

UC Davis

UC Davis Previously Published Works

Title

How Can We Define “Optimal Microbiota?”: A Comparative Review of Structure and Functions of Microbiota of Animals, Fish, and Plants in Agriculture

Permalink

<https://escholarship.org/uc/item/0536p84c>

Authors

Ikeda-Ohtsubo, Wakako
Brugman, Sylvia
Warden, Craig H
[et al.](#)

Publication Date

2018

DOI

10.3389/fnut.2018.00090

Peer reviewed



How Can We Define “Optimal Microbiota?”: A Comparative Review of Structure and Functions of Microbiota of Animals, Fish, and Plants in Agriculture

Wakako Ikeda-Ohtsubo^{1*}, Sylvia Brugman², Craig H. Warden³, Johanna M. J. Rebel⁴, Gert Folkerts⁵ and Corné M. J. Pieterse⁶

¹ Laboratory of Animal Products Chemistry, Graduate School of Agricultural Science, Tohoku University, Sendai, Japan, ² Cell Biology and Immunology Group, Wageningen University and Research, Wageningen, Netherlands, ³ Departments of Pediatrics, Neurobiology Physiology and Behavior, University of California, Davis, Davis, CA, United States, ⁴ Wageningen Livestock Research, Wageningen University and Research, Wageningen, Netherlands, ⁵ Division of Pharmacology, Utrecht Institute for Pharmaceutical Sciences, Faculty of Science, Utrecht University, Utrecht, Netherlands, ⁶ Plant–Microbe Interactions, Department of Biology, Science4Life, Utrecht University, Utrecht, Netherlands

OPEN ACCESS

Edited by:

Raquel Hontecillas,
Virginia Tech, United States

Reviewed by:

Abdel Qawasmeh,
Hebron University, Palestine
Julio Villena,
Centro de Referencia para
Lactobacilos, Argentina

*Correspondence:

Wakako Ikeda-Ohtsubo
wakako.ohtsubo@tohoku.ac.jp

Specialty section:

This article was submitted to
Nutritional Immunology,
a section of the journal
Frontiers in Nutrition

Received: 09 June 2018

Accepted: 07 September 2018

Published: 02 October 2018

Citation:

Ikeda-Ohtsubo W, Brugman S, Warden CH, Rebel JMJ, Folkerts G and Pieterse CMJ (2018) How Can We Define “Optimal Microbiota?”: A Comparative Review of Structure and Functions of Microbiota of Animals, Fish, and Plants in Agriculture. *Front. Nutr.* 5:90. doi: 10.3389/fnut.2018.00090

All multicellular organisms benefit from their own microbiota, which play important roles in maintaining the host nutritional health and immunity. Recently, the number of studies on the microbiota of animals, fish, and plants of economic importance is rapidly expanding and there are increasing expectations that productivity and sustainability in agricultural management can be improved by microbiota manipulation. However, optimizing microbiota is still a challenging task because of the lack of knowledge on the dominant microorganisms or significant variations between microbiota, reflecting sampling biases, different agricultural management as well as breeding backgrounds. To offer a more generalized view on microbiota in agriculture, which can be used for defining criteria of “optimal microbiota” as the goal of manipulation, we summarize here current knowledge on microbiota on animals, fish, and plants with emphasis on bacterial community structure and metabolic functions, and how microbiota can be affected by domestication, conventional agricultural practices, and use of antimicrobial agents. Finally, we discuss future tasks for defining “optimal microbiota,” which can improve host growth, nutrition, and immunity and reduce the use of antimicrobial agents in agriculture.

Keywords: microbiota, agriculture, animal husbandry, aquaculture, rhizosphere, phyllosphere, agricultural immunology

INTRODUCTION

Today, biologists in agricultural science, regardless of the organism of their interest, focus significant attention on the microbiota, i.e., the complex communities of microorganisms colonizing host animals, fish, and plants (1). Meta-analyses of 16S rRNA genes from different body parts of animals, fish, and plants are frequently performed expecting that some changes of microbiota will explain the effectiveness of treatments such as feed changes, fertilizer amendment, or gene knockouts on host organisms, which have been conducted with aims to improve productivity and sustainability in agriculture (2). However, it is often the case that no apparent

changes are observed in the microbial structure corresponding to the specific treatment, or if present, the functions of the responding microorganisms are not well-known [e.g., (3–5)]. Besides, it is often difficult for researchers in agricultural sciences to exploit the microbial data to improve the host factors because of the lack of definition and criteria of “optimal microbiota” in animals, fish, and plants.

Compared to a large body of studies on microbiota of human subjects (6) or experimental models using rodents(7), zebrafish (8), or *Arabidopsis* (9), there are a very limited number of studies on economically important animals, fish, and plants. Microbiota datasets obtained from livestock animals, aquaculture fish, and crop plants grown may significantly be affected by complex environmental factors such as climates, cultivation scales, and uses of antibiotics and fertilizers, which can vary between different countries and regions. Besides, the microbiota of agricultural organisms may also reflect the great variability of host species and genotypes, biological functions at different developmental stages, and macro- and microstructures of the colonizing sites, which are not thoroughly studied as the laboratory models. Due to the overall limited understanding of the microbiota in agricultural ecosystems at this point, it is not an easy task to define “optimal microbiota,” which can optimize the growth, host nutrition, and immunity of agricultural organisms.

The importance of understanding the structure and functions of microbiota in agriculture is also widely discussed in the context of the spread of antimicrobial resistance (AMR) from agricultural sites to human society (10). While manipulation of microbiota is a promising strategy to tackle the AMR (11), it is prerequisite for researchers to interpret and exploit the rapidly expanding datasets of the microbiota in animals, fish, and plants in agriculture with a more generalized view. By sharing knowledge on the ecophysiology of microbiota in different host organisms with respect to their structure and metabolites and understanding how the host factors and ambient conditions can alter them, we would be able to refine targets of microbial manipulation and reduce uses of chemicals and antimicrobial agents in agricultural fields.

The aim of this review is to summarize and generalize the current knowledge on the microbiota on animals, fish, and plants in agriculture with emphasis on structure and functions of bacterial communities, which may contribute to the health of the host organisms and can strongly be impacted by agricultural practices such as uses of antimicrobial agents. We finally provide important yet overlooked aspects of microbiota in animals, fish, and plants in agriculture, which should be considered in future studies to reach the goal of defining the “optimal microbiota.”

STRUCTURE AND FUNCTION OF MICROBIOTA OF ANIMALS, FISH, AND PLANTS

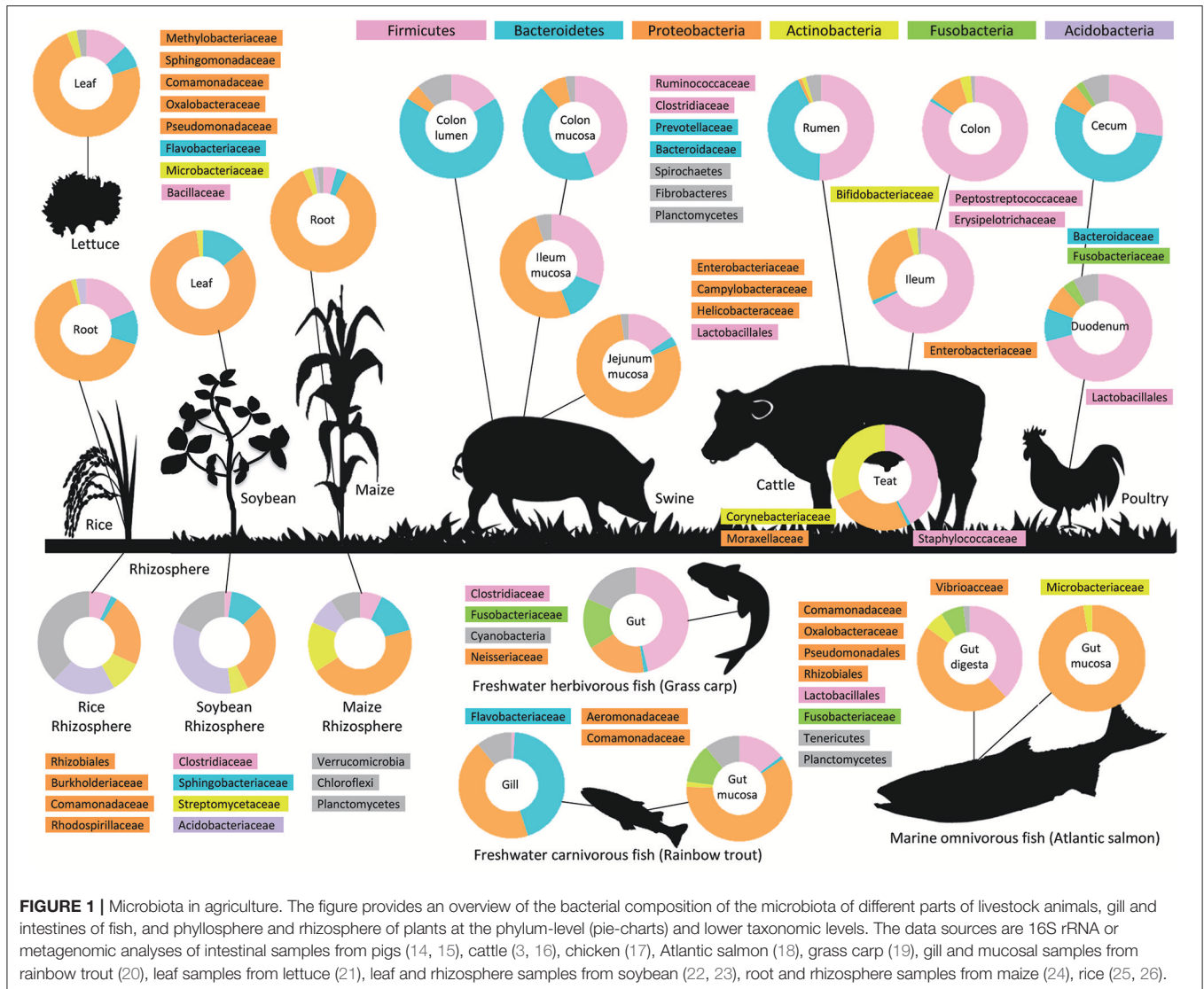
General Overview of Microbiota of Animals, Fish, and Plants

The body of organisms provides a wide variety of ecological niche, in which the environmental conditions such as

temperature, pH, and oxygen level as well as nutrition availability affect the composition of microbiota residing there. While archaea and eukaryotic microorganisms such as fungi and protozoa account for a significant proportion of microbiota in the plant rhizosphere (12) and cow rumen (13), bacterial communities have been primarily focused on in many studies in agricultural science in terms of their functional contribution to host nutrition and health. The 16S rRNA gene-based approach with a next-generation sequencing platform has revealed diversity and dynamics of bacterial communities colonizing animals, fish, and plants in agriculture, which have enabled us to grasp a general overview of compositional similarities and differences of microbiota among these organisms (**Figure 1**). Microbiota of animal, fish, and plants are highly diverse and can harbor up to 20 bacterial phyla, but it is a common trait that three phyla: Proteobacteria, Firmicutes, and Bacteroidetes, dominate the bacterial community (**Figure 1**). Less abundant phyla include Actinobacteria, which are commonly found but variable at lower taxonomic levels (e.g., Streptomycetaceae, Microbacteriaceae, and Corynebacteriaceae), while Fusobacteria and Acidobacteria are more specified to animal/fish and plants, respectively (**Figure 1**). Fusobacteria can represent a major bacterial group of “core gut microbiome” of some marine and freshwater fish (8, 27). Chloroflexi, Cyanobacteria, Planctomycetes, Spirochaetes, and Verrucomicrobia sporadically occur as subdominant phyla (**Figure 1**).

The high abundance of Proteobacteria in animals, fish, and plants (**Figure 1**) reflects the advantages of facultative anaerobes in the host proximity, where strict anaerobes are exposed to the risk of oxygen toxicity but strict aerobes may face a severe competition over oxygen as an electron acceptor. Such oxic-anoxic interface is ubiquitous as microenvironments in and around the host organism and is an important determinant of the composition of microbiota (28). Facultative anaerobic bacteria have highly flexible metabolic properties; they are able to generate energy by fermentation or use inorganic nitrogen compounds such as nitrate as an alternative electron acceptor when oxygen is depleted from the environment. Under oxic conditions, they grow rapidly using oxygen and break down and build up a wide variety of organic compounds, which essentially change the surrounding organochemical conditions (29). Such exceptional adaptability to multiple environmental conditions, which have been characterized by their high genetic and phenotypic plasticity, enable Proteobacteria to be specialists of host association, as represented by major target symbionts and pathogens in agriculture (30–32).

While Proteobacteria are ubiquitous and their association is often described to be opportunistic, they show apparent host specificities in some microbiota. In fish intestinal microbiota, Aeromonadaceae (Gammaproteobacteria) represents the most abundant symbionts in freshwater fish, while Vibrionaceae (Gammaproteobacteria) replace the occupation in marine fish (27, 33). In microbiota of livestock animals, the proteobacterial community is predominated by Enterobacteriaceae, followed by Campylobacteriaceae and



Helicobacteraceae, which are a major source of foodborne diseases of human (3, 34, 35).

Plant phyllosphere has been found to be dominated by strict aerobes, represented by Methylobacteriaceae and Sphingomonadaceae (Alphaproteobacteria), but their abundance is low in the plant root, being replaced by a large diversity of facultative anaerobes (36). Interestingly, microbiota of plants and fish have several groups of facultative anaerobes in common. Multiple apertures of fish, i.e., skin, gills, and gut, are constantly in contact with ambient water, each of which is covered with thick mucus biofilm, which bears resemblance to the plant root system. Comamonadaceae and Oxalobacteraceae (Betaproteobacteria) as well as Flavobacteriaceae (Bacteroidetes), which dominate the leaf-to-root microbiota of plants, are also abundant in fish mucus, especially in gill microbiota [Figure 1; (36, 37)]. Also, bacteria known as plant growth-promoting microbes (PGPM) such as Pseudomonadaceae (Gammaproteobacteria) and Rhizobiales (Alphaproteobacteria) are also frequent colonizers in fish

intestinal microbiota (33, 38). In contrast, these bacterial groups are not commonly found in animal intestinal microbiota.

Firmicutes population in animals, fish, and plants can be roughly classified into two groups: Lactic acid bacteria from the orders Bacillales and Lactobacillales, and anaerobic fermentative bacteria affiliated with Clostridiales such as Clostridiaceae and Ruminococcaceae. The former represents microbiota in oxic-to microoxic regions like plant phyllosphere, fish mucosa, and small intestines of animals, while the latter represents anoxic fermentative digestive tracts like rumen and large intestine (Figure 1). Lactobacillales is one of the most frequently found and most studied bacterial groups in animal and fish microbiota with some family-level variations, as Lactobacillaceae is a stable and important colonizer in the small intestine of pigs (39, 40) and chickens (17), Carnobacteriaceae [rainbow trout; (20)], and Leuconostocaceae [Atlantic salmon; (18)] are characteristic in fish microbiota, and Enterococcaceae and Streptococcaceae are generally found both in animals

and fish microbiota (41). Clostridiaceae is phylogenetically and functionally diverse and widely distributed in anaerobic environments from plant rhizosphere to animal and fish intestines (42). Studies on *Clostridium* spp. from various origins have shown their metabolic versatility and dexterous switching of their fermentation pathway in response to environmental changes (43, 44).

Bacteroidetes colonizing animals, fish, and plants can also be affiliated with two types: aerobic Flavobacteriaceae, which is adapted to the oxic interface of plants and fish as mentioned above, and anaerobic fermentative bacteria such as Bacteroidaceae and Prevotellaceae (Figure 1). Flavobacteriaceae is often recognized as threatening pathogens of animals and fish (45), but it also represents PGPM and has been introduced to industrial applications (46). Bacteroidaceae and Prevotellaceae are important primary fermenters in animal and fish intestinal tracts, through which complex carbohydrates derived from plants and undigested proteins enter the microbial metabolic network and provide soluble sugars and amino acids available for all type of cells (47). They dominate the rumen and colonic microbiota of animals (48, 49) but are rarely found in fish and plants, which suggests their speciation to animal digestive tracts.

Relationship Between Intestinal Microbiota Composition and Host Nutrition of Animals and Fish

Microbiota of animals and fish associated with different host physiological conditions have been widely studied to elucidate the relationship between the structure and functions of microbiota and host nutritional health. Although results are variable across studies, which may be attributed to different experimental designs, analytical methods, or individual variations, a few general aspects can be inferred from recent studies.

In animal husbandry, feed efficiency and growth performance are often focused on as the most important physiological factors. Recent studies on pig microbiota have reported the enrichment of Clostridiales, as well as microbial functional genes involved in fermenting dietary polysaccharides and amino acid metabolism, are positively associated with porcine feed efficiency (50, 51). Similarly, positive correlations of Clostridiales (family Lachnospiraceae) to good feed efficiency have also been found in the cattle rumen (49) and chicken caeca (52). In fish, the intestinal microbiota of prebiotics-treated fish with improved growth performance also showed an increased number of *Clostridium* spp. (53). All of these studies have attributed the positive effects of Clostridiales on the host feed efficiency and growth performance to the high energy yields by the production of short-chain fatty acids (SCFA) such as butyrate, which has also been suggested in human gut microbiota studies (54).

Lactobacillales are thought to improve feed efficiency of animals and multiple *Lactobacillus* strains are widely used as feed additives especially in pig farming (55). While studies have suggested a positive correlation of *Lactobacillus* to a better feed

efficiency of cattle, chickens, and fish (17, 49, 53), contrasting effects of *Lactobacillus* spp. on growth performance have been also reported on chicken (17, 52, 56). Therefore, species- and strain-level variation should be considered when the abundance of *Lactobacillus* strains is used as a criterion for evaluating the health and growth performance of animals and fish.

Both in animals and fish, intestinal microbial colonization has been shown to promote epithelial cell turnover and regulate transcription of genes involved in nutrient metabolism and immunity, and the corresponding gene modules are universally conserved between mammals and fish (57–59).

Rhizosphere vs. Phyllosphere: Difference of Microbiota Composition

Plant microbiota significantly differed from those of animals and fish, in that in addition to the complex bacterial community, a large variety of archaea as well as eukaryotic macro- and microorganisms can directly and constantly affect the health of the host plant (12, 60). In response to this challenge, plants have a finely regulated immunological capacity, which recognizes different exogenous molecules and responds by activating specific defense mechanisms (61). The plant rhizosphere is home of a high density [10^{10} - 10^{12} cells per gram soil; (62)] of microorganisms and a large pool of microbial metabolites influence the nutritional conditions of the host plant as well as the composition of microbial populations (63). In contrast, microbiota in the plant phyllosphere, i.e., leaf and root, are enriched with restricted groups of bacteria. As mentioned already, Proteobacteria and Flavobacteria have been found as endophytes or epiphytes of the host plant, while any other bacterial phyla dominating the rhizosphere, such as the phylum Acidobacteria and Firmicutes, or prominent rhizobacteria such as Streptomycetaceae (Actinobacteria) and Burkholderiaceae (Betaproteobacteria) are segregated at different levels of the phyllosphere (64). This suggests the existence of a “selecting gate” between the rhizosphere and the phyllosphere, or different compartments of the plant root (65).

Studies across different plant species including the *Arabidopsis* model indicate that the enrichment of Proteobacteria is a common trait in the plant phyllosphere (Figure 1). However, the enriched bacterial species, i.e., members selected by the host plant, seem to differ significantly between plant species. Pseudomonadaceae (Gammaproteobacteria) and Streptomycetaceae (Actinobacteria), which are frequent colonizers in the root of *Arabidopsis*, are not found in grass plants such as barley, maize, and wheat (24, 66). Also, leaf microbiota between plant species can be very different, as exemplified that Enterobacteriaceae, Bacillaceae, and *Pantoea* spp. dominating spinach and lettuce leaves are not abundant in *Arabidopsis* (21, 67). While the bacterial composition of microbiota of the phyllosphere of economically important plants is very limited compared to that of the rhizosphere (36, 68), the composition of microbiota in different plant compartment may provide useful insights into site-specific selection mechanisms of the host plant.

Acquisition of Microbiota in the Early Life of Animals, Fish, and Plants

In animals, the intestinal immunity is known to be developed in the course of frequent interaction with microbiota, which are formed and fluctuated in response to the host dynamics (69). It has been shown that the early-life transfer of microbiota from the mother to the child via the birth channel and colostrum milk can impact on subsequent intestinal microbial diversity and immune processes in piglets (70). The transition from nursing, weaning to conventional diets can dramatically affect intestinal microbiota. Milk provides immunological factors such as Immunoglobulin A (IgA), leukocytes, and peptides, which suppress inflammatory cytokine expression, and lactose and oligosaccharides contained in milk can stimulate the growth of early-colonizing microorganisms such as lactic acid bacteria. (71). Comparative analyses of the intestinal microbiota of nursing and weaning piglets have shown that the dietary change from sow milk to a starter diet composed of plant and animal-based components has a significant impact on the microbial structure as well as its functional capacities (34). In their study, Enterobacteriaceae, Bacteroidaceae, and Clostridiaceae dominating the nursing piglets almost disappeared as the piglet diet shifted to a starter diet, which has been characterized by the dominance of Prevotellaceae and Ruminococcaceae associated with plant polysaccharide degradation. The early colonization and subsequent disappearance of Enterobacteriaceae as well as the maturation of microbiota associated with the domination of plant-polysaccharide degraders in the early life have also been commonly found in other mammals including human (72) and also in chickens (73). Proliferation of pathogenic members of Enterobacteriaceae can be regulated by selective binding activities of host-derived IgA, which seems to be one of the most important mechanisms affecting early development of intestinal microbiota in animals (74).

Fish develop from eggs that are directly exposed to microorganisms in their surroundings. The eggs are quickly coated with microorganisms present in the surrounding water, of which some have been shown to protect the eggs from infection with oomycete *Saprolegnia*, a deleterious pathogen causing economic loss in the salmon industry (75). Since fish represents the largest number of vertebrates (>28,000 species), a lot of interspecies variation may exist based on the receptors or binding moieties on the egg surface. Although microbiota of fish larvae is poorly understood (76, 77), some studies have suggested that the microbiota composition of fish larva greatly depend on the microorganisms present on the eggs, in the live feed and rearing water (78, 79). Since microorganisms are able to enter the fish larvae before it starts feeding (3–4 days after fertilization), initial microbial infection in the larval intestine probably occurs before the feed specific species grow to abundance [(76, 80) Lopez Nadal, unpublished observations]. In early life stages of Coho salmon, *Pseudomonas* sp. present on the eggs has been predominantly found in the juvenile gastrointestinal tract, but not in the culture water or food, which suggests that a maternal transfer may occur in the early developmental stages of the salmon (81).

The development of seedlings from largely sterile plant seeds is one of the most critical stages of a plant's life cycle. Yet, very

little is known about the role of microbiota in the early life of plants (82). Starting inoculum on the ripening seed may be important for the establishment of microbiota and preliminary enrichment of the soil microbiota by the parental plant will form ideal environments for germination of seeds in the same soil. Interestingly, it has recently been shown that diseased plants can recruit themselves a consortium of beneficial, immunostimulatory microbes from the soil environment and let them colonize germinating seedlings, which suggests that plants are capable of selecting soil microbiota for protecting a successive generation of plants against the causal agent of the disease (83, 84).

Post-translational Host Modulation by Microbiota

Host epigenomics has recently been shown to be one of the most important factors significantly affected by microbiota. Anti-inflammatory effects of some intestinal microorganisms such as *Clostridium* spp. have been attributed to their metabolite butyrate, an epigenetic substance known to inhibit activities of histone deacetylases and modulate gene expression patterns of host animals (85). Also in fish, promotion of resistance to viral infection of conventionally reared zebrafish has been shown to be associated with microbe-induced epigenetic changes in the host (86). In plants, not only bacterial pathogens but also fungal and other eukaryotic organisms have been known to manipulate their host epigenetically to favor themselves (87). While low-molecular-weight microbial metabolites such as SCFAs and polyamines from mammalian intestinal microbiota have been shown to be involved in various epigenomic mechanisms in the mammalian host (88), modulatory effects of microbial structural components such as LPS, peptidoglycan, and exopolysaccharides from microbiota in most agricultural organisms have not yet been well-studied except for those from some pathogens and probiotics (see below). Molecular mechanisms how microbiota modulates host epigenomics have recently been attracting major attention, which may also contribute to understanding functions of microbiota in animals, fish, and plants in agriculture (89).

MICROBIAL METABOLITES: BENEFICIAL AND DELETERIOUS EFFECTS OF METABOLITES PRODUCED BY MICROBIOTA OF ANIMALS, FISH, AND PLANTS

“Optimal microbiota” of agricultural organisms are expected to provide beneficial effects on their host nutritional health and immunological resistance. Microorganisms influence the host health by producing a large variety of metabolites, which can have both beneficial effects and detrimental effects on the host physiology (Table 1).

Short Chain Fatty Acid (SCFA)

Fermentative microorganisms break down carbohydrates and proteins into SCFA. In the gut of animals and fish, major SCFAs produced by fermentative microorganisms are acetate,

TABLE 1 | Important microbial metabolites and their effects on host animals, fish, and plants.

Microbial metabolites	Examples	Hosts ^a	Beneficial effects	Detrimental effects
Short-chain fatty acids (SCFAs)	Butyrate	A, F	Energy homeostasis anti-inflammatory effect, improve intestinal barrier	Mucosal disruption
	Propionate			Neurotoxicity
Organic acids	Lactate	A, F,	Increase butyrate production	Acidosis, inflammation, neurotoxicity,
	Succinate	A, F, P	Glycemic control, feed PGPM, mineral solubilization	Feed pathogens
Ammonia and amino acid derivatives	Ammonia, ammonium	A, F, P	Nitrogen nutritional source, pH neutralization	Inflammation, mucosal damages, increase oxidative stress
	Biogenic amines	A, F	Synthesis of neurotransmitter (serotonin)	Production of uremic toxins, carcinogenesis
Signaling molecules acting on the host	IAA, 2,4-DAPG, GABA	A, F, P	Growth promotion, anti-inflammation	
Signaling molecules acting on other microbes	AHL, AI-2		Maintenance of microbial structure (e.g., biofilm formation), cell-to-cell communication between microbes	
Antimicrobial compounds	Bacteriocins, RiPPs	A, F, P	Defense against pathogens, immunomodulatory effects	Cytotoxicity
Vitamins	Vitamin B ₁₂ , vitamin K, D	A, F	Provisioning of host nutrition, immunomodulation	
Microbial cellular components	LPS, Polysaccharide A	A, F, P	Immunomodulation, maintenance of intestinal homeostasis	Inflammation

^aA, animals; F, fish; P, plants.

propionate, and butyrate, while relatively low amounts of formate, valerate, caproate, and branched-chain SCFAs, i.e., isobutyrate, 2-methyl-butyrate, and isovalerate, which are used as a marker of undesired intestinal protein fermentation (90), are also present (91, 92). SCFAs can modulate the gene expression of the host epithelial cells in multiple ways and their physiological concentrations may significantly affect the host nutritional health and immunity (93, 94). SCFAs produced by gut microbiota are known to serve as a major energy source for ruminant animals, which consume cellulose fibers and complex carbohydrates as the main diet, but also play a crucial role for young monogastric animals for maintaining the body weight after weaning (95, 96). Additional roles of SCFA include defense mechanisms, mineral solubilization, and the anti-inflammatory effects (97, 98). SCFA produced by intestinal microbiota improve intestinal barrier functions and suppress inflammation through signaling pathways such as activating G-protein coupled receptors, inhibiting histone deacetylase, stimulation of histone acetyltransferase activity, and stabilizing hypoxia-inducible factor (HIF), which have been extensively studied with rodent models (99, 100).

While SCFAs produced by intestinal microbiota are generally considered to be beneficial to the host, excessive SCFAs can cause intestinal injuries in animals with premature or weakening mucosal conditions (101, 102). Formate, which concentration increases along with dysbiosis, can enhance the growth of unwanted enterobacterial pathogens (103). High levels

of propionate are often found in human and animals with psychological and behavioral disorders and thought to have a neurotoxic potential (104, 105).

Lactate and Other Organic Acids

Lactate is an important intermediate in anaerobic fermentation of carbohydrates. While host-derived lactate has been known for regulatory functions on the energy homeostasis and brain metabolism (106, 107), lactate produced by microbiota may also play important roles in the intestinal ecosystem, such as turnover of host epithelial cells (108), in addition to their role as a major food source for other SCFA producing bacteria (109). In the small intestine of animals and fish, lactic acid bacteria such as Lactobacillales (**Figure 1**) are known to produce lactate as a primary metabolite, while *Turicibacter* (Erysipelotrichaceae) represent the major lactate producers in the large intestine. Residuous oxygen may increase relative abundance of intestinal lactic acid bacteria, which generally show high tolerance against oxygen (110, 111), and lactate productions and consumption profiles may differ significantly between upper and lower intestines (112).

Succinate is another major organic acid released from microbiota during carbohydrate fermentation. Prevotellaceae and Veillonellaceae, which are predominant bacterial groups in the rumen and in the colon of pigs, are major succinate producers. A large variety of bacteria including Enterobacteriaceae and Clostridiaceae can grow on succinate, and succinate accumulation would increase a risk of infection

by pathogenic bacteria (113). Recent studies have reported succinate production by gut microbiota is strongly correlated to the metabolic fluctuation of host animals (114, 115).

Accumulation of lactate and succinate has been reported in the intestine of pigs with gastric problems (116, 117), which has been shown to be inversely related to the SCFA concentrations (118). Increased concentrations of lactate and succinate can cause a decline in pH and drastic changes in metabolic patterns in animal and fish intestines, which can lead to deleterious outcomes such as acidosis and inflammation (119). To avoid this, a rapid turnover of lactate by gut microbiota seems to be crucial for intestinal homeostasis in animals and fish (28).

Plant root exudates contain a high amount of organic acids, such as citrate, succinate, and malate, which can significantly affect the composition of the microbial community in the rhizosphere (120, 121). The high amount of organic acids exudated from the host plant feed and control proximal microbiota consisting of plant growth-promoting microorganisms (PGPM) as well as pathogens, and the microbiota in rhizosphere may also affect the concentration of organic acids excreted from the plant host by modulating their regulatory genes (122, 123). As organic acids can affect the growth and plant-promoting activities of PGPM, e.g., suppressing phosphate stabilization (124), the concentration of organic acids should be well fine-tuned by the host-microbe regulatory network (125).

Ammonia and Amino Acid Derivatives

Ammonia (NH_3) and ammonium (NH_4^+) play an important physiological role in the body of animals, fish, and plants as it provides usable forms of nitrogen required for the synthesis of DNA, RNA, and proteins. Ammonia not only serves as a major nitrogen source but also are responsible for buffering the ecosystem such as rumen by neutralizing excess acids. Many bacteria are able to generate ammonia via protein or peptide degradation and N_2 fixation. Fixed atmospheric N_2 in the NH_4^+ form is an important source of nitrogen in the soil ecosystem, which concentration in agricultural soils is approximately between 20 and 200 μM (126), and many plants are highly dependent on endophytic or rhizospheric nitrogen-fixing bacteria for their nitrogen demands. Ammonia is also an important metabolite in the microbiota of animals and fish and millimolar level concentration of ammonia can be generally found in the intestinal ecosystem [e.g., 10–70 mM in colonic lumen; (127)]. Many bacteria such as *E. coli* and *Bacteroides* spp. are known to require ammonia or ammonium for their growth in the intestinal system, while they are able to provide amino acids and their derivatives to other intestinal bacteria and the host (29, 128).

Toxicity of ammonia (NH_3) and ammonium (NH_4^+) from microbiota poses a risk to the host as well. When excess protein is present in the intestine, ammonia production by microbial deamination will exceed microbial ammonia assimilation (129). Urea produced by the host animals is also converted to ammonia and further to ammonium hydroxide by microbiota, which can elevate luminal pH at the level of causing mucosal damage and irritation (130). Accumulated ammonia has multiple adverse

effects on host epithelial cells (129). Ammonium toxicity is also documented in plants, but the cause for this phenomenon and involvement of microbiota is still unknown (131).

Increased protein and peptide concentrations in a microbial ecosystem may facilitate active amino acid conversion to various nitrogenous derivatives. Many facultative and obligate anaerobic bacteria ferment amino acids into a wide variety of intermediate metabolites such as indoles, phenols, cresols, and their derivatives as well as biogenic amines (132). Biogenic amines such as tyramine, putrescine, histamine, methylamine, and tryptamine, are produced by decarboxylation of amino acids, which have significant physiological and toxicological functions in eukaryotic cells (132, 133). Biogenic amines serve as precursors of various bioactive compounds, which can directly regulate physiology and behavior of the host. For example, tryptamine, a β -arylamine neurotransmitter derived from tryptophan metabolism, influences modes of serotonin production in enterochromaffin (EC) cells and therefore affect host behavior (134, 135).

Secondary Metabolites

Secondary metabolites from microbiota such as tryptamine, which can serve as hormones or signaling molecules (136) to “control” the host physiology and behavior, are also known for plants. Indole-3-acetic acid (IAA), one of the most important plant growth regulators, is also derived from the tryptophan metabolism of PGPM such as Pseudomonadaceae (137). Pseudomonadaceae are also known to produce a wide variety of secondary metabolites including antibiotic compounds and siderophores, which can protect the host plant from invasive pathogens not only in the rhizosphere but also in phyllosphere (138, 139). Specific secondary metabolites of Pseudomonadaceae such as 2,4-diacetylphloroglucinol (2,4-DAPG) are of special interests for controlling specific plant-microbe interaction (140). In animals and fish, secondary metabolites produced by gut microbiota such as gamma-aminobutyric acid (GABA) are likely to have more general but significant influence on physiological and psychological properties of the host (141).

Antimicrobial compounds such as bacteriocins, siderophores, and lipopeptide biosurfactants enable some microorganisms to outcompete and eliminate pathogens and shape the structure of microbiota by also affecting the host immunity (139, 142, 143). Although bacteriocins and siderophores have been well-documented in some beneficial or pathogenic strains, genes encoding these compounds could be commonly found in a wide range of microorganisms (144). In human microbiota, ribosomally synthesized post-translationally modified peptides (RiPPs), which include lantibiotics, thiazole/oxazole-modified microcins (TOMMs) as well as thiopeptides antibiotics, are one of the most widely distributed and variable microbial metabolites (145).

Signaling molecules known as autoinducers play important roles in cell-to-cell communication between microorganisms and shape the synchronized behavior of microbial community such as biofilm formation (146). In contrast to the well-known quorum-sensing molecule AHL (N-acyl homoserine

lactone), which are produced as virulence factors by many gram-negative pathogenic bacteria and probably uncommon in healthy intestinal microbiota in animals and fish (147, 148), AI-2 (autoinducer 2) are present in many intestinal bacteria such as Firmicutes and Bacteroidetes and known to modify the structure and behavior of intestinal microbiota (149, 150).

Vitamins

Animals depend on their gut microbiota for various vitamins, which are often deficient in their normal diet. Deficiencies in vitamin B₁₂ and other B-complex vitamins, as well as vitamin K and D in animals and fish has been correlated to the absence of intestinal microorganisms producing those vitamins (151, 152). In addition to the crucial role for the host nutritional health (153–155), vitamins formed by microbiota are also provisioned to other microorganisms in proximity thereby supporting the cross-feeding metabolic network in gut microbiota (152). Some vitamins are also known to participate in host epigenomic mechanisms by altering the transcriptional machinery of the host cells (88). While most plants can synthesize vitamins and do not depend on their microbiota for their vitamin requirements, some algae have been known to benefit from the microbial provision of vitamin B₁₂ (156).

Microbial Cellular Components

Microbial metabolites affecting host health also include structural compounds of microorganisms themselves. Exo- and lipopolysaccharides (LPS), peptidoglycan, flagellin, and some unique peptides and nucleic acids released from the microbial community, which are often collectively called as microorganism-associated molecular patterns (MAMPs), are specifically detected as “non-self” and distinguished by pattern recognition receptors (PRRs) of the host cells and trigger immune responses in animals, fish (86), and plants (157, 158). It has long been recognized that MAMPs from pathogens play a crucial role for host immunity in animals and plant (96, 159–161), but recent studies have revealed that MAMPs from commensal microbiota may also control the host immune system to maintain intestinal homeostasis (162). Common intestinal residential bacteria such as *Clostridium* and *Bacteroides* have been shown to stimulate the production of cytokines such as IL-6 and TNF α that protect intestinal tissues from injury (163), and also to induce the proliferation of immune cells such as FOXP3⁺ regulatory T (Treg) cells (164). Although MAMPs required for induction of each host factor are not well-understood, species-specific polysaccharides such as Polysaccharide A found on the capsule of *Bacteroides fragilis* may play important roles for initial binding and recognition to the host cells (165).

In the model plant *Arabidopsis thaliana* it was recently shown that MAMPs from beneficial root microbiota members are similarly recognized by the plant immune system as MAMPs from pathogens, but the downstream immune response was suppressed by so far unknown mechanisms (89).

INFLUENCE OF AGRICULTURAL MANAGEMENT PRACTICES ON MICROBIOTA IN ANIMALS, FISH, AND PLANTS

Recent comparative studies on gut microbiota between urban and hunter-gatherer human population have suggested continuous decreases in microbial diversity over generations during worldwide industrialization (166–168). Similar changes, i.e., loss of diversity in domestic vs. wild counterparts, have been documented in primates and Przewalski's horses (169, 170), but the diversity level of gut microbiota has been found to be consistent in mice (171) and a vice versa situation has also been observed in cloacal microbiota in parrots (172). Nevertheless, many studies have shown that the reduced diversity of gut microbiota is characteristic to many diseases and disorders in human [e.g., (173, 174)], therefore the loss of diversity in gut microbiota over generations may have negatively affected the health of not only human but also other animals and fish. Although reduced microbial diversity is not often discussed for plants, the long-term agricultural practices may have served as strong selective pressures on the microbiota of the phyllosphere and rhizosphere (175, 176). Supplementation of the “lost” population could improve host fitness, as has been shown in mice (177), but the cause of the loss of certain microbial groups and its consequences are not fully understood. For optimizing microbiota in agricultural organisms, it is important to evaluate how domestication and agricultural management practices can affect the microbiota and host nutritional health and immunity.

Domestication

While some livestock animals and farmed fish have evolved into domestic species distinct from wild relatives, studies on the microbiota of wild representatives of agricultural organisms provide insights into how domestication may have affected the microbial composition of agricultural organisms. For example, a comparative study of domestic pig microbiota with that of wild boars has revealed that *Lactobacillus* spp. and Enterobacteriaceae, which are considered to be dominant bacterial groups in pig intestinal microbiota (178), are not common in wild boars (179). Interestingly, recently domesticated wild boars have been found to harbor Enterobacteriaceae as a major group, which collectively suggests that gut microbiota of domestic pigs may reflect the recent agricultural management practices (179). Since the increased abundance of Enterobacteriaceae has been reported to be correlated to post-weaning diarrhea (180), agricultural management is likely to have a significant impact on the health of domestic pigs via the fluctuation of gut microbiota. In cattle, inoculation with bison rumen contents has been shown to increase protein digestibility and nitrogen retention but not fiber digestibility, which suggests that microbiota of ancestors of livestock animals may have had higher capacities to extract nitrogen nutrition from crude materials (181).

While a study on gut microbiota of laboratory-reared and recently-caught zebrafish has shown little influence from domestication on intestinal microbiota of fish and shrimps (8,

182), some bacterial groups in wild fish have been found to disappear upon captivity (183) and therefore careful investigation should be needed in future studies.

Plant microbiota can also be affected by domestication, i.e., plant breeding in combination with yield-increasing agricultural practices and the use of chemical fertilizers and pesticides, which has resulted in the selection of specific plant traits maximizing profitable functions from the root microbiome (184). Studies have shown distinct features of the microbial community associated with wild and domesticated crop species such as rhizosphere microbiota from sugar beets (185) and endophytic population from grapevines (186). Nevertheless, plant hosts respond to various microbial factors by changing their physiology and thereby can modulate their microbial composition (187), it is important to obtain more insights into the physiological and structural differences between wild and domesticated plant species.

Agricultural Management Practices

Agricultural management practices include multiple and long-term stress factors such as selective breeding, confinement, nutritional changes, close contact with people, and antimicrobial usage, all of which can affect the composition of microbiota to a greater extent.

Selective breeding produces a new type of organism with a phenotype different from its parental organisms, which can affect the composition of the host-specific microbiota. Gut microbiota in livestock animals including cattle and pigs have been reported to show a host-specificity and habitability over generations (188, 189), which suggests that the host genetics are correlated to microbial structure and functions (190). Although fish gut microbiota are largely affected by ecological factors, several studies have shown host selection plays an important role in shaping intestinal and gill microbiota (191, 192). In plants, rhizosphere microbiota has been shown to have specific profiles unique to its host plant species, genotype, and cultivar (193, 194).

Confinement such as indoor breeding and aquaculture has been reported to affect microbiota of animals and fish to various extents. There is no clear evidence on how housing systems (indoor vs. outdoor) can affect microbiota of animals, since previously reported changes in gut microbiota in response to different housing methods can be better explained by dietary changes (195, 196). As intestinal tracts of fish are constantly exposed to water and a large number of microorganisms in their surroundings, it is not surprising that the conditions of aquaculture such as water quality (e.g., salinity) and external microbial community significantly affect gut microbiota of fish (197–199).

Nutritional changes including grazing, feeding, and weaning in animals or fertilizer amendment for plants, are one of the most important factors shaping the structure and functions of microbiota in agriculture. Availability of microbiota-accessible organic compounds is a crucial determinant for the survivability of individual microorganisms in the host systems (200). In livestock animals, starch grains, plant fibers and crude proteins in feed are digested by rumen or colonic microbiota to different degrees, which can essentially

change the structure and functions of gut microbiota and the host nutritional health as already mentioned above (51, 201, 202). Weaning can cause serious fluctuation of rumen and intestinal microbiota of young animals (34, 203), which can occasionally lead to dysbiosis and post-weaning diarrhea (204). Also similar to animals, dietary changes such as feeding high-cellulose diet has been shown to increase cellulolytic bacteria in the fish intestine (205, 206), and relationships between the dietary components and gut microbiota are extensively studied (207). Continuous cropping and fertilizer amendment can modulate nutritional status in the agricultural soil and affect plant microbiota (208). The growth inhibition caused by the continuous cropping, such as the imbalance of inorganic nutrients and prevalence of pathogenic fungi, could be mitigated by native microbiota (209), which suggests the resilience of agricultural soils highly depend on their microbial activities.

Continuous close contact with human and animals seems to allow inter-species transmission of certain bacterial groups even between the intestinal microbiota. As the microbial composition in human gut has been found to be affected by adjacent livestock animals or companion animals (210, 211), the microbiota of animals, fish, and plants could be affected by the human microbial assemblage. The predominance of *Bifidobacterium* spp. in the modern human gut microbiota reflecting the dietary transition from fiber-rich plant-based diet to western diet (212), has also been observed in animals, which have experienced close contact with humans (213).

Influence of Usage of Antimicrobial Agents on Microbiota in Agriculture

Antimicrobial agents have been used as a common agricultural practice over the decades, which aims not only to treat infectious diseases of animals and fish but also to promote growth and improve productivity (214). The worldwide overuse of antimicrobial agents has already brought major concerns: the spread of AMR in microorganisms through the global ecosystem. Microbiota of animals, fish, and plants, which have been treated with antimicrobial agents, can serve as a reservoir of resistance genes where commensal microorganisms may confer AMR to pathogenic microorganisms by horizontal transfer events (215). Use of low-dose antibiotics as antimicrobial growth promoters (AGPs) in livestock farming is still a common practice in many countries, which poses a great risk to accelerate emergence and spread of antibiotic-resistant bacteria (216).

It has been shown that antimicrobial agents can alter intestinal microbiota of animals in location-specific ways, (i) structural and functional disruption of foregut microbiota, and (ii) increase of AMR in hindgut microbiota (217). The major risk of disruption of foregut microbiota in piglet is characterized by the increased number of *Streptococcus suis* and Enterobacteriaceae, which are known to cause infectious diseases like pneumonia and post-weaning diarrhea (218). Such effects by the early-life exposure to antimicrobial agents can be retained throughout the life of animals (70, 219). Reduced diversity of intestinal microbiota by antimicrobial treatments, which can increase the

host susceptibility to pathogens, has also been documented in fish (220).

The risk of AMR in agriculture is not restricted to livestock farms. Livestock manure is frequently used for composting and eventually amended to agricultural soils as fertilizers and the high frequency of AMR in manure can be transferred to the microbial community in the soil, which can also affect the plant microbiota (221, 222). Aquaculture ponds are considered to be a significant reservoir of AMR (223), especially in countries where livestock manure is used for feeding fish in farming ponds (224). Once entering to the agricultural food chain, AMR is transmitted and exchanged between microbiota associated with animals, fish, and plants and spread over agricultural food products, which can be eventually introduced to human microbiota (225–227).

FUTURE TASKS FOR DEFINING “OPTIMAL MICROBIOTA” OF ANIMALS, FISH, AND PLANTS

The cross-sectional view of the microbiota of animals, fish, and plants reviewed here may provide an idea of what aspects should be particularly considered in the future investigation for elucidating structure and functions of “optimal microbiota” and applying the knowledge for improving host nutrition and immunology to maximize productivity and sustainability in agriculture.

Quantitative Understanding of Microbiota

Many past studies employ 16S rRNA gene sequencing approach to study microbiota of animals, fish, and plants. While the composition of microbiota, which can be estimated by the number of sequencing reads, allows understanding of the diversity and distribution of specific microbial groups, the density of microorganisms is often overlooked. Microbial density is especially important when studying a specific host region where microbial activities play crucial roles or the host immunological factors respond to a certain density threshold. The microbiota of animal rumen, hindguts or rhizosphere with the extremely high density of microorganisms [10^{11} - 10^{12} cells per mL or gram; (62, 228)] and animal foreguts or the plant phyllosphere with a smaller number of microbial cells in several orders of magnitude (10^4 - 10^7 cells) should be considered as a separate ecosystem themselves, i.e., habitats with different types and levels of microbial structure and functions, which could be differently recognized by the host immunity (162). Therefore, using fecal samples for studying gut microbiota should be done with a special caution, since the foregut microbiota are highly underrepresented in the feces and its compositional and functional changes can be completely masked by the hindgut microbiota (229).

While no direct counts of the number of microbial metabolites produced are available for any host organism, as their composition constantly changes depending on environmental, host and microbial factors, attempts have been made to use sequencing information to estimate the number of compounds that may be produced from human microbiota. Donia et al. have

identified 14,000 predicted small-molecule biosynthetic gene clusters (BGCs) by shotgun sequencing human gut metagenome where they have shown that 3,118 BGCs have been found in the healthy human microbiota, among which 599 clusters can be affiliated with typical human gut microbiota while 1,061 clusters with the typical oral cavity (90). They reported that gene cluster classes in the human microbiota differed from those in non-human microbiota, which suggests that species-specific analyses of BGCs will also be useful for agricultural organisms. To make the best use of such useful approach, it is worth summarizing what kind of microbial metabolites can occur and how they affect physiology and growth properties of the host animals, fish, and plants.

Cultivation of the Uncultured Majority

The limitation of our current knowledge on microbiota of animals, fish, and plants can be largely attributed to the predominance of uncultured microorganisms in each microbiota. For example, a study by Stanley et al. has been able to identify several bacterial phylotypes in chicken caeca, which are negatively correlated with growth performance of the host chicken, but all of these phylotypes have been affiliated with unknown and uncultured bacterial groups of the phylum Firmicutes (35). In cattle, 44.6% of all microbial sequences obtained from gastrointestinal tracts have failed to be identified at the genus level (3). Similarly, eggs at the fertilization stage of grass carp have been reported to be colonized by a large proportion (>50%) of uncultured bacteria (230). In maize rhizosphere, important functional genes for microbial nitrogen metabolism such as nitrogen-fixation and denitrification have been mainly affiliated with uncultured bacteria (231).

These findings underscore the importance of cultivation of the uncultured members of microbiota colonizing animals, fish, and plants. The difficulties of conventional cultivation techniques are now able to be addressed by modern technologies featured by single-cell (meta)genomics (232) and culturomics (233), which combine the analytical methods such as the index fluorescence-activated cell sorting (FACS) or the matrix-assisted laser desorption/ionization-time of flight (MALDI-TOF) with multiple culture conditions and high-throughput 16S rRNA gene sequencing (233, 234). Isotope probing (SIP) (235) has also been used as a powerful method to identify uncultured microorganisms with specific activities such as host-protein utilization in animal gut microbiota (236) or the pesticide degradation in the rhizosphere microbiota (237). Individual profiles of microbial metabolites (Table 1) are also an important aspect to understand the role of uncultured microorganisms, which can be extensively assessed by recently advancing metabolomic approach integrated with genomic and proteomic datasets (238).

Systematic Investigation of Microbial Functions

Many well-described Proteobacteria species, which are widely distributed in healthy animals, fish and plants, also behave as opportunistic pathogens (239, 240). *Campylobacter* spp. and *Salmonella* spp., two major food-borne pathobionts have

been found to be stable colonizers of livestock animals and human, which are usually harmless but occasionally cause diseases of the hosts (241). Plant-associated bacteria of the class Gammaproteobacteria such as Pseudomonadaceae, Erwiniaceae, Xanthomonadaceae show species- and strain-level differences in their traits as pathogens, antagonists of the pathogens, or PGPM (240). Most microorganisms consisting the microbiota of animals, fish, and plants seem to be opportunistic symbionts, which colonization can result in beneficial and detrimental, or no effects on the host, which might be determined not only by genetic properties of each microorganism but also by various environmental conditions and host factors (242, 243). The question whether the identified bacterial groups are beneficial or detrimental cannot be answered only by 16S rRNA gene-based analyses, and the microbial metabolites (244), host-specific selective marker genes including virulence or symbiotic factors in the microbial genomes (145, 245), and metabolites involved in modulations of the host cell immunity (246) should be systematically investigated for elucidating the roles of microbiota. A study on the fruit fly *Drosophila* has shown that occurrence of a single protein of plant pathogenic bacterium *Erwinia carotovora*, i.e., *evf* factor determines the successfulness of persistence in the gut of the host (247). Similar unknown mechanisms may present in microorganisms associated with animals, fish, and plants, which are responsible for the host-specific selection of individual microorganisms.

As the majority is still uncultured, microbial physiology is not fully resolved and many important microbial processes in natural ecosystems have still not well-discussed in microbiota research on animals, fish, and plants.

Nitrogen fixation, as well as ammonia oxidation and denitrification (reduction of nitrate, nitrite, and N_2O), are globally important processes conducted by microorganisms fueling the nitrogen cycle of most ecological systems (248) but have been poorly investigated for animal and fish microbiota. Nitrogen fixation and other inorganic nitrogen conversion have been known to maintain nutritional status of termite gut microbiota, where nitrogen-poor wood polysaccharides (cellulose and hemicellulose) serve as major sources of nutrition (249, 250). Recent findings of a genetic diversity of the nitrogen fixation gene *nifH* in human microbiota indicate that inorganic nitrogen metabolism may play an important role in animal microbiota (251), but it is still unknown which microorganisms are responsible for the processes.

Microbial removal of hydrogen (H_2) generated in the course of fermentation of fiber-rich carbohydrates is a critical process in every anaerobic system, including gut microbiota of animals and fish (252). H_2 -consuming intestinal microorganisms such as methanogenic archaea, sulfate-reducing bacteria, and reductive acetogens are therefore as important as primary fermenters such as Bacteroidetes for maintaining the redox balance and conserving energy (253), and are crucial for the stable SCFA production in the ecosystem (254, 255). In contrast to the well-studied rumen microbiota, little is known about H_2 -consuming microorganisms for monogastric animal guts, but a study by Rey et al. have shown that genes encoding Wood-Ljungdahl pathway, which are key components for reductive acetogenesis, have been

shown to be highly represented among expressed RNAs in human gut microbiota than marker genes for methanogenesis or sulfate reduction (256).

Clarification of Optimization Purposes

Recently, Lloyd-Price et al. has suggested “healthy human microbiome” can be defined in terms of microbial composition, function, dynamics, and ecology (6). Although this definition can be applied for defining “optimal microbiota” of animals, fish, and plants in agriculture, the dataset from each target organism may be highly limited compared with that of human gut microbiome (257) and additional criteria should be considered in the context of productivity and sustainability.

In plant science, improving growth speeds, conferring resistance against environmental stresses, or improving nutritional values have been successfully accomplished by inoculating PGPM consisting of specific bacterial groups or amending materials promoting the growth of PGPM (258). In contrast, the impact of inoculation of putatively beneficial microorganisms i.e., probiotics to animals and fish seems to be less pronounced (259). Striking similarities between gut microbiota in antibiotic-treated pigs, which gain weight and have high-feed efficiency, and gut microbiota linked to human obesity (218, 260) indicate that the “optimal microbiota” are not necessarily identical to the “healthy microbiota” in agricultural contexts. Therefore, clarifying purposes of the microbiota optimization, i.e., prevention of specific diseases or addition of nutritional values in the products, is prerequisite for the employment of microbiota manipulation techniques, which have been reviewed by Brugman et al., in this issue (11). As discussed above, the worldwide threat of AMR should be combated by reducing the amount of unnecessary use of antimicrobial agents in agricultural practice and by manipulation of microbiota, which can minimize the risk of diseases and optimize the growth performance of target organisms. Host–microbe interaction in individual agricultural organisms should be studied with close reference to the current knowledge available from laboratory models and humans, through which new ideas for modulating microbiota as alternative strategies to antibiotic use can be shared and discussed interdisciplinarily.

AUTHOR CONTRIBUTIONS

WI-O and CP conceived the idea of the review, WI-O wrote the manuscript, and all authors discussed the contents and contributed to the writing of the manuscript.

ACKNOWLEDGMENTS

This review is based on the workshop The Dawn of Agricultural Immunology hosted and supported by the Lorentz Center in Leiden, the Netherlands, during September 19–23, 2016. The authors would like to thank the Lorentz Center as well as the organizers and participants of the workshop for providing us with the opportunity to discuss the ideas in this review with the aim to integrate knowledge of microbiota of animals, fish, and plants in agriculture.

REFERENCES

- Hacquard S, Garrido-Oter R, González A, Spaepen S, Ackermann G, Lebeis S, et al. Microbiota and host nutrition across plant and animal kingdoms. *Cell Host Microbe* (2015) 17:603–16. doi: 10.1016/j.chom.2015.04.009
- Mueller UG, Sachs JL. Engineering microbiomes to improve plant and animal health. *Trends Microbiol.* (2015) 23:606–17. doi: 10.1016/j.tim.2015.07.009
- Mao S, Zhang M, Liu J, Zhu W. Characterising the bacterial microbiota across the gastrointestinal tracts of dairy cattle: membership and potential function. *Sci Rep.* (2015) 5:16116. doi: 10.1038/srep16116
- Thibodeau A, Letellier A, Yergeau É, Larrivière-Gauthier G, Fravallo P. Lack of evidence that selenium-yeast improves chicken health and modulates the caecal microbiota in the context of colonization by *Campylobacter jejuni*. *Front Microbiol.* (2017) 8:451. doi: 10.3389/fmicb.2017.00451
- Wemheuer F, Kaiser K, Karlovsky P, Daniel R, Vidal S, Wemheuer B. Bacterial endophyte communities of three agricultural important grass species differ in their response towards management regimes. *Sci Rep.* (2017) 7:40914. doi: 10.1038/srep40914
- Lloyd-Price J, Abu-Ali G, Huttenhower C. The healthy human microbiome. *Genome Med.* (2016) 8:51. doi: 10.1186/s13073-016-0307-y
- Xiao L, Feng Q, Liang S, Sonne SB, Xia Z, Qiu X, et al. A catalog of the mouse gut metagenome. *Nat Biotechnol.* (2015) 33:1103. doi: 10.1038/nbt.3353
- Roeselers G, Mitge EK, Stephens WZ, Parichy DM, Cavanaugh CM, Guillemin K, et al. Evidence for a core gut microbiota in the zebrafish. *ISME J.* (2011) 5:1595–608. doi: 10.1038/ismej.2011.38
- Bai Y, Müller DB, Srinivas G, Garrido-Oter R, Potthoff E, Rott M, et al. Functional overlap of the *Arabidopsis* leaf and root microbiota. *Nature* (2015) 528:364. doi: 10.1038/nature16192
- Thanner S, Drissner D, Walsh F. Antimicrobial resistance in agriculture. *mBio* (2016) 7:e02227-15. doi:10.1128/mBio.02227-15
- Brugman S, Ikeda-Ohtsubo W, Braber S, Folkerts G, Pieterse CMJ, Bakker PAHM. A comparative review on microbiota manipulation: lessons from fish, plants, livestock and human research. *Front Nutr.* 5:80. doi: 10.3389/fnut.2018.00080
- Buée M, De Boer W, Martin F, van Overbeek L, Jurkevitch E. The rhizosphere zoo: an overview of plant-associated communities of microorganisms, including phages, bacteria, archaea, and fungi, and of some of their structuring factors. *Plant Soil.* (2009) 321:189–212. doi: 10.1007/s11104-009-9991-3
- Henderson G, Cox F, Ganesh S, Jonker A, Young W, Collaborators GRC, et al. Rumen microbial community composition varies with diet and host, but a core microbiome is found across a wide geographical range. *Sci Rep.* (2015) 5:14567. doi: 10.1038/srep14567
- Burrough ER, Arruda BL, Plummer PJ. Comparison of the luminal and mucosa-associated microbiota in the colon of pigs with and without swine dysentery. *Front Vet Sci.* (2017) 4:139. doi: 10.3389/fvets.2017.00139
- Kelly J, Daly K, Moran AW, Ryan S, Bravo D, Shirazi-Beechey SP. Composition and diversity of mucosa-associated microbiota along the entire length of the pig gastrointestinal tract; dietary influences. *Environ Microbiol.* (2017) 19:1425–38. doi: 10.1111/1462-2920.13619
- Braem G, De Vliegher S, Verbist B, Heyndrickx M, Leroy F, De Vuyst L. Culture-independent exploration of the teat apex microbiota of dairy cows reveals a wide bacterial species diversity. *Vet Microbiol.* (2012) 157:383–90. doi: 10.1016/j.vetmic.2011.12.031
- Yan W, Sun C, Yuan J, Yang N. Gut metagenomic analysis reveals prominent roles of *Lactobacillus* and cecal microbiota in chicken feed efficiency. *Sci Rep.* (2017) 7:45308. doi: 10.1038/srep45308
- Gajardo K, Rodiles A, Kortner TM, Krogdahl Å, Bakke AM, Merrifield DL, et al. A high-resolution map of the gut microbiota in Atlantic salmon (*Salmo salar*): a basis for comparative gut microbial research. *Sci Rep.* (2016) 6:30893. doi: 10.1038/srep30893
- Ni J, Yan Q, Yu Y, Zhang T. Factors influencing the grass carp gut microbiome and its effect on metabolism. *FEMS Microbiol Ecol.* (2014) 87:704–14. doi: 10.1111/1574-6941.12256
- Lyons P, Turnbull J, Dawson K, Crumlish M. Exploring the microbial diversity of the distal intestinal lumen and mucosa of farmed rainbow trout *Oncorhynchus mykiss* (Walbaum) using next generation sequencing (NGS). *Aquacult Res.* (2017) 48:77–91. doi: 10.1111/are.12863
- Rastogi G, Sbodio A, Tech JJ, Suslow TV, Coaker GL, Leveau JHJ. Leaf microbiota in an agroecosystem: spatiotemporal variation in bacterial community composition on field-grown lettuce. *ISME J.* (2012) 6:1812–22. doi: 10.1038/ismej.2012.32
- Delmotte N, Knief C, Chaffron S, Innerebner G, Roschitzki B, Schlapbach R, et al. Community proteogenomics reveals insights into the physiology of phyllosphere bacteria. *Proc Natl Acad Sci USA.* (2009) 106:16428–33. doi: 10.1073/pnas.0905240106
- Liang J, Sun S, Ji J, Wu H, Meng F, Zhang M, et al. Comparison of the rhizosphere bacterial communities of Zigongdongdou soybean and a high-methionine transgenic line of this cultivar. *PLoS ONE* (2014) 9:e103343. doi: 10.1371/journal.pone.0103343
- Niu B, Paulson JN, Zheng X, Kolter R. Simplified and representative bacterial community of maize roots. *Proc Natl Acad Sci USA* (2017) 114:E2450–9. doi: 10.1073/pnas.1616148114
- Lu Y, Rosencrantz D, Liesack W, Conrad R. Structure and activity of bacterial community inhabiting rice roots and the rhizosphere. *Environ Microbiol.* (2006) 8:1351–60. doi: 10.1111/j.1462-2920.2006.01028.x
- Breidenbach B, Conrad R. Seasonal dynamics of bacterial and archaeal methanogenic communities in flooded rice fields and effect of drainage. *Front Microbiol.* (2014) 5:752. doi: 10.3389/fmicb.2014.00752
- Tarnecki AM, Burgos FA, Ray CL, Arias CR. Fish intestinal microbiome: diversity and symbiosis unravelled by metagenomics. *J Appl Microbiol.* (2017) 123:2–17. doi: 10.1111/jam.13415
- Brune A, Frenzel P, Cypionka H. Life at theoxic-anoxic interface: microbial activities and adaptations. *FEMS Microbiol Rev.* (2000) 24:691–710. doi: 10.1111/j.1574-6976.2000.tb00567.x
- Bennett BD, Kimball EH, Gao M, Osterhout R, Van Dien SJ, Rabinowitz JD. Absolute metabolite concentrations and implied enzyme active site occupancy in *Escherichia coli*. *Nat Chem Biol.* (2009) 5:593–9. doi: 10.1038/nchembio.186
- Uden G, Becker S, Bongaerts J, Holighaus G, Schirawski J, Six S. O₂-sensing and O₂-dependent gene regulation in facultatively anaerobic bacteria. *Arch Microbiol.* (1995) 164:81–90. doi: 10.1007/s002030050238
- Dobrindt U, Hochhut B, Hentschel U, Hacker J. Genomic islands in pathogenic and environmental microorganisms. *Nat Rev Microbiol.* (2004) 2:414. doi: 10.1038/nrmicro884
- Silby MW, Winstanley C, Godfrey SAC, Levy SB, Jackson RW. *Pseudomonas* genomes: diverse and adaptable. *FEMS Microbiol Rev.* (2011) 35:652–80. doi: 10.1111/j.1574-6976.2011.00269.x
- Sullam KE, Essinger SD, Lozupone CA, O'Connor MP, Rosen GL, Knight R, et al. Environmental and ecological factors that shape the gut bacterial communities of fish: a meta-analysis. *Mol Ecol.* (2012) 21:3363–78. doi: 10.1111/j.1365-294X.2012.05552.x
- Frese SA, Parker K, Calvert CC, Mills DA. Diet shapes the gut microbiome of pigs during nursing and weaning. *Microbiome* (2015) 3:28. doi: 10.1186/s40168-015-0091-8
- Stanley D, Hughes RJ, Moore RJ. Microbiota of the chicken gastrointestinal tract: influence on health, productivity and disease. *Appl Microbiol Biotechnol.* (2014) 98:4301–10. doi: 10.1007/s00253-014-5646-2
- Müller DB, Vogel C, Bai Y, Vorholt JA. The plant microbiota: systems-level insights and perspectives. *Annu Rev Genet.* (2016) 50:211–34. doi: 10.1146/annurev-genet-120215-034952
- Lowrey L, Woodhams DC, Tacchi L, Salinas I. Topographical mapping of the rainbow trout (*Oncorhynchus mykiss*) microbiome reveals a diverse bacterial community with antifungal properties in the skin. *Appl Environ Microbiol.* (2015) 81:6915–25. doi: 10.1128/AEM.01826-15
- Estruch G, Collado MC, Peñaranda DS, Tomás Vidal A, Jover Cerdá M, Pérez Martínez G, et al. Impact of fishmeal replacement in diets for gilthead sea bream (*Sparus aurata*) on the gastrointestinal microbiota determined by pyrosequencing the 16s rRNA gene. *PLoS ONE* (2015) 10:e0136389. doi: 10.1371/journal.pone.0136389
- Levesque CL, Hooda S, Swanson KS, de Lange K. Alterations in ileal mucosa bacteria related to diet complexity and growth performance in young pigs. *PLoS ONE* (2014) 9:e108472. doi: 10.1371/journal.pone.0108472

40. Valeriano VD, Balolong MP, Kang D-K. Probiotic roles of *Lactobacillus* sp. in swine: insights from gut microbiota. *J Appl Microbiol.* (2017) 122:554–67. doi: 10.1111/jam.13364
41. Hardie JM, Whiley RA. Classification and overview of the genera *Streptococcus* and *Enterococcus*. *J Appl Microbiol.* (2003) 83:1S–11S. doi: 10.1046/j.1365-2672.83.s1.1.x
42. Wiegel J, Tanner R, Rainey FA. An introduction to the family Clostridiaceae BT. In: Dworkin M, Falkow S, Rosenberg E, Schleifer K-H, Stackebrandt E, editors. *The Prokaryotes, Vol. 4, Bacteria: Firmicutes, Cyanobacteria*. New York, NY: Springer US (2006). pp. 654–78. doi: 10.1007/0-387-30744-3_20
43. Grupe H, Gottschalk G. Physiological events in *Clostridium acetobutylicum* during the shift from acidogenesis to solventogenesis in continuous culture and presentation of a model for shift induction. *Appl Environ Microbiol.* (1992) 58:3896–902.
44. Fischbach MA, Sonnenburg JL. Eating for two: how metabolism establishes interspecies interactions in the gut. *Cell Host Microbe* (2011) 10:336–47. doi: 10.1016/j.chom.2011.10.002
45. Loch TP, Faisal M. Emerging flavobacterial infections in fish: a review. *J Adv Res.* (2015) 6:283–300. doi: 10.1016/j.jare.2014.10.009
46. Nishioka T, Elsharkawy MM, Suga H, Kageyama K, Hyakumachi M, Shimizu M. Development of culture medium for the isolation of *Flavobacterium* and *Chryseobacterium* from rhizosphere soil. *Microb Environ.* (2016) 31:104–10. doi: 10.1264/jisme2.ME15144
47. El Kaoutari A, Armougom F, Gordon JI, Raoult D, Henrissat B. The abundance and variety of carbohydrate-active enzymes in the human gut microbiota. *Nat Rev Microbiol.* (2013) 11:497. doi: 10.1038/nrmicro3050
48. Holman DB, Brunelle BW, Trachsel J, Allen HK. Meta-analysis to define a core microbiota in the swine gut. *mSystems* (2017) 2:e00004-17. doi: 10.1128/mSystems.00004-17
49. Li F, Guan LL. Metatranscriptomic profiling reveals linkages between the active rumen microbiome and feed efficiency in beef cattle. *Appl Environ Microbiol.* (2017) 83:e00061-17. doi: 10.1128/AEM.00061-17
50. McCormack UM, Curiao T, Buzoianu SG, Prieto ML, Ryan T, Varley P, et al. Exploring a possible link between the intestinal microbiota and feed efficiency in pigs. *Appl Environ Microbiol.* (2017) 83:e00380-17. doi: 10.1128/AEM.00380-17
51. Yang H, Huang X, Fang S, He M, Zhao Y, Wu Z, et al. Unraveling the fecal microbiota and metagenomic functional capacity associated with feed efficiency in pigs. *Front Microbiol.* (2017) 8:1555. doi: 10.3389/fmicb.2017.01555
52. Stanley D, Hughes RJ, Geier MS, Moore RJ. Bacteria within the gastrointestinal tract microbiota correlated with improved growth and feed conversion: challenges presented for the identification of performance enhancing probiotic bacteria. *Front Microbiol.* (2016) 7:187. doi: 10.3389/fmicb.2016.00187
53. Geraylou Z, Souffreau C, Rurangwa E, D'Hondt S, Callewaert L, Courtin CM, et al. Effects of arabinoxylan-oligosaccharides (AXOS) on juvenile Siberian sturgeon (*Acipenser baerii*) performance, immune responses and gastrointestinal microbial community. *Fish Shellfish Immunol.* (2012) 33:718–24. doi: 10.1016/j.fsi.2012.06.010
54. Ley RE, Bäckhed F, Turnbaugh P, Lozupone CA, Knight RD, Gordon JI. Obesity alters gut microbial ecology. *Proc Natl Acad Sci USA* (2005) 102:11070–5. doi: 10.1073/pnas.0504978102
55. Dowarah R, Verma AK, Agarwal N. The use of *Lactobacillus* as an alternative of antibiotic growth promoters in pigs: a review. *Anim Nutr.* (2017) 3:1–6. doi: 10.1016/j.aninu.2016.11.002
56. Crisol-Martinez E, Stanley D, Geier MS, Hughes RJ, Moore RJ. Understanding the mechanisms of zinc bacitracin and avilamycin on animal production: linking gut microbiota and growth performance in chickens. *Appl Microbiol Biotechnol.* (2017) 101:4547–59. doi: 10.1007/s00253-017-8193-9
57. Savage DC, Siegel JE, Snellen JE, Whitt DD. Transit time of epithelial cells in the small intestines of germfree mice and ex-germfree mice associated with indigenous microorganisms. *Appl Environ Microbiol.* (1981) 42:996–1001.
58. Rawls JF, Samuel BS, Gordon JI. Gnotobiotic zebrafish reveal evolutionarily conserved responses to the gut microbiota. *Proc Natl Acad Sci USA.* (2004) 101:4596–601. doi: 10.1073/pnas.0400706101
59. Sommer F, Nookaew I, Sommer N, Fogelstrand P, Bäckhed F. Site-specific programming of the host epithelial transcriptome by the gut microbiota. *Genome Biol.* (2015) 16:62. doi: 10.1186/s13059-015-0614-4
60. Sapp M, Ploch S, Fiore-Donno AM, Bonkowski M, Rose LE. Protists are an integral part of the *Arabidopsis thaliana* microbiome. *Environ Microbiol.* (2017) 20:30–43. doi: 10.1111/1462-2920.13941
61. Pieterse CMJ, Leon-Reyes A, Van der Ent S, Van Wees SCM. Networking by small-molecule hormones in plant immunity. *Nat Chem Biol.* (2009) 5:308. doi: 10.1038/nchembio.164
62. Foster RC. Microenvironments of soil microorganisms. *Biol Fertil Soils* (1988) 6:189–203. doi: 10.1007/BF00260816
63. Berendsen RL, Pieterse CMJ, Bakker PAHM. The rhizosphere microbiome and plant health. *Trends Plant Sci.* (2012) 17:478–86. doi: 10.1016/j.tplants.2012.04.001
64. Hernández M, Dumont MG, Yuan Q, Conrad R. Different bacterial populations associated with the roots and rhizosphere of rice incorporate plant-derived carbon. *Appl Environ Microbiol.* (2015) 81:2244–53. doi: 10.1128/AEM.03209-14
65. Edwards J, Johnson C, Santos-Medellin C, Lurie E, Podishetty NK, Bhatnagar S, et al. Structure, variation, and assembly of the root-associated microbiomes of rice. *Proc Natl Acad Sci USA.* (2015) 112:E911–20. doi: 10.1073/pnas.1414592112
66. Bulgarelli D, Garrido-Oter R, Munch PC, Weiman A, Droge J, Pan Y, et al. Structure and function of the bacterial root microbiota in wild and domesticated barley. *Cell Host Microbe* (2015) 17:392–403. doi: 10.1016/j.chom.2015.01.011
67. Bodenhausen N, Horton MW, Bergelson J. Bacterial communities associated with the leaves and the roots of *Arabidopsis thaliana*. *PLoS ONE* (2013) 8:e56329. doi: 10.1371/journal.pone.0056329
68. Knief C, Delmotte N, Chaffron S, Stark M, Innerebner G, Wassmann R, et al. Metaproteogenomic analysis of microbial communities in the phyllosphere and rhizosphere of rice. *ISME J.* (2012) 6:1378–90. doi: 10.1038/ismej.2011.192
69. Francino MP. Early development of the gut microbiota and immune health. *Pathogens* (2014) 3:769–90. doi: 10.3390/pathogens3030769
70. Schokker D, Zhang J, Zhang L, Vastenhouw SA, Heilig HGHJ, Smidt H, et al. Early-life environmental variation affects intestinal microbiota and immune development in new-born piglets. *PLoS ONE* (2014) 9:e100040. doi: 10.1371/journal.pone.0100040
71. Moeser AJ, Pohl CS, Rajput M. Weaning stress and gastrointestinal barrier development: implications for lifelong gut health in pigs. *Anim Nutr.* (2017) 3:313–21. doi: 10.1016/j.aninu.2017.06.003
72. Matsuki T, Yahagi K, Mori H, Matsumoto H, Hara T, Tajima S, et al. A key genetic factor for fucosyllactose utilization affects infant gut microbiota development. *Nat Commun.* (2016) 7:11939. doi: 10.1038/ncomms11939
73. Videnska P, Sedlar K, Lukac M, Faldynova M, Gerzova L, Cejkova D, et al. Succession and replacement of bacterial populations in the caecum of egg laying hens over their whole life. *PLoS ONE* (2014) 9:e115142. doi: 10.1371/journal.pone.0115142
74. Okai S, Usui F, Yokota S, Hori-i Y, Hasegawa M, Nakamura T, et al. High-affinity monoclonal IgA regulates gut microbiota and prevents colitis in mice. *Nat Microbiol.* (2016) 1:16103. doi: 10.1038/nmicrobiol.2016.103
75. Liu Y, de Bruijn I, Jack ALH, Drynan K, van den Berg AH, Thoen E, et al. Deciphering microbial landscapes of fish eggs to mitigate emerging diseases. *ISME J.* (2014) 8:2002. doi: 10.1038/ismej.2014.44
76. Tinh NTN, Dierckens K, Sorgeloos P, Bossier P. A review of the functionality of probiotics in the larviculture food chain. *Mar Biotechnol (NY).* (2008) 10:1–12. doi: 10.1007/s10126-007-9054-9
77. Llewellyn MS, Boutin S, Hoseinifar SH, Derome N. Teleost microbiomes: the state of the art in their characterization, manipulation and importance in aquaculture and fisheries. *Front Microbiol.* (2014) 5:207. doi: 10.3389/fmicb.2014.00207
78. Ringo E, Birkbeck TH. Intestinal microflora of fish larvae and fry. *Aquac Res.* (1999) 30:73–93. doi: 10.1046/j.1365-2109.1999.00302.x
79. Olafsen JA. Interactions between fish larvae and bacteria in marine aquaculture. *Aquaculture* (2001) 200:223–47. doi: 10.1016/S0044-8486(01)00702-5

80. Lauzon HL, Gudmundsdottir S, Petursdottir SK, Reynisson E, Steinarrsson A, Oddgeirsson M, et al. Microbiota of Atlantic cod (*Gadus morhua* L.) rearing systems at pre- and posthatch stages and the effect of different treatments. *J Appl Microbiol.* (2010) 109:1775–89. doi: 10.1111/j.1365-2672.2010.04806.x
81. Romero J, Navarrete P. 16S rDNA-based analysis of dominant bacterial populations associated with early life stages of coho salmon (*Oncorhynchus kisutch*). *Microb Ecol.* (2006) 51:422–30. doi: 10.1007/s00248-006-9037-9
82. Nelson EB. The seed microbiome: origins, interactions, and impacts. *Plant Soil* (2018) 422:7–34. doi: 10.1007/s11104-017-3289-7
83. Bakker PAHM, Pieterse CMJ, de Jonge R, Berendsen RL. The soil-borne legacy. *Cell* (2018) 172:1178–80. doi: 10.1016/j.cell.2018.02.024
84. Berendsen RL, Vismans G, Yu K, Song Y, de Jonge R, Burgman WP, et al. Disease-induced assemblage of a plant-beneficial bacterial consortium. *ISME J.* (2018) 12:1496–1507. doi: 10.1038/s41396-018-0093-1
85. Marks PA, Richon VM, Rifkind RA. Histone deacetylase inhibitors: inducers of differentiation or apoptosis of transformed cells. *J Natl Cancer Inst.* (2000) 92:1210–6. doi: 10.1093/jnci/92.15.1210
86. Galindo-Villegas J, Garcia-Moreno D, de Oliveira S, Meseguer J, Mulero V. Regulation of immunity and disease resistance by commensal microbes and chromatin modifications during zebrafish development. *Proc Natl Acad Sci USA.* (2012) 109:E2605–14. doi: 10.1073/pnas.1209920109
87. Ramirez-Prado JS, Piquerez SJM, Bendahmane A, Hirt H, Raynaud C, Benhamed M. Modify the histone to win the battle: chromatin dynamics in plant–pathogen interactions. *Front Plant Sci.* (2018) 9:355. doi: 10.3389/fpls.2018.00355
88. Bhat MI, Kapila R. Dietary metabolites derived from gut microbiota: critical modulators of epigenetic changes in mammals. *Nutr Rev.* (2017) 75:374–89. doi: 10.1093/nutrit/nux001
89. Stringlis IA, Proietti S, Hickman R, Van Verk MC, Zamioudis C, Pieterse CMJ. Root transcriptional dynamics induced by beneficial rhizobacteria and microbial immune elicitors reveal signatures of adaptation to mutualists. *Plant J.* (2018) 93:166–80. doi: 10.1111/tpj.13741
90. Macfarlane GT, Gibson GR, Beatty E, Cummings JH. Estimation of short-chain fatty acid production from protein by human intestinal bacteria based on branched-chain fatty acid measurements. *FEMS Microbiol Lett.* (1992) 101:81–8. doi: 10.1111/j.1574-6968.1992.tb05764.x
91. Kihara M, Sakata T. Fermentation of dietary carbohydrates to short-chain fatty acids by gut microbes and its influence on intestinal morphology of a detritivorous teleost tilapia (*Oreochromis niloticus*). *Comp Biochem Physiol Physiol.* (1997) 118:1201–7. doi: 10.1016/S0300-9629(97)00052-2
92. Krajmalnik-Brown R, Ilhan Z-E, Kang D-W, DiBaise JK. Effects of gut microbes on nutrient absorption and energy regulation. *Nutr Clin Pract.* (2012) 27:201–14. doi: 10.1177/0884533611436116
93. Macfarlane GT, Macfarlane S. Bacteria, colonic fermentation, and gastrointestinal health. *J AOAC Int.* (2012) 95:50–60. doi: 10.5740/jaoacint.SGE_Macfarlane
94. Koh A, De Vadder F, Kovatcheva-Datchary P, Bäckhed F. From dietary fiber to host physiology: short-chain fatty acids as key bacterial metabolites. *Cell* (2016) 165:1332–45. doi: 10.1016/j.cell.2016.05.041
95. van Beers-Schreurs HM, Nabuurs MJ, Vellenga L, Kalsbeek-van der Valk HJ, Wensing T, Breukink HJ. Weaning and the weanling diet influence the villous height and crypt depth in the small intestine of pigs and alter the concentrations of short-chain fatty acids in the large intestine and blood. *J Nutr.* (1998) 128:947–53. doi: 10.1093/jn/128.6.947
96. Montagne L, Pluske JR, Hampson DJ. A review of interactions between dietary fibre and the intestinal mucosa, and their consequences on digestive health in young non-ruminant animals. *Anim Feed Sci Technol.* (2003) 108:95–117. doi: 10.1016/S0377-8401(03)00163-9
97. Smith PM, Howitt MR, Panikov N, Michaud M, Gallini CA, Bohlooly-Y M, et al. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science* (2013) 341:569LP–73. doi: 10.1126/science.1241165
98. Guilloteau P, Martin L, Eeckhaut V, Ducatelle R, Zabielski R, Van Immerseel F. From the gut to the peripheral tissues: the multiple effects of butyrate. *Nutr Res Rev.* (2010) 23:366–84. doi: 10.1017/S0954422410000247
99. Tan J, McKenzie C, Potamitis M, Thorburn AN, Mackay CR, Macia L. The role of short-chain fatty acids in health and disease. *Adv Immunol.* (2014) 121:91–119. doi: 10.1016/B978-0-12-800100-4.00003-9
100. Kelly CJ, Zheng L, Campbell EL, Saeedi B, Scholz CC, Bayless AJ, et al. Crosstalk between microbiota-derived short-chain fatty acids and intestinal epithelial HIF augments tissue barrier function. *Cell Host Microbe* (2015) 17:662–71. doi: 10.1016/j.chom.2015.03.005
101. Nafday SM, Chen W, Peng L, Babyatsky MW, Holzman IR, Lin J. Short-chain fatty acids induce colonic mucosal injury in rats with various postnatal ages. *Pediatr Res.* (2005) 57:201–4. doi: 10.1203/01.PDR.0000150721.83224.89
102. Peng L, He Z, Chen W, Holzman IR, Lin J. Effects of butyrate on intestinal barrier function in a Caco-2 cell monolayer model of intestinal barrier. *Pediatr Res.* (2007) 61:37. doi: 10.1203/01.pdr.0000250014.92242.f3
103. Hughes ER, Winter MG, Duerkop BA, Spiga L, Furtado de Carvalho T, Zhu W, et al. Microbial respiration and formate oxidation as metabolic signatures of inflammation-associated dysbiosis. *Cell Host Microbe* (2017) 21:208–19. doi: 10.1016/j.chom.2017.01.005
104. El-Ansary AK, Ben Bacha A, Kotb M. Etiology of autistic features: the persisting neurotoxic effects of propionic acid. *J Neuroinflamm.* (2012) 9:74. doi: 10.1186/1742-2094-9-74
105. Wiley NC, Dinan TG, Ross RP, Stanton C, Clarke G, Cryan JF. The microbiota-gut-brain axis as a key regulator of neural function and the stress response: implications for human and animal health. *J Anim Sci.* (2017) 95:3225–46. doi: 10.2527/jas.2016.1256
106. Sola-Penna M. Metabolic regulation by lactate. *IUBMB Life* (2008) 60:605–8. doi: 10.1002/iub.97
107. Proia P, Di Liegro MC, Schiera G, Fricano A, Di Liegro I. Lactate as a metabolite and a regulator in the central nervous system. *Int J Mol Sci.* (2016) 17: doi: 10.3390/ijms17091450
108. Okada T, Fukuda S, Hase K, Nishiumi S, Izumi Y, Yoshida M, et al. Microbiota-derived lactate accelerates colon epithelial cell turnover in starvation-refed mice. *Nat Commun.* (2013) 4:1654. doi: 10.1038/ncomms2668
109. Bourriaud C, Robins RJ, Martin L, Kozłowski F, Tenailleu E, Cherbut C, et al. Lactate is mainly fermented to butyrate by human intestinal microfloras but inter-individual variation is evident. *J Appl Microbiol.* (2005) 99:201–12. doi: 10.1111/j.1365-2672.2005.02605.x
110. Condon S. Responses of lactic acid bacteria to oxygen*. *FEMS Microbiol Rev.* (1987) 3:269–80. doi: 10.1111/j.1574-6968.1987.tb02465.x
111. Guo T, Zhang L, Xin Y, Xu Z, He H, Kong J. Oxygen-inducible conversion of lactate to acetate in heterofermentative *Lactobacillus brevis* ATCC 367. *Appl Environ Microbiol.* (2017) 83: doi: 10.1128/AEM.01659-17
112. Kahlert S, Junnikkala S, Renner L, Hynönen U, Hartig R, Nossol C, et al. Physiological concentration of exogenous lactate reduces antimycin A triggered oxidative stress in intestinal epithelial cell line IPEC-1 and IPEC-J2 *in vitro*. *PLoS ONE* (2016) 11:e0153135. doi: 10.1371/journal.pone.0153135
113. Ferreyra JA, Wu KJ, Hryckowian AJ, Bouley DM, Weimer BC, Sonnenburg JL. Gut microbiota-produced succinate promotes *C. difficile* infection after antibiotic treatment or motility disturbance. *Cell Host Microbe* (2014) 16:770–7. doi: 10.1016/j.chom.2014.11.003
114. De Vadder F, Kovatcheva-Datchary P, Zitoun C, Duchamp A, Bäckhed F, Mithieux G. Microbiota-produced succinate improves glucose homeostasis via intestinal gluconeogenesis. *Cell Metab.* (2016) 24:151–7. doi: 10.1016/j.cmet.2016.06.013
115. Serena C, Ceperuelo-Mallafre V, Keiran N, Queipo-Ortuño MI, Bernal R, Gomez-Huelgas R, et al. Elevated circulating levels of succinate in human obesity are linked to specific gut microbiota. *ISME J.* (2018) doi: 10.1038/s41396-018-0068-2
116. Tsukahara T, Ushida K. Organic acid profiles in feces of pigs with pathogenic or non-pathogenic diarrhea. *J Vet Med Sci.* (2001) 63:1351–4. doi: 10.1292/jvms.63.1351
117. Tsukahara T, Ushida K. Succinate accumulation in pig large intestine during antibiotic-associated diarrhea and the constitution of succinate-producing flora. *J Gen Appl Microbiol.* (2002) 48:143–54. doi: 10.2323/jgam.48.143
118. Højberg O, Canibe N, Poulsen HD, Hedemann MS, Jensen BB. Influence of dietary zinc oxide and copper sulfate on the gastrointestinal ecosystem in newly weaned piglets. *Appl Environ Microbiol.* (2005) 71:2267–77. doi: 10.1128/AEM.71.5.2267-2277.2005
119. Hernández J, Benedito JL, Abuelo A, Castillo C. Ruminal acidosis in feedlot: from aetiology to prevention. *Sci World J.* (2014) 2014:702572. doi: 10.1155/2014/702572

120. Doornbos RF, van Loon LC, Bakker PAHM. Impact of root exudates and plant defense signaling on bacterial communities in the rhizosphere. a review. *Agron Sustain Dev.* (2012) 32:227–43. doi: 10.1007/s13593-011-0028-y
121. Stringlis IA, Yu K, Feussner K, de Jonge R, Van Bentum S, Van Verk MC, et al. MYB72-dependent coumarin exudation shapes root microbiome assembly to promote plant health. *Proc Natl Acad Sci USA.* (2018). 115:E5213–22. doi: 10.1073/pnas.1722335115
122. Glick BR. Modulation of plant ethylene levels by the bacterial enzyme ACC deaminase. *FEMS Microbiol Lett.* (2005) 251:1–7. doi: 10.1016/j.femsle.2005.07.030
123. Kamilova F, Kravchenko L V, Shaposhnikov AI, Makarova N, Lugtenberg B. Effects of the tomato pathogen *Fusarium oxysporum* f. sp. *radicis-lycopersici* and of the biocontrol bacterium *Pseudomonas fluorescens* WCS365 on the composition of organic acids and sugars in tomato root exudate. *Mol Plant Microbe Interact.* (2006) 19:1121–6. doi: 10.1094/MPMI-19-1121
124. Iyer B, Rajput MS, Rajkumar S. Effect of succinate on phosphate solubilization in nitrogen fixing bacteria harbouring chick pea and their effect on plant growth. *Microbiol Res.* (2017) 202:43–50. doi: 10.1016/j.micres.2017.05.005
125. Wu H, Wu L, Zhu Q, Wang J, Qin X, Xu J, et al. The role of organic acids on microbial deterioration in the *Radix pseudostellariae* rhizosphere under continuous monoculture regimes. *Sci Rep.* (2017) 7:3497. doi: 10.1038/s41598-017-03793-8
126. Loqué D, von Witrén N. Regulatory levels for the transport of ammonium in plant roots. *J Exp Bot.* (2004) 55:1293–1305. doi: 10.1093/jxb/erh147
127. Hrnjez BJ, Song JC, Prasad M, Mayol JM, Matthews JB. Ammonia blockade of intestinal epithelial K⁺ conductance. *Am J Physiol.* (1999) 277:G521–32.
128. Davila A-M, Blachier F, Gotteland M, Andriamihaja M, Benetti P-H, Sanz Y, et al. Intestinal luminal nitrogen metabolism: role of the gut microbiota and consequences for the host. *Pharmacol Res.* (2013) 68:95–107. doi: 10.1016/j.phrs.2012.11.005
129. Apajalahti J, Vienola K. Interaction between chicken intestinal microbiota and protein digestion. *Anim Feed Sci Technol.* (2016) 221:323–330. doi: 10.1016/j.anifeeds.2016.05.004
130. Tsujii M, Kawano S, Tsuji S, Fusamoto H, Kamada T, Sato N. Mechanism of gastric mucosal damage induced by ammonia. *Gastroenterology* (1992) 102:1881–8.
131. Podgorska A, Burian M, Gieczewska K, Ostaszewska-Bugajska M, Zebrowski J, Solecka D, et al. Altered cell wall plasticity can restrict plant growth under ammonium nutrition. *Front Plant Sci.* (2017) 8:1344. doi: 10.3389/fpls.2017.01344
132. Premont RT, Gainetdinov RR, Caron MG. Following the trace of elusive amines. *Proc Natl Acad Sci USA.* (2001) 98:9474–5. doi: 10.1073/pnas.181356198
133. Spano G, Russo P, Lonvaud-Funel A, Lucas P, Alexandre H, Grandvalet C, et al. Biogenic amines in fermented foods. *Eur J Clin Nutr.* (2010) 64(Suppl 3):S95–100. doi: 10.1038/ejcn.2010.218
134. Williams BB, Van Benschoten AH, Cimermanic P, Donia MS, Zimmermann M, Taketani M, et al. Discovery and characterization of gut microbiota decarboxylases that can produce the neurotransmitter tryptamine. *Cell Host Microbe* (2014) 16:495–503. doi: 10.1016/j.chom.2014.09.001
135. Yano JM, Yu K, Donaldson GP, Shastri GG, Ann P, Ma L, et al. Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. *Cell* (2015) 161:264–76. doi: 10.1016/j.cell.2015.02.047
136. Galligan JJ. Beneficial actions of microbiota-derived tryptophan metabolites. *Neurogastroenterol Motil.* (2018) 30:e13283. doi: 10.1111/nmo.13283
137. Spaepen S, Vanderleyden J, Remans R. Indole-3-acetic acid in microbial and microorganism-plant signaling. *FEMS Microbiol Rev.* (2007) 31:425–48. doi: 10.1111/j.1574-6976.2007.00072.x
138. Dowling DN, O’Gara F. Metabolites of *Pseudomonas* involved in the biocontrol of plant disease. *Trends Biotechnol.* (1994) 12:133–41. doi: 10.1016/0167-7799(94)90091-4
139. Stringlis IA, Zhang H, Pieterse CMJ, Bolton MD, de Jonge R. Microbial small molecules - weapons of plant subversion. *Nat Prod Rep.* (2018) 35:410–33. doi: 10.1039/c7np00062f
140. Loeschcke A, Thies S. *Pseudomonas putida*—a versatile host for the production of natural products. *Appl Microbiol Biotechnol.* (2015) 99:6197–14. doi: 10.1007/s00253-015-6745-4
141. Sharon G, Garg N, Debelius J, Knight R, Dorrestein PC, Mazmanian SK. Specialized metabolites from the microbiome in health and disease. *Cell Metab.* (2014) 20:719–30. doi: 10.1016/j.cmet.2014.10.016
142. Miethke M, Marahiel MA. Siderophore-based iron acquisition and pathogen control. *Microbiol Mol Biol Rev.* (2007) 71:413–51. doi: 10.1128/MMBR.00012-07
143. Garcia-Gutierrez E, Mayer MJ, Cotter PD, Narbad A. Gut microbiota as a source of novel antimicrobials. *Gut Microbes* (2018) 27:1–21. doi: 10.1080/19490976.2018.1455790
144. Walsh CJ, Guinane CM, Hill C, Ross RP, O’Toole PW, Cotter PD. *In silico* identification of bacteriocin gene clusters in the gastrointestinal tract, based on the Human Microbiome Project’s reference genome database. *BMC Microbiol.* (2015) 15:183. doi: 10.1186/s12866-015-0515-4
145. Donia MS, Cimermanic P, Schulze CJ, Wieland Brown LC, Martin J, Mitreva M, et al. A systematic analysis of biosynthetic gene clusters in the human microbiome reveals a common family of antibiotics. *Cell* (2014) 158:1402–14. doi: 10.1016/j.cell.2014.08.032
146. Taga ME, Bassler BL. Chemical communication among bacteria. *Proc Natl Acad Sci USA* (2003) 100(Suppl.):14549–54. doi: 10.1073/pnas.1934514100
147. Bruhn JB, Dalsgaard I, Nielsen KE, Buchholtz C, Larsen JL, Gram L. Quorum sensing signal molecules (acylated homoserine lactones) in gram-negative fish pathogenic bacteria. *Dis Aquat Organ.* (2005) 65:43–52. doi: 10.3354/dao065043
148. Swearingen MC, Sabag-Daigle A, Ahmer BMM. Are there acyl-homoserine lactones within mammalian intestines? *J Bacteriol.* (2013) 195:173–9. doi: 10.1128/JB.01341-12
149. Federle MJ. Autoinducer-2-based chemical communication in bacteria: complexities of interspecies signaling. *Contrib Microbiol.* (2009) 16:18–32. doi: 10.1159/000219371
150. Thompson JA, Oliveira RA, Djukovic A, Ubeda C, Xavier KB. Manipulation of the quorum sensing signal AI-2 affects the antibiotic-treated gut microbiota. *Cell Rep.* (2015) 10:1861–71. doi: 10.1016/j.celrep.2015.02.049
151. Sugita H, Miyajima C, Deguchi Y. The vitamin B12-producing ability of the intestinal microflora of freshwater fish. *Aquaculture* (1991) 92:267–76. doi: 10.1016/0044-8486(91)90028-6
152. Degnan PH, Taga ME, Goodman AL. Vitamin B12 as a modulator of gut microbial ecology. *Cell Metab.* (2014) 20:769–78. doi: 10.1016/j.cmet.2014.10.002
153. Hooper LV, Midtvedt T, Gordon JL. How host-microbial interactions shape the nutrient environment of the mammalian intestine. *Annu Rev Nutr.* (2002) 22:283–307. doi: 10.1146/annurev.nutr.22.011602.092259
154. Magnúsdóttir S, Ravcheev D, de Crécy-Lagard V, Thiele I. Systematic genome assessment of B-vitamin biosynthesis suggests co-operation among gut microbes. *Front Genet.* (2015) 6:148. doi: 10.3389/fgenet.2015.00148
155. Biesalski HK. Nutrition meets the microbiome: micronutrients and the microbiota. *Ann N Y Acad Sci.* (2016) 1372:53–64. doi: 10.1111/nyas.13145
156. Smith AG, Croft MT, Moulin M, Webb ME. Plants need their vitamins too. *Curr Opin Plant Biol.* (2007) 10:266–75. doi: 10.1016/j.pbi.2007.04.009
157. Bittel P, Robatzek S. Microbe-associated molecular patterns (MAMPs) probe plant immunity. *Curr Opin Plant Biol.* (2007) 10:335–41. doi: 10.1016/j.pbi.2007.04.021
158. Newman M-A, Sundelin T, Nielsen JT, Erbs G. MAMP (microbe-associated molecular pattern) triggered immunity in plants. *Front Plant Sci.* (2013) 4:139. doi: 10.3389/fpls.2013.00139
159. Nürnberger T, Brunner F, Kemmerling B, Piater L. Innate immunity in plants and animals: striking similarities and obvious differences. *Immunol Rev.* (2004) 198:249–66. doi: 10.1111/j.0105-2896.2004.01119.x
160. Ausubel FM. Are innate immune signaling pathways in plants and animals conserved? *Nat Immunol.* (2005) 6:973. doi: 10.1038/ni1253
161. Mackey D, McFall A. MAMPs and MIMPs: proposed classifications for inducers of innate immunity. *Mol Microbiol.* (2006) 61:1365–71. doi: 10.1111/j.1365-2958.2006.05311.x
162. Lathrop SK, Bloom SM, Rao SM, Nutsch K, Lio C-W, Santacruz N, et al. Peripheral education of the immune system by colonic commensal microbiota. *Nature* (2011) 478:250. doi: 10.1038/nature10434

163. Zheng Y, Chaudhry A, Kas A, deRoos P, Kim JM, Chu T-T, et al. Regulatory T-cell suppressor program co-opts transcription factor IRF4 to control T(H)2 responses. *Nature* (2009) 458:351–6. doi: 10.1038/nature07674
164. Atarashi K, Tanoue T, Shima T, Imaoka A, Kuwahara T, Momose Y, et al. Induction of colonic regulatory T cells by indigenous Clostridium species. *Science* (2011) 331:337–41. doi: 10.1126/science.1198469
165. Mazmanian SK, Liu CH, Tzianabos AO, Kasper DL. An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. *Cell* (2005) 122:107–18. doi: 10.1016/j.cell.2005.05.007
166. Schnorr SL, Candela M, Rampelli S, Centanni M, Consolandi C, Basaglia G, et al. Gut microbiome of the Hadza hunter-gatherers. *Nat Commun.* (2014) 5:3654. doi: 10.1038/ncomms4654
167. Obregon-Tito AJ, Tito RY, Metcalf J, Sankaranarayanan K, Clemente JC, Ursell LK, et al. Subsistence strategies in traditional societies distinguish gut microbiomes. *Nat Commun.* (2015) 6:6505. doi: 10.1038/ncomms7505
168. Sonnenburg ED, Smits SA, Tikhonov M, Higginbottom SK, Wingreen NS, Sonnenburg JL. Diet-induced extinction in the gut microbiota compounds over generations. *Nature* (2016) 529:212–5. doi: 10.1038/nature16504
169. Clayton JB, Vangay P, Huang H, Ward T, Hillmann BM, Al-Ghalith GA, et al. Captivity humanizes the primate microbiome. *Proc Natl Acad Sci USA.* (2016) 113:10376–81. doi: 10.1073/pnas.1521835113
170. Metcalf JL, Song SJ, Morton JT, Weiss S, Seguin-Orlando A, Joly F, et al. Evaluating the impact of domestication and captivity on the horse gut microbiome. *Sci Rep.* (2017) 7:15497. doi: 10.1038/s41598-017-15375-9
171. Kreisinger J, Dagmar C, Vohánka J, Piálek J. Gastrointestinal microbiota of wild and inbred individuals of two house mouse subspecies assessed using high-throughput parallel pyrosequencing. *Mol Ecol.* (2014) 23:5048–60. doi: 10.1111/mec.12909
172. Xenoulis PG, Gray PL, Brightsmith D, Palculict B, Hoppes S, Steiner JM, et al. Molecular characterization of the cloacal microbiota of wild and captive parrots. *Vet Microbiol.* (2010) 146:320–5. doi: 10.1016/j.vetmic.2010.05.024
173. Bisgaard H, Li N, Bonnelykke K, Chawes BLK, Skov T, Paludan-Muller G, et al. Reduced diversity of the intestinal microbiota during infancy is associated with increased risk of allergic disease at school age. *J Allergy Clin Immunol.* (2011) 128:645–6. doi: 10.1016/j.jaci.2011.04.060
174. Ott SJ, Musfeldt M, Wenderoth DF, Hampe J, Brant O, Fölsch UR, et al. Reduction in diversity of the colonic mucosa associated bacterial microflora in patients with active inflammatory bowel disease. *Gut* (2004) 53:685–93. doi: 10.1136/gut.2003.025403
175. O'Donnell AG, Seasman M, Macrae A, Waite I, Davies JT. Plants and fertilisers as drivers of change in microbial community structure and function in soils. *Plant Soil* (2001) 232:135–45. doi: 10.1023/A:1010394221729
176. Williams TR, Marco ML. Phyllosphere microbiota composition and microbial community transplantation on lettuce plants grown indoors. *MBio* (2014) 5:e01564-14. doi: 10.1128/mBio.01564-14
177. Rosshart SP, Vassallo BG, Angeletti D, Hutchinson DS, Morgan AP, Takeda K, et al. Wild mouse gut microbiota promotes host fitness and improves disease resistance. *Cell* (2017) 171:1015.e13–28.e13. doi: 10.1016/j.cell.2017.09.016
178. Castillo M, Martín-Ortúe SM, Manzanilla EG, Badiola I, Martín M, Gasa J. Quantification of total bacteria, enterobacteria and lactobacilli populations in pig digesta by real-time PCR. *Vet Microbiol.* (2006) 114:165–70. doi: 10.1016/j.vetmic.2005.11.055
179. Ushida K, Tsuchida S, Ogura Y, Toyoda A, Maruyama F. Domestication and cereal feeding developed domestic pig-type intestinal microbiota in animals of suidae. *Anim Sci J.* (2016) 87:835–41. doi: 10.1111/asj.12492
180. Dou S, Gadonna-Widehem P, Rome V, Hamoudi D, Rhazi L, Lakhali L, et al. Characterisation of early-life fecal microbiota in susceptible and healthy pigs to post-weaning diarrhoea. *PLoS ONE* (2017) 12:e0169851. doi: 10.1371/journal.pone.0169851
181. Ribeiro GO, Oss DB, He Z, Gruninger RJ, Elekwachi C, Forster RJ, et al. Repeated inoculation of cattle rumen with bison rumen contents alters the rumen microbiome and improves nitrogen digestibility in cattle. *Sci Rep.* (2017) 7:1276. doi: 10.1038/s41598-017-01269-3
182. Rungrasamee W, Klanchui A, Maibunkaew S, Chaiyapechara S, Jiravanichpaisal P, Karoonathaisiri N. Characterization of intestinal bacteria in wild and domesticated adult black tiger shrimp (*Penaeus monodon*). *PLoS ONE* (2014) 9:e91853. doi: 10.1371/journal.pone.0091853
183. Dhanasiri AKS, Brunvold L, Brinchmann MF, Korsnes K, Bergh Ø, Kiron V. Changes in the intestinal microbiota of wild atlantic cod *Gadus morhua* L. upon captive rearing. *Microb Ecol.* (2011) 61:20–30. doi: 10.1007/s00248-010-9673-y
184. Pieterse CMJ, de Jonge R, Berendsen RL. The soil-borne supremacy. *Trends Plant Sci.* (2016) 21:171–3. doi: 10.1016/j.tplants.2016.01.018
185. Zachow C, Müller H, Tilcher R, Berg G. Differences between the rhizosphere microbiome of *Beta vulgaris* ssp. *maritima*—ancestor of all beet crops—and modern sugar beets. *Front Microbiol.* (2014) 5:415. doi: 10.3389/fmicb.2014.00415
186. Campisano A, Pancher M, Puopolo G, Puddu A, López-Fernández S, Biagini B, et al. Diversity in endophyte populations reveals functional and taxonomic diversity between wild and domesticated grapevines. *Am J Enol Vitic.* (2015) 66:12 LP-21. doi: 10.5344/ajev.2014.14046
187. Pieterse CMJ, Van der Does D, Zamioudis C, Leon-Reyes A, Van Wees SCM. Hormonal Modulation of Plant Immunity. *Annu Rev Cell Dev Biol.* (2012) 28:489–521. doi: 10.1146/annurev-cellbio-092910-154055
188. Roehre R, Dewhurst RJ, Duthie C-A, Rooke JA, McKain N, Ross DW, et al. Bovine host genetic variation influences rumen microbial methane production with best selection criterion for low methane emitting and efficiently feed converting hosts based on metagenomic gene abundance. *PLoS Genet.* (2016) 12:e1005846. doi: 10.1371/journal.pgen.1005846
189. Camarinha-Silva A, Maushammer M, Wellmann R, Vital M, Preuss S, Bennewitz J. Host genome influence on gut microbial composition and microbial prediction of complex traits in pigs. *Genetics* (2017) 206:1637–44. doi: 10.1534/genetics.117.200782
190. Benson AK, Kelly SA, Legge R, Ma F, Low SJ, Kim J, et al. Individuality in gut microbiota composition is a complex polygenic trait shaped by multiple environmental and host genetic factors. *Proc Natl Acad Sci USA.* (2010) 107:18933–8. doi: 10.1073/pnas.1007028107
191. Li X, Yu Y, Feng W, Yan Q, Gong Y. Host species as a strong determinant of the intestinal microbiota of fish larvae. *J Microbiol.* (2012) 50:29–37. doi: 10.1007/s12275-012-1340-1
192. Pratte ZA, Besson M, Hollman RD, Stewart FJ. The gills of reef fish support a distinct microbiome influenced by host-specific factors. *Appl Environ Microbiol.* (2018) 84:e00063-18. doi: 10.1128/AEM.00063-18
193. Pérez-Jaramillo JE, Mendes R, Raaijmakers JM. Impact of plant domestication on rhizosphere microbiome assembly and functions. *Plant Mol Biol.* (2016) 90:635–44. doi: 10.1007/s11103-015-0337-7
194. Mendes LW, Raaijmakers JM, de Hollander M, Mendes R, Tsai SM. Influence of resistance breeding in common bean on rhizosphere microbiome composition and function. *ISME J.* (2018) 12:212–24. doi: 10.1038/ismej.2017.158
195. Doyle CJ, Gleeson D, O'Toole PW, Cotter PD. Impacts of seasonal housing and teat preparation on raw milk microbiota: a high-throughput sequencing study. *Appl Environ Microbiol.* (2017) 83:e02694-16. doi: 10.1128/AEM.02694-16
196. Kubasova T, Davidova-Gerzova L, Merlot E, Medvecký M, Polansky O, Gardan-Salmon D, et al. Housing systems influence gut microbiota composition of sows but not of their piglets. *PLoS ONE* (2017) 12:e0170051. doi: 10.1371/journal.pone.0170051
197. Dehler CE, Secombes CJ, Martin SAM. Seawater transfer alters the intestinal microbiota profiles of Atlantic salmon (*Salmo salar* L.). *Sci Rep.* (2017) 7:13877. doi: 10.1038/s41598-017-13249-8
198. Giatsis C, Sipkema D, Smidt H, Heilig H, Benvenuti G, Verreth J, et al. The impact of rearing environment on the development of gut microbiota in tilapia larvae. *Sci Rep.* (2015) 5:18206. doi: 10.1038/srep18206
199. Schmidt, VT, Smith, KF, Melvin, DW, Amaral-Zettler, LA. Community assembly of a euryhaline fish microbiome during salinity acclimation. *Mol Ecol.* (2015) 24:2537–50. doi: 10.1111/mec.13177
200. Sonnenburg ED, Sonnenburg JL. Starving our microbial self: the deleterious consequences of a diet deficient in microbiota-accessible carbohydrates. *Cell Metab.* (2014) 20:779–86. doi: 10.1016/j.cmet.2014.07.003
201. Niu Q, Li P, Hao S, Zhang Y, Kim SW, Li H, et al. Dynamic distribution of the gut microbiota and the relationship with apparent crude fiber digestibility and growth stages in pigs. *Sci Rep.* (2015) 5:9938. doi: 10.1038/srep09938
202. Shanks OC, Kelty CA, Archibeque S, Jenkins M, Newton RJ, McLellan SL, et al. Community structures of fecal bacteria in cattle from different

- animal feeding operations. *Appl Environ Microbiol.* (2011) 77:2992–3001. doi: 10.1128/AEM.02988-10
203. Meale SJ, Li S, Azevedo P, Derakhshani H, Plaizier JC, Khafipour E, et al. Development of ruminal and fecal microbiomes are affected by weaning but not weaning strategy in dairy calves. *Front Microbiol.* (2016) 7:582. doi: 10.3389/fmicb.2016.00582
204. Gresse R, Chaucheyras-Durand F, Fleury MA, Van de Wiele T, Forano E, Blanquet-Diot S. Gut microbiota dysbiosis in postweaning piglets: understanding the keys to health. *Trends Microbiol.* (2017) 25:851–73. doi: 10.1016/j.tim.2017.05.004
205. Wu S, Wang G, Angert ER, Wang W, Li W, Zou H. Composition, diversity, and origin of the bacterial community in grass carp intestine. *PLoS ONE* (2012) 7:e30440. doi: 10.1371/journal.pone.0030440
206. Li H, Wu S, Wirth S, Hao Y, Wang W, Zou H, et al. Diversity and activity of cellulolytic bacteria, isolated from the gut contents of grass carp (*Ctenopharyngodon idellus*) (Valenciennes) fed on Sudan grass (*Sorghum sudanense*) or artificial feedstuffs. *Aquac Res.* (2014) 47:153–64. doi: 10.1111/are.12478
207. Ringø E, Zhou Z, Vecino JLG, Wadsworth S, Romero J, Krogdahl Å, et al. Effect of dietary components on the gut microbiota of aquatic animals. A never-ending story? *Aquac Nutr.* (2015) 22:219–82. doi: 10.1111/anu.12346
208. Jangid K, Williams MA, Franzluebbers AJ, Sanderlin JS, Reeves JH, Jenkins MB, et al. Relative impacts of land-use, management intensity and fertilization upon soil microbial community structure in agricultural systems. *Soil Biol Biochem.* (2008) 40:2843–53. doi: 10.1016/j.soilbio.2008.07.030
209. Santhanam R, Luu VT, Weinhold A, Goldberg J, Oh Y, Baldwin IT. Native root-associated bacteria rescue a plant from a sudden-wilt disease that emerged during continuous cropping. *Proc Natl Acad Sci USA.* (2015) 112:E5013–20. doi: 10.1073/pnas.1505765112
210. Song SJ, Lauber C, Costello EK, Lozupone CA, Humphrey G, Berg-Lyons D, et al. Cohabiting family members share microbiota with one another and with their dogs. *Elife* (2013) 2:e00458. doi: 10.7554/eLife.00458
211. Kraemer JG, Ramette A, Aebi S, Oppliger A, Hilty M. Influence of pig farming on the human's nasal microbiota: the key role of the airborne microbial communities. *Appl Environ Microbiol.* (2018) 12:AEM.02470-17. doi: 10.1128/AEM.02470-17
212. Soverini M, Rampelli S, Turroni S, Schnorr SL, Quercia S, Castagnetti A, et al. Variations in the post-weaning human gut metagenome profile as result of *Bifidobacterium* acquisition in the western microbiome. *Front Microbiol.* (2016) 7:1058. doi: 10.3389/fmicb.2016.01058
213. Milani C, Mangifesta M, Mancabelli L, Lugli GA, James K, Duranti S, et al. Unveiling bifidobacterial biogeography across the mammalian branch of the tree of life. *ISME J.* (2017) 11:2834. doi: 10.1038/ismej.2017.138
214. Dibner JJ, Richards JD. Antibiotic growth promoters in agriculture: history and mode of action. *Poult Sci.* (2005) 84:634–43. doi: 10.1093/ps/84.4.634
215. van Schaik W. The human gut resistome. *Philos Trans R Soc Lond B Biol Sci.* (2015) 370:20140087. doi: 10.1098/rstb.2014.0087
216. Andersson DI, Hughes D. Microbiological effects of sublethal levels of antibiotics. *Nat Rev Microbiol.* (2014) 12:465–78. doi: 10.1038/nrmicro3270
217. Mu C, Yang Y, Su Y, Zoetendal EG, Zhu W. Differences in microbiota membership along the gastrointestinal tract of piglets and their differential alterations following an early-life antibiotic intervention. *Front Microbiol.* (2017) 8:797. doi: 10.3389/fmicb.2017.00797
218. Looft T, Johnson TA, Allen HK, Bayles DO, Alt DP, Stedtfeld RD, et al. In-feed antibiotic effects on the swine intestinal microbiome. *Proc Natl Acad Sci USA.* (2012) 109:1691–6. doi: 10.1073/pnas.1120238109
219. Van Vleck Pereira R, Lima S, Siler JD, Foditsch C, Warnick LD, Bicalho RC. Ingestion of milk containing very low concentration of antimicrobials: longitudinal effect on fecal microbiota composition in preweaned calves. *PLoS ONE* (2016) 11:e0147525. doi: 10.1371/journal.pone.0147525
220. Navarrete P, Mardones P, Opazo R, Espejo R, Romero J. Oxytetracycline treatment reduces bacterial diversity of intestinal microbiota of Atlantic salmon. *J Aquat Anim Health* (2011) 20:177–83. doi: 10.1577/H07-043.1
221. Hammesfahr U, Heuer H, Manzke B, Smalla K, Thiele-Bruhn S. Impact of the antibiotic sulfadiazine and pig manure on the microbial community structure in agricultural soils. *Soil Biol Biochem.* (2008) 40:1583–91. doi: 10.1016/j.soilbio.2008.01.010
222. Liu F, Ying G-G, Tao R, Zhao J-L, Yang J-F, Zhao L-F. Effects of six selected antibiotics on plant growth and soil microbial and enzymatic activities. *Environ Pollut.* (2009) 157:1636–42. doi: 10.1016/j.envpol.2008.12.021
223. Xiong W, Sun Y, Zhang T, Ding X, Li Y, Wang M, et al. Antibiotics, antibiotic resistance genes and bacterial community composition in fresh water aquaculture environment in China. *Microb Ecol.* (2015) 70:425–32. doi: 10.1007/s00248-015-0583-x
224. Petersen A, Andersen JS, Kaewmak T, Somsiri T, Dalsgaard A. Impact of integrated fish farming on antimicrobial resistance in a pond environment. *Appl Environ Microbiol.* (2002) 68:6036–42. doi: 10.1128/AEM.68.12.6036-6042.2002
225. Cabello FC. Heavy use of prophylactic antibiotics in aquaculture: a growing problem for human and animal health and for the environment. *Environ Microbiol.* (2006) 8:1137–44. doi: 10.1111/j.1462-2920.2006.01054.x
226. Marshall BM, Levy SB. Food animals and antimicrobials: impacts on human health. *Clin Microbiol Rev.* (2011) 24:718–33. doi: 10.1128/CMR.00002-11
227. Jechalke S, Heuer H, Siemens J, Amelung W, Smalla K. Fate and effects of veterinary antibiotics in soil. *Trends Microbiol.* (2014) 22:536–45. doi: 10.1016/j.tim.2014.05.005
228. Mark Welch JL, Hasegawa Y, McNulty NP, Gordon JI, Borisy GG. Spatial organization of a model 15-member human gut microbiota established in gnotobiotic mice. *Proc Natl Acad Sci USA.* (2017) 114:E9105–14. doi: 10.1073/pnas.1711596114
229. Marteau P, Pochart P, Doré J, Béra-Maillet C, Bernalier A, Corthier G. Comparative study of bacterial groups within the human cecal and fecal microbiota. *Appl Environ Microbiol.* (2001) 67:4939–42. doi: 10.1128/AEM.67.10.4939-4942.2001
230. Wang W, Wu S, Zheng Y, Cheng Y, Li W, Zou H, et al. Characterization of the bacterial community associated with early-developmental stages of grass carp (*Ctenopharyngodon idella*). *Aquac Res.* (2014) 46:2728–35. doi: 10.1111/are.12428
231. Li X, Rui J, Xiong J, Li J, He Z, Zhou J, et al. Functional potential of soil microbial communities in the maize rhizosphere. *PLoS ONE* (2014) 9:e112609. doi: 10.1371/journal.pone.0112609
232. Stepanauskas R. Single cell genomics: an individual look at microbes. *Curr Opin Microbiol.* (2012) 15:613–20. doi: 10.1016/j.mib.2012.09.001
233. Lagier J-C, Khelaifa S, Alou MT, Ndongo S, Dione N, Hugon P, et al. Culture of previously uncultured members of the human gut microbiota by culturomics. *Nat Microbiol.* (2016) 1:16203. doi: 10.1038/nmicrobiol.2016.203
234. Stepanauskas R, Fergusson EA, Brown J, Poulton NJ, Tupper B, Labonté JM, et al. Improved genome recovery and integrated cell-size analyses of individual uncultured microbial cells and viral particles. *Nat Commun.* (2017) 8:84. doi: 10.1038/s41467-017-00128-z
235. Radajewski S, Ineson P, Parekh NR, Murrell JC. Stable-isotope probing as a tool in microbial ecology. *Nature* (2000) 403:646–9. doi: 10.1038/35001054
236. Berry D, Stecher B, Schintlmeister A, Reichert J, Brugiroux S, Wild B, et al. Host-compound foraging by intestinal microbiota revealed by single-cell stable isotope probing. *Proc Natl Acad Sci USA.* (2013) 110:4720–5. doi: 10.1073/pnas.1219247110
237. Jiang B, Jin N, Xing Y, Su Y, Zhang D. Unraveling uncultivable pesticide degraders via stable isotope probing (SIP). *Crit Rev Biotechnol.* (2018) 38:1025–48. doi: 10.1080/07388551.2018.1427697
238. Vernocchi P, Del Chierico F, Putignani L. Gut microbiota profiling: metabolomics based approach to unravel compounds affecting human health. *Front Microbiol.* (2016) 7:1144. doi: 10.3389/fmicb.2016.01144
239. Berg G, Eberl L, Hartmann A. The rhizosphere as a reservoir for opportunistic human pathogenic bacteria. *Environ Microbiol.* (2005) 7:1673–85. doi: 10.1111/j.1462-2920.2005.00891.x
240. Hardoim PR, van Overbeek LS, Berg G, Pirttilä AM, Compant S, Campisano A, et al. The hidden world within plants: ecological and evolutionary considerations for defining functioning of microbial endophytes. *Microbiol Mol Biol Rev.* (2015) 79:293–320. doi: 10.1128/MMBR.00050-14
241. Iraola G, Forster SC, Kumar N, Lehours P, Bekal S, García-Peña FJ, et al. Distinct *Campylobacter fetus* lineages adapted as livestock pathogens and human pathogens in the intestinal microbiota. *Nat Commun.* (2017) 8:1367. doi: 10.1038/s41467-017-01449-9

242. Paulsen IT, Seshadri R, Nelson KE, Eisen JA, Heidelberg JF, Read TD, et al. The *Brucella suis* genome reveals fundamental similarities between animal and plant pathogens and symbionts. *Proc Natl Acad Sci USA*. (2002) 99:13148–53. doi: 10.1073/pnas.192319099
243. Rosenberg E, Zilber-Rosenberg I. Pathogens as symbionts BT. In: Rosenberg E, Zilber-Rosenberg I. *The Hologenome Concept: Human, Animal and Plant Microbiota* (Cham: Springer International Publishing), 129–49. doi: 10.1007/978-3-319-04241-1_9
244. Postler TS, Ghosh S. Understanding the holobiont: how microbial metabolites affect human health and shape the immune system. *Cell Metab*. (2017) 26:110–30. doi: 10.1016/j.cmet.2017.05.008
245. Toft C, Andersson SGE. Evolutionary microbial genomics: insights into bacterial host adaptation. *Nat Rev Genet*. (2010) 11:465. doi: 10.1038/nrg2798
246. Hevia A, Delgado S, Sánchez B, Margolles A. Molecular players involved in the interaction between beneficial bacteria and the immune system. *Front Microbiol*. (2015) 6:1285. doi: 10.3389/fmicb.2015.01285
247. Quevillon-Cheruel S, Leulliot N, Muniz CA, Vincent M, Gally J, Argentini M, et al. Evf, a virulence factor produced by the *Drosophila* pathogen *Erwinia carotovora*, is an S-palmitoylated protein with a new fold that binds to lipid vesicles. *J Biol Chem*. (2009) 284:3552–62. doi: 10.1074/jbc.M808334200
248. Jetten MSM. The microbial nitrogen cycle. *Environ Microbiol*. (2008) 10:2903–9. doi: 10.1111/j.1462-2920.2008.01786.x
249. Lilburn TG, Kim KS, Ostrom NE, Byzek KR, Leadbetter JR, Breznak JA. Nitrogen fixation by symbiotic and free-living spirochetes. *Science* (2001) 292:2495–8. doi: 10.1126/science.1060281
250. Ngugi DK, Brune A. Nitrate reduction, nitrous oxide formation, and anaerobic ammonia oxidation to nitrite in the gut of soil-feeding termites (*Cubitermes* and *Ophiotermes* spp.). *Environ Microbiol*. (2012) 14:860–71. doi: 10.1111/j.1462-2920.2011.02648.x
251. Igai K, Itakura M, Nishijima S, Tsurumaru H, Suda W, Tsutaya T, et al. Nitrogen fixation and *nifH* diversity in human gut microbiota. *Sci Rep*. (2016) 6:31942. doi: 10.1038/srep31942
252. Morris, BE, Henneberger R, Huber H, Moissl-Eichinger C. Microbial syntrophy: interaction for the common good. *FEMS Microbiol Rev*. (2013) 37:384–406. doi: 10.1111/1574-6976.12019
253. Carbonero F, Benefiel AC, Gaskins HR. Contributions of the microbial hydrogen economy to colonic homeostasis. *Nat Rev Gastroenterol Hepatol*. (2012) 9:504–18. doi: 10.1038/nrgastro.2012.85
254. Schink B. Energetics of syntrophic cooperation in methanogenic degradation. *Microbiol Mol Biol Rev*. (1997) 61:262–80.
255. Chung KT. Inhibitory effects of H₂ on growth of *Clostridium cellobioparum*. *Appl Environ Microbiol*. (1976) 31:342–8.
256. Rey FE, Faith JJ, Bain J, Muehlbauer MJ, Stevens RD, Newgard CB, et al. Dissecting the *in vivo* metabolic potential of two human gut acetogens. *J Biol Chem*. (2010) 285:22082–90. doi: 10.1074/jbc.M110.117713
257. Schmidt TSB, Raes J, Bork P. The human gut microbiome: from association to modulation. *Cell* (2018) 172:1198–215. doi: 10.1016/j.cell.2018.02.044
258. Lucy M, Reed E, Glick BR. Applications of free living plant growth-promoting rhizobacteria. *Anton Van Leeuwen*. (2004) 86:1–25. doi: 10.1023/B:ANTO.0000024903.10757.6e
259. Sachsenröder J, Twardziok SO, Scheuch M, John R. The general composition of the faecal virome of pigs depends on age, but not on feeding with a probiotic bacterium. *PLoS ONE* (2014) 9:e88888. doi: 10.1371/journal.pone.0088888
260. Wu GD, Chen J, Hoffmann C, Bittinger K, Chen Y-Y, Keilbaugh SA, et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science* (2011) 334:105–8. doi: 10.1126/science.1208344

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Ikeda-Ohtsubo, Brugman, Warden, Rebel, Folkerts and Pieterse. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.