- [50] Lammas DA, Stober C, Harvey CJ, Kendrick N, Panchalingam S, Kumararatne DS. ATP-induced killing of mycobacteria by human macrophages is mediated by purinergic P2Z(P2X7) receptors. *Immunity* 1997;7:433–44.
- [51] Lees MP, Fuller SJ, McLeod R, Boulter NR, Miller CM, Zakrzewski AM, et al. P2X7 receptor-mediated killing of an intracellular parasite, *Toxoplasma gondii*, by human and murine macrophages. *J Immunol* 2010;184:7040–6.
- [52] Thamavongsa V, Kern JW, Missiakas DM, Schneewind O. *Staphylococcus aureus* synthesizes adenosine to escape host immune responses. *J Exp Med* 2009;206:2417–27.
- [53] Mahamed DA, Mills JH, Egan CE, Denkers EY, Bynoe MS. CD73-generated adenosine facilitates *Toxoplasma gondii* differentiation to long-lived tissue cysts in the central nervous system. *Proc Natl Acad Sci USA* 2012;109:16312–7.
- [54] Spooner R, DeGuzman J, Lee KL, Yilmaz O. Danger signal adenosine via adenosine 2a receptor stimulates growth of *Porphyromonas gingivalis* in primary gingival epithelial cells. *Mol Oral Microbiol* 2014;29:67–78.
- [55] Sitkovsky MV, Ohta A. The 'danger' sensors that STOP the immune response: the A2 adenosine receptors? *Trends Immunol* 2005;26:299–304.
- [56] Thiel M, Caldwell CC, Sitkovsky MV. The critical role of adenosine A2A receptors in downregulation of inflammation and immunity in the pathogenesis of infectious diseases. *Microbes Infect* 2003;5:515–26.
- [57] Coutinho-Silva R, Correa G, Sater AA, Ojcius DM. The P2X(7) receptor and intracellular pathogens: a continuing struggle. *Purinergic Signal* 2009;5:197–204.
- [58] Pettengill MA, Marques-da-Silva C, Avila ML, d'Arc dos Santos OS, Lam VW, Ollawa I, et al. Reversible inhibition of *Chlamydia trachomatis* infection in epithelial cells due to stimulation of P2X(4) receptors. *Infect Immun* 2012;80:4232–8.
- [59] Marques-da-Silva C, Chaves MM, Chaves SP, Figliuolo VR, Meyer-Fernandes JR, Corte-Real S, et al. Infection with *Leishmania amazonensis* upregulates purinergic receptor expression and induces host-cell susceptibility to UTP-mediated apoptosis. *Cell Microbiol* 2011;13:1410–28.
- [60] Figliuolo VR, Chaves SP, Santoro GF, Coutinho CM, Meyer-Fernandes JR, Rossi-Bergmann B, et al. Periodate-oxidized ATP modulates macrophage functions during infection with *Leishmania amazonensis*. *Cytometry A* 2014;85:588–600.
- [61] Moreira-Souza AC, Marinho Y, Correa G, Santoro GF, Coutinho CM, Vommaro RC, et al. Pyrimidinergic receptor activation controls *Toxoplasma gondii* infection in macrophages. *PLoS One* 2015;10:e0133502.
- [62] Yilmaz O, Yao L, Maeda K, Rose TM, Lewis EL, Duman M, et al. ATP scavenging by the intracellular pathogen *Porphyromonas gingivalis* inhibits P2X7-mediated host-cell apoptosis. *Cell Microbiol* 2008;10:863–75.
- [63] Choi CH, Spooner R, DeGuzman J, Koutouzis T, Ojcius DM, Yilmaz O. *Porphyromonas gingivalis*-nucleoside-diphosphate-kinase inhibits ATP-induced reactive-oxygen-species via P2X7 receptor/NADPH-oxidase signalling and contributes to persistence. *Cell Microbiol* 2013;15:961–76.
- [64] Attwood PV, Wieland T. Nucleoside diphosphate kinase as protein histidine kinase. *Naunyn Schmiedeb Arch Pharmacol* 2015;388:153–60.
- [65] Dar HH, Chakraborti PK. Intermolecular phosphotransfer is crucial for efficient catalytic activity of nucleoside diphosphate kinase. *Biochem J* 2010;430:539–49.
- [66] Spooner R, Yilmaz O. Nucleoside-diphosphate-kinase: a pleiotropic effector in microbial colonization under interdisciplinary characterization. *Microbes Infect* 2012;14:228–37.
- [67] Sundin GW, Shankar S, Chugani SA, Chopade BA, Kavanaugh-Black A, Chakrabarty AM. Nucleoside diphosphate kinase from *Pseudomonas aeruginosa*: characterization of the gene and its role in cellular growth and exopolysaccharide alginate synthesis. *Mol Microbiol* 1996;20:965–79.
- [68] Johnson L, Atanasova KR, Bui PQ, Lee J, Hung SC, Yilmaz O, et al. *Porphyromonas gingivalis* attenuates ATP-mediated inflammasome activation and HMGB1 release through expression of a nucleoside-diphosphate kinase. *Microbes Infect* 2015;17:369–77.
- [69] Kim YJ, Lee JH, Lee Y, Jia J, Paek SH, Kim HB, et al. Nucleoside diphosphate kinase and flagellin from *Pseudomonas aeruginosa* induce interleukin 1 expression via the Akt/NF-kappaB signaling pathways. *Infect Immun* 2014;82:3252–60.
- [70] Alam MS, Costales MG, Cavanaugh C, Williams K. Extracellular adenosine generation in the regulation of pro-inflammatory responses and pathogen colonization. *Biomolecules* 2015;5:775–92.
- [71] Pettengill MA, Lam VW, Ojcius DM. The danger signal adenosine induces persistence of chlamydial infection through stimulation of A2b receptors. *PLoS One* 2009;4:e8299.
- [72] Bisaggio DF, Peres-Sampaio CE, Meyer-Fernandes JR, Souto-Padron T. Ecto-ATPase activity on the surface of *Trypanosoma cruzi* and its possible role in the parasite-host cell interaction. *Parasitol Res* 2003;91:273–82.
- [73] Meyer-Fernandes JR, Saad-Nehme J, Peres-Sampaio CE, Belmont-Firpo R, Bisaggio DF, Do Couto LC, et al. A Mg-dependent ecto-ATPase is increased in the infective stages of *Trypanosoma cruzi*. *Parasitol Res* 2004;93:41–50.
- [74] Berredo-Pinho M, Peres-Sampaio CE, Chrispim PP, Belmont-Firpo R, Lemos AP, Martiny A, et al. A Mg-dependent ecto-ATPase in *Leishmania amazonensis* and its possible role in adenosine acquisition and virulence. *Arch Biochem Biophys* 2001;391:16–24.
- [75] Mempin R, Tran H, Chen C, Gong H, Kim HK, Lu S. Release of extracellular ATP by bacteria during growth. *BMC Microbiol* 2013;13:301.
- [76] Ding Q, Quah SY, Tan KS. Secreted ATP from *Actinomyces comitans* triggers chemokine response. *Mol Oral Microbiol* 2015:1–12 [Epub ahead of print].
- [77] Sikora A, Liu J, Brosnan C, Buell G, Chessell I, Bloom BR. Cutting edge: purinergic signaling regulates radical-mediated bacterial killing mechanisms in macrophages through a P2X7-independent mechanism. *J Immunol* 1999;163:558–61.
- [78] Park E, Na HS, Song YR, Shin SY, Kim YM, Chung J. Activation of NLRP3 and AIM2 inflammasomes by *Porphyromonas gingivalis* infection. *Infect Immun* 2014;82:112–23.
- [79] Lamkanfi M, Dixit VM. Modulation of inflammasome pathways by bacterial and viral pathogens. *J Immunol* 2011;187:597–602.
- [80] Guo H, Callaway JB, Ting JP. Inflammasomes: mechanism of action, role in disease, and therapeutics. *Nat Med* 2015;21:677–87.
- [81] Martinon F, Burns K, Tschopp J. The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL-beta. *Mol Cell* 2002;10:417–26.
- [82] Lamkanfi M, Dixit VM. Mechanisms and functions of inflammasomes. *Cell* 2014;157:1013–22.

- [83] Mariathasan S, Weiss DS, Newton K, McBride J, O'Rourke K, Roose-Girma M, et al. Cryopyrin activates the inflammasome in response to toxins and ATP. *Nature* 2006;440:228–32.
- [84] Yang D, He Y, Munoz-Planillo R, Liu Q, Nunez G. Caspase-11 requires the Pannexin-1 channel and the purinergic P2X7 pore to mediate pyroptosis and endotoxic shock. *Immunity* 2015;43:923–32.
- [85] Garlanda C, Dinarello CA, Mantovani A. The interleukin-1 family: back to the future. *Immunity* 2013;39:1003–18.
- [86] Dinarello CA. Anti-inflammatory agents: present and future. *Cell* 2010;140:935–50.
- [87] Santana PT, Martel J, Lai HC, Perfettini JL, Kanellopoulos JM, Young JD, et al. Is the inflammasome relevant for epithelial cell function? *Microbes Infect* 2015;18(2):93–101.
- [88] Kayagaki N, Warming S, Lamkanfi M, Vande WL, Louie S, Dong J, et al. Non-canonical inflammasome activation targets caspase-11. *Nature* 2011;479:117–21.
- [89] Luna-Gomes T, Santana PT, Coutinho-Silva R. Silica-induced inflammasome activation in macrophages: role of ATP and P2X7 receptor. *Immunobiology* 2015;220:1101–6.
- [90] Bostanci N, Emingil G, Saygan B, Turkoglu O, Atilla G, Curtis MA, et al. Expression and regulation of the NALP3 inflammasome complex in periodontal diseases. *Clin Exp Immunol* 2009;157:415–22.
- [91] Ramos-Junior ES, Morandini AC, Almeida-da-Silva CL, Franco EJ, Potempa J, Nguyen KA, et al. A dual role for P2X7 receptor during *Porphyromonas gingivalis* infection. *J Dent Res* 2015;94:1233–42.
- [92] Morandini AC, Ramos-Junior ES, Potempa J, Nguyen KA, Oliveira AC, Bellio M, et al. *Porphyromonas gingivalis* fimbriae dampen P2X7-dependent interleukin-1beta secretion. *J Innate Immun* 2014;6:831–45.
- [93] Yilmaz O, Sater AA, Yao L, Koutouzis T, Pettengill M, Ojcius DM. ATP-dependent activation of an inflammasome in primary gingival epithelial cells infected by *Porphyromonas gingivalis*. *Cell Microbiol* 2010;12:188–98.
- [94] Hung SC, Choi CH, Said-Sadier N, Johnson L, Atanasova KR, Sellami H, et al. P2X4 assembles with P2X7 and pannexin-1 in gingival epithelial cells and modulates ATP-induced reactive oxygen species production and inflammasome activation. *PLoS One* 2013;8:e70210.