

UCSF

UC San Francisco Previously Published Works

Title

β 1-integrin via NF- κ B signaling is essential for acquisition of invasiveness in a model of radiation treated in situ breast cancer

Permalink

<https://escholarship.org/uc/item/02f2h88k>

Journal

Breast Cancer Research, 15(4)

ISSN

1465-5411

Authors

Nam, Jin-Min
Ahmed, Kazi M
Costes, Sylvain
et al.

Publication Date

2013-08-01

DOI

10.1186/bcr3454

Peer reviewed

RESEARCH ARTICLE

Open Access

β 1-integrin via NF- κ B signaling is essential for acquisition of invasiveness in a model of radiation treated *in situ* breast cancer

Jin-Min Nam¹, Kazi M Ahmed², Sylvain Costes², Hui Zhang², Yasuhito Onodera³, Adam B Olshen⁴, Kanako C Hatanaka⁵, Rumiko Kinoshita¹, Masayori Ishikawa⁶, Hisataka Sabe³, Hiroki Shirato¹ and Catherine C Park^{2,7*}

Abstract

Introduction: Ductal carcinoma *in situ* (DCIS) is characterized by non-invasive cancerous cell growth within the breast ducts. Although radiotherapy is commonly used in the treatment of DCIS, the effect and molecular mechanism of ionizing radiation (IR) on DCIS are not well understood, and invasive recurrence following radiotherapy remains a significant clinical problem. This study investigated the effects of IR on a clinically relevant model of Akt-driven DCIS and identified possible molecular mechanisms underlying invasive progression in surviving cells.

Methods: We measured the level of phosphorylated-Akt (p-Akt) in a cohort of human DCIS specimens by immunohistochemistry (IHC) and correlated it with recurrence risk. To model human DCIS, we used Akt overexpressing human mammary epithelial cells (MCF10A-Akt) which, in three-dimensional laminin-rich extracellular matrix (lrECM) and *in vivo*, form organotypic DCIS-like lesions with lumina expanded by pleiomorphic cells contained within an intact basement membrane. In a population of cells that survived significant IR doses in three-dimensional lrECM, a malignant phenotype emerged creating a model for invasive recurrence.

Results: P-Akt was up-regulated in clinical DCIS specimens and was associated with recurrent disease. MCF10A-Akt cells that formed DCIS-like structures in three-dimensional lrECM showed significant apoptosis after IR, preferentially in the luminal compartment. Strikingly, when cells that survived IR were repropagated in three-dimensional lrECM, a malignant phenotype emerged, characterized by invasive activity, up-regulation of fibronectin, α 5 β 1-integrin, matrix metalloproteinase-9 (MMP-9) and loss of E-cadherin. In addition, IR induced nuclear translocation and binding of nuclear factor-kappa B (NF- κ B) to the β 1-integrin promoter region, associated with up-regulation of α 5 β 1-integrins. Inhibition of NF- κ B or β 1-integrin signaling abrogated emergence of the invasive activity.

Conclusions: P-Akt is up-regulated in some human DCIS lesions and is possibly associated with recurrence. MCF10A-Akt cells form organotypic DCIS-like lesions in three-dimensional lrECM and *in vivo*, and are a plausible model for some forms of human DCIS. A population of Akt-driven DCIS-like spheroids that survive IR progresses to an invasive phenotype in three-dimensional lrECM mediated by β 1-integrin and NF- κ B signaling.

Keywords: ductal carcinoma *in situ*, DCIS, integrin, ionizing radiation

Introduction

Ductal carcinoma *in situ* (DCIS) is comprised of cancerous cells that are contained within the milk duct and separated from the stroma by a basement membrane and is associated with risk for developing invasive cancer [1].

With the advent of screening mammography, DCIS represents approximately 20% of all new cases of breast cancer diagnosed in the United States annually. Lumpectomy followed by radiation therapy (RT) is the most common treatment for DCIS, the efficacy of which is supported by randomized trials and meta-analysis demonstrating a reduction in the risk for local recurrence by approximately 50% [2,3]. Although the primary goal of therapy is to prevent invasive recurrence, 50% of all local

* Correspondence: cpark@radonc.ucsf.edu

²Life Sciences Division, Ernest Orlando Lawrence Berkeley National Laboratory, 1 Cyclotron Road, Berkeley, 94720, USA

Full list of author information is available at the end of the article

recurrences after RT are invasive cancer [2]. Relevant experimental models to investigate molecular effects of ionizing radiation (IR) on invasive recurrence, however, have not been established.

Although structurally separated from the stromal breast tissue by a basement membrane, DCIS lesions in humans have the ability to elicit stromal remodeling of the extracellular matrix (ECM); indeed, stromal neoangiogenesis and fibronectin (FN) deposition have been well documented [4-6]. β 1-integrins are critical mediators of normal cell-ECM interactions and have been shown to play a multifaceted role in malignant progression [7]. Others and we have shown that IR results in up-regulation of β 1-integrins in invasive breast cancer, leading to increased cell survival [8-10].

Recently, we turned our attention to other mediators of the acute phase response to IR. It is known that nuclear factor-kappa B (NF- κ B) is a pleiotropic regulator of many genes involved in inflammation, growth regulation and apoptosis, and IR [11-13]. In fact, several reports place integrins upstream of NF- κ B [14-16]. We recently showed that upon IR exposure in breast cancer cells, NF- κ B binds directly to the β 1-integrin promoter region, resulting in increased β 1-integrin transcripts and radioresistance [17]. Here, we verified the importance of NF- κ B regulation of β 1-integrin post-IR in the context of Akt-driven progression.

In the present study, we show that phosphorylated-Akt (p-Akt) is up-regulated in clinical DCIS specimens and is associated with recurrent disease. To investigate the possible molecular mechanisms of IR on DCIS, we used active Akt-overexpressing human mammary epithelial cells, MCF10A-Akt, in three-dimensional IrECM cultures as a model of DCIS, which we validated *in vivo*. When propagated in three-dimensional IrECM, the MCF10A-Akt cells recapitulate an organotypic duct-like structure that retains basal polarity, but with lumina expanded by pleiomorphic cells resembling human DCIS [18]. DCIS-like MCF10A-Akt structures in three-dimensional IrECM show a significant increase in apoptosis in response to IR, preferentially in the luminal compartment. To determine whether surviving cells remained viable after IR, we selected and repropagated them in three-dimensional IrECM. Strikingly, we observed the emergence of a malignant phenotype in a sub-population of survivors, with increased β 1-integrin expression, matrix metalloproteinase-9 (MMP-9) and invasive activity. In addition, among the malignant population, IR induced nuclear translocation and binding of NF- κ B p65 to the β 1-integrin promoter region, associated with up-regulation of β 1-integrins. Inhibition of NF- κ B translocation to the nucleus or inhibition of β 1-integrin signaling abrogated the emergence of the invasive phenotype. These results indicate that regulation of β 1-integrin signaling via NF- κ B may play an important

role in the emergence of invasive disease after radiation treatment of Akt-driven DCIS-like lesions.

Methods

Tissue specimens

Clinical specimens were obtained from 24 patients with pure DCIS, who were treated at the Hokkaido University Hospital from 1998 to 2008. Patients underwent breast-conserving surgery followed by external beam fractionated radiotherapy to the whole breast. Among the 24 patients, five had ipsilateral invasive breast tumor recurrence within five years. This study was approved by the Institutional Review Board of Hokkaido University Hospital (010-0203). The requirement for written consent was waived by our institutional board according to the Ethical Guidelines for Clinical Studies of the Japanese Ministry of Health, Labor and Welfare.

Immunohistochemistry and pathologic scoring of human DCIS tissues

Immunohistochemistry (IHC) of human DCIS tissues was performed on 4 μ m-thick formalin-fixed paraffin-embedded serial sections. Immunohistochemical staining of p-Akt was performed by using the CSAII Biotin-free Tyramide Signal Amplification System (DAKO, Tokyo, Japan) according to the manufacturer's protocol. Each slide was deparaffinized in xylene and dehydrated through graded alcohols, and processed for antigen retrieval by ethylenediaminetetraacetic acid (EDTA) (pH 9.0) at 95°C for 40 minutes. Endogenous peroxidase was blocked by 3% hydrogen peroxidase at room temperature for 10 minutes and then blocked by serum-free protein in buffer for 10 minutes. Primary antibody against p-Akt (1:50, Cell Signaling Technology, Danvers, MA, USA) was incubated overnight at 4°C. Slides were washed and then followed by sequential incubation for 15 minutes with anti-rabbit immunoglobulins-horseradish peroxidase (HRP) (1:200), fluorescyl-tyramide hydrogen peroxide and anti-fluorescein-HRP. For β 1-integrin staining, EnVision™ system (DAKO) was used. After deparaffinization, the slides were treated with antigen retrieval reagent (pH 9.0) at 95°C for 40 minutes. Slides were washed and incubated in 3% H₂O₂ and then blocked. After rinsing, the sections were incubated with primary antibody against β 1-integrin (1:150) overnight at 4°C. Antibody detection was performed using the EnVision™ system. The color was developed with 3, 3'-diaminobenzidine tetrahydrochloride (DAB)/hydrogen peroxide. Each slide was counterstained with hematoxylin. Blinded samples were reviewed by a pathologist and scored for nuclear grade and Van Nuys classification. IHC was scored based on the intensity of signal (0 = none, 1 = light, 2 = moderate, 3 = heavy) and the percentage of positive cells (0 = <10%, 1 = 10% to 25%, 2 = 25% to 50%,

3 = >50%). All variables were made binary before analysis. All patients had at least five years of follow-up, and thus, we estimated five-year recurrence-free survival rates. For the other variables, if they were not already binary, such as estrogen receptor (ER) status, a threshold was used that led to a balance in the counts of the two groups. Associations between variables were assessed using Fisher's exact test. Tests with *P*-values less than 0.05 were considered significant.

***In vivo* study**

To mimic human DCIS, intraductal transplantation was performed as described previously [19]. Adult 9-week old virgin female SCID mice were anesthetized, a Y-incision was made on the abdomen, and then the nipple of the 4th inguinal mammary gland was identified and transected. Forty thousand MCF10A-Akt cells in 2 to 4 μ l medium containing 0.1% trypan blue were injected *via* the cleaved nipple of mouse mammary ducts. A 50 μ l capacity Hamilton syringe with a 30-gauge blunt-ended 1/2-inch needle was used to inject the cells. After the injection, the skin was closed with wound clips. A sterile tamoxifen pellet (Innovative Research of America, Sarasota, FL, USA) was inserted subcutaneously. At the end of the experiments, mammary glands were harvested, and paraffin or frozen blocks were made. All animal experimental procedures were approved by the Lawrence Berkeley National Laboratory Animal Welfare and Safety Committee.

Immunohistochemistry and immunofluorescence on *in vivo* tumor sections

IHC of thin sections of mouse mammary gland tissues containing human-like DCIS lesions was performed using standard avidin-biotin-peroxidase (ABC) methods. Each slide was baked at 60°C for 30 minutes, dewaxed in xylene and rehydrated through graded alcohols. For β 1-integrin staining, slides were incubated with proteinase K at 37°C for 10 minutes. After washing, slides were microwaved for 10 minutes in 10 mmol/L sodium citrate (pH 6.0) and cooled for 30 minutes. Each section was blocked with 10% normal rabbit serum for β 1-integrin or normal goat serum for p-Akt and cleaved caspase-3 staining for 15 minutes. Primary antibody against β 1-integrin (1:50, Millipore, Billerica, MA, USA) was incubated for one hour. Primary antibody against p-Akt (1:50, Cell Signaling Technologies) or cleaved caspase-3 (1:300, Cell Signaling Technologies) was incubated overnight at 4°C. Antibody detection was performed using an avidin-biotinylated enzyme. The coloring reaction was performed with DAB. Each section was counterstained with hematoxylin. For immunofluorescence (IF) staining of Ki-67, each slide was baked and deparaffinized in xylene, and rehydrated through a graded series of ethanol and then microwaved for 10 minutes in 10 mmol/L sodium

citrate (pH 6.0). After cooling for 20 minutes, the sections were blocked by 10% goat serum for 10 minutes. Primary antibody against Ki-67 (1:500, Vector laboratories, Burlingame, CA, USA) was incubated overnight at 4°C. Slides were washed and incubated with secondary antibody for 40 minutes. Nuclei were counter stained with 4', 6-diamino-2-phenylindole (DAPI; Sigma-Aldrich, St. Louis, MO, USA). The images of IHC and IF were acquired by using a Zeiss Axioskop inverted microscope.

Cell culture

MCF10A cells expressing ER-Akt (MCF10A-Akt), a kind gift from Dr. Jayanta Debnath (University of California in San Francisco, UCSF) were cultured in (D)MEM/F12 supplemented with 5% horse serum (Life technologies, Carlsbad, CA, USA); 20 ng/ml epidermal growth factor (EGF; Roche, Basel, Schweiz); 10 μ g/ml insulin (Sigma-Aldrich); 0.5 μ g/ml hydrocortisone (Sigma-Aldrich); 100 ng/ml cholera toxin (Sigma-Aldrich) and penicillin-streptomycin (Life technologies) [20]. For ER-Akt activation, 4-hydroxytamoxifen (4-HT; Sigma-Aldrich) was added to the culture medium on day two of three-dimensional culture, and EtOH was used as a control.

Three-dimensional lrECM culture

For three-dimensional lrECM cultures, MCF10A-Akt cells were plated on top of commercially available growth factor-reduced Basement Membrane Extract (BD Biosciences, San Jose, CA, USA) as described previously [21]. MCF10A-Akt cells were cultured on three-dimensional lrECM for 12 days in the presence of EtOH or 1 μ M 4HT. At Day 12, cultures were exposed to Sham, 2 or 8 Gy X-ray and harvested on Day 15. For the recurrence model, the colonies were isolated and dissociated from three-dimensional lrECM to make single cells using 0.05% trypsin-EDTA, and expanded on two-dimensional plastic on Day 15. Single cells were re-plated on top of three-dimensional lrECM and propagated for 12 additional days.

Lysis from three-dimensional lrECM

To release cells from three-dimensional lrECM, cultures were first treated with ice-cold 5 mM EDTA/PBS on ice and then cells were lysed in 1% radioimmunoprecipitation assay (RIPA) buffer (1% Noidet P-40, 150 mM NaCl, 50 mM Tris-HCl (pH 7.4), 5 mM EDTA, 1% Na-deoxychorate, 0.1% SDS, 1 mM Na₃VO₄, 10 μ M Na₂MoO₄, and protease inhibitor cocktail (Millipore)). After sonication, the solution was centrifuged and the supernatant was collected.

Immunoblotting

Cell lysates were aliquoted onto Novex[®] 4-20% Tris-Glycine gels (Life technologies) in equal amounts and

separated using 110 voltage current. Protein bands were transferred onto a polyvinylidene fluoride (PVDF) membrane, and blots were blocked with 5% skim milk/TBST (Tris-buffered saline Tween-20; 25mM Tris-HCl (pH7.4), 120 mM NaCl, 3 mM KCl and 0.1% Tween 20). Blots were probed with primary antibody (1:100 to 1:2,000 dilution in 3% skim milk/TBST) overnight at 4°C. After washing three times with TBST for 10 minutes, membranes were incubated with secondary antibody for one hour, then washed with TBST, and exposed by enhanced chemiluminescence.

Immunostaining

Cells from three-dimensional cultures were fixed with 4% paraformaldehyde for 15 minutes, washed three times with glycine/PBS for 10 minutes and blocked with blocking buffer (10% goat serum (Life technologies), 1% Goat F(ab')₂ anti-mouse antibody (Life technologies) in IF wash buffer (0.05% NaN₃, 0.1% BSA, 0.2% Triton-X 100 and 0.05% Tween 20 in PBS)) for 1.5 hours in a humidified chamber. After removing blocking buffer, samples were incubated with primary antibody in blocking buffer overnight at 4°C and then washed three times with IF wash buffer, followed by incubating with Alexa Fluor 488-conjugated secondary antibody (Life technologies) for 40 minutes in a dark humidified chamber and then washed three times with IF wash buffer for 20 minutes. Nuclei were counterstained with DAPI, washed twice with PBS for 10 minutes and mounted. Images were acquired using a Zeiss Axioskop inverted microscope or a Nikon A1R confocal microscope system.

Antibodies

The following antibodies were used: anti-β1-integrin, clone 18 (BD Biosciences); anti-β1-integrin, AIB2 (Sierra Biosource, Morgan Hill, CA, USA); anti-α5-integrin (Millipore); anti-α6-integrin, NKI-GoH3 (Millipore); anti-FN, IST-4 (Sigma); anti-FN EDA+ (Abcam, Cambridge, England); anti-cleaved caspase-3 (Cell signaling Technology); anti-NF-κB p65 (Immuno-Biological Laboratories Co., Ltd., Gunma, Japan); anti-histone H1 (abcam) anti-β-actin (Sigma-Aldrich); ECL™ anti-mouse immunoglobulin G (IgG), HRP linked whole antibody (from sheep), NA931V (GE Healthcare, Buckinghamshire, UK); ECL™ anti-rabbit IgG, HRP linked whole antibody (from donkey), NA934V (GE Healthcare).

Matrix degradation assay

Glass-bottomed dishes (MatTek, Ashland, MA, USA) were coated with DQ-gelatin from pig skin fluorescein conjugate (Invitrogen), which was subsequently cross-linked with 0.5% glutaraldehyde on ice. Cells from three-dimensional IrECM cultures at day 30 were replated onto the coated dishes, incubated for 24 hours and then fixed

with 4% paraformaldehyde for 15 minutes. The number of cells positive for matrix degradation was counted from 100 cells.

Zymography

Gelatin gels for the zymography assay were purchased from Invitrogen and handled according to the manufacturer's instructions. Cells were repropagated in three-dimensional IrECM cultures for five days and the medium was replaced by (D)MEM/F12 supplemented with additives (20 ng/ml EGF; 10 μg/ml insulin; 0.5 μg/ml hydrocortisone; 100 ng/ml cholera toxin). After 24 hours, the medium absent of serum was collected and denatured in SDS buffer under non-reducing conditions, and run on a Novex® Zymogram gel. After electrophoresis, the gel was incubated in Zymogram renaturing buffer and then equilibrated in Zymogram developing buffer.

Matrigel chemoinvasion assay

The matrigel chemoinvasion assay was performed using Biocoat Matrigel invasion chambers (BD Biosciences) as described previously [22]. Briefly, 5×10^4 cells were seeded onto the upper wells of 24-well chambers in the absence of serum, and the lower wells were filled with culture medium. After incubation for 18 hours, cells that migrated out onto the lower surface of the membranes were fixed with 4% paraformaldehyde at room temperature for 15 minutes. These cells were stained with 1% crystal violet and analyzed using AxioCam (Carl Zeiss, Jena, Germany). Data were collected from three independent experiments.

Apoptosis assay

Apoptosis was detected using terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling (TUNEL) as described previously [10]. Samples from three-dimensional IrECM were fixed with 4% paraformaldehyde and permeabilized in 0.1% Triton X-100 in 0.1% sodium citrate. After washing with PBS, cells were incubated in TUNEL reaction mixture (In Situ Cell Death Detection Kit, Roche) at 37°C for 60 minutes.

Image acquisition, processing and analysis

Cells were viewed and imaged using a Zeiss Axio Observer Z1 automated microscope (Carl Zeiss). Images were acquired using a high NA Zeiss plan-apochromat 20X (NA of 0.8) and a sensitive scientific-grade EM-CCD camera (AxioCam, Carl Zeiss). In order to acquire data for a large number of colonies, minimize acquisition time and simplify analysis, the centers of individual acini were marked with the 'Mark and Find' function of Axiovision and images for all acini were acquired at once. In order to compare immunostaining intensity, for a given antibody, all images across treatments were acquired with the same exposure time, making sure images were not

saturated. Each treatment group contained 100 to 300 acini in duplicate experiments. Image processing was performed under Matlab (MathWorks Inc, Natick, MA, USA) and DIP image (image processing toolbox for Matlab, Delft University of Technology, The Netherlands). Statistical tests were done with the Statistics module of Matlab. Results of three-dimensional studies are expressed as mean \pm SE. Data were analyzed by Student's *t* test. *P* values of < 0.05 were considered significant. Cells positive for cleaved caspase-3 were identified by setting a constant threshold across all treatments above which pixels were considered positive if they were also overlapping with the nuclear DAPI area. The percent of positive cells was computed by dividing the positive area with the nuclear DAPI area. All imaging parameters and methods used in this manuscript have recently been described [23].

Results

Phosphorylated-Akt and β 1-integrin are up-regulated in human DCIS specimens and correlate with disease recurrence

Oncogenic Akt activation has been implicated in breast cancer progression and is also up-regulated in clinical human DCIS tissue specimens [24]. To investigate the clinical importance of p-Akt and β 1-integrin levels in human DCIS, we asked whether their expression level was correlated with local recurrence after breast conserving surgery and RT in a limited cohort of patients. First, we confirmed that both p-Akt and β 1-integrin were indeed present in human DCIS specimens (Figure 1A and Additional file 1A). Among the 24 patients, 22 (91.7%) scored positively (1 to 3 versus 0) for p-Akt intensity and percentage. Importantly, 17 (70.8%) cases had a percentage score of 3, or the highest level of expression (Figure 1B). We then analyzed p-Akt staining for association with local recurrence within five years (Figure 1C). Among the five recurrent cases, all five (100%) had a percentage score of 3 compared with the non-recurrent cases (63.2%). For p-Akt intensity, moderate to high expression (2 to 3 versus 0 to 1) was strongly associated with recurrent cases (80%), whereas 0 to 1 scores are dominant in non-recurrent cases (63.2%) (Figure 1C; odds ratio (OR) = 6.32, *P* = 0.11, Fisher's exact test). Since nuclear grade is known to be one of the most important clinical factors associated with disease recurrence in DCIS, we also tested for the association between p-Akt expression and nuclear grade. Among the 22 scorable cases, 12 cases are nuclear grade 3, and all 12 had a moderate to high percentage (2 or 3) of p-Akt.

We also examined β 1-integrin expression, comparing recurrent versus non-recurrent cases [see Additional file 1]. We found that a high percentage of β 1-integrin expression is associated with recurrent cases compared

with non-recurrent cases (40% versus 5.3%, OR = 10.8, *P* = 0.09). These results are consistent with our previous study in early stage invasive breast cancer where we showed that β 1-integrin expression was associated with more aggressive disease and significantly reduced overall and disease-free survival [25]. These data suggest that p-Akt and β 1-integrin are important in identifying aggressive forms of DCIS that is more likely to recur following treatment.

Akt-overexpressing MCF10A cells formed DCIS-like structures in three-dimensional IrECM cultures and *in vivo*

We sought to investigate the effect of radiation and its underlying molecular mechanisms in DCIS. As human DCIS lesions were found to up-regulate p-Akt, we modeled DCIS by propagating MCF10A human mammary epithelial cells with inducible active ER-Akt [18,24]. Typically, MCF10A cells form a hollow lumen in three-dimensional IrECM [20], recapitulating normal *in vivo* ductal structures. However, cells induced to express activate Akt by adding 4-HT formed overgrown large colonies containing lumen filled with atypical appearing cells (Figure 2A and 2B), emulating premalignant *in situ* lesions. Notably, IF of α 6-integrin indicates that the basal polarity is not disrupted, confirming the non-invasive structure (Figure 2A, inset).

To determine the validity of MCF10A-Akt cells as a model for human DCIS in three-dimensional IrECM, we recapitulated the DCIS-like structures *in vivo* (Figure 2C) using an established intraductal transplantation method [19]. The MCF10A-Akt cells were directly injected into the mammary duct of female SCID mice via a cleaved nipple and mice were monitored for up to 10 weeks. Using H & E staining from sections of harvested mammary glands, we detected intraductal tumors generated from MCF10A-Akt cells that represented human DCIS-like lesions (Figure 2D). Confirming that the molecular phenotype of these lesions emulated that observed in three-dimensional IrECM, co-overexpression of p-Akt and β 1-integrin was detected by IHC (Figure 2D). The expression of cleaved caspase-3, an apoptosis marker, was negatively correlated with proliferation, measured by Ki-67 nuclear antigen, and negatively correlated with active Akt positive regions (Figure 2D). These results show that MCF10A-Akt cells form human DCIS-like structures *in vitro* in three-dimensional IrECM and *in vivo* that are proliferative and have high β 1-integrin expression. Our *in vivo* data further validated the use of the MCF10A-Akt cells in the three-dimensional IrECM culture model as a surrogate for testing the effects of IR on human DCIS. Moreover, the *in vitro* system would allow us to further investigate the biology of recurrence following IR treatment of DCIS, which to date has not been adequately addressed.

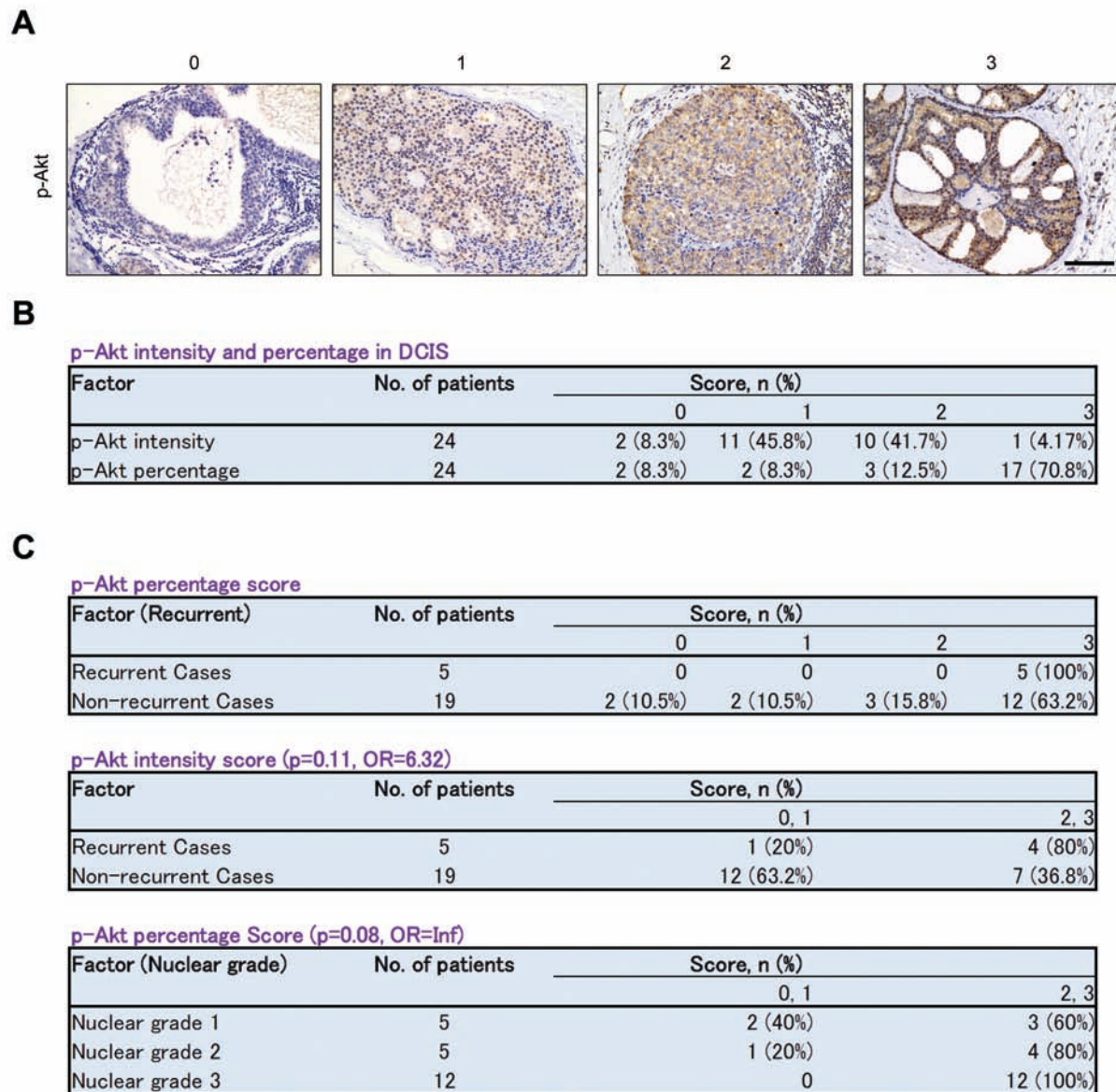
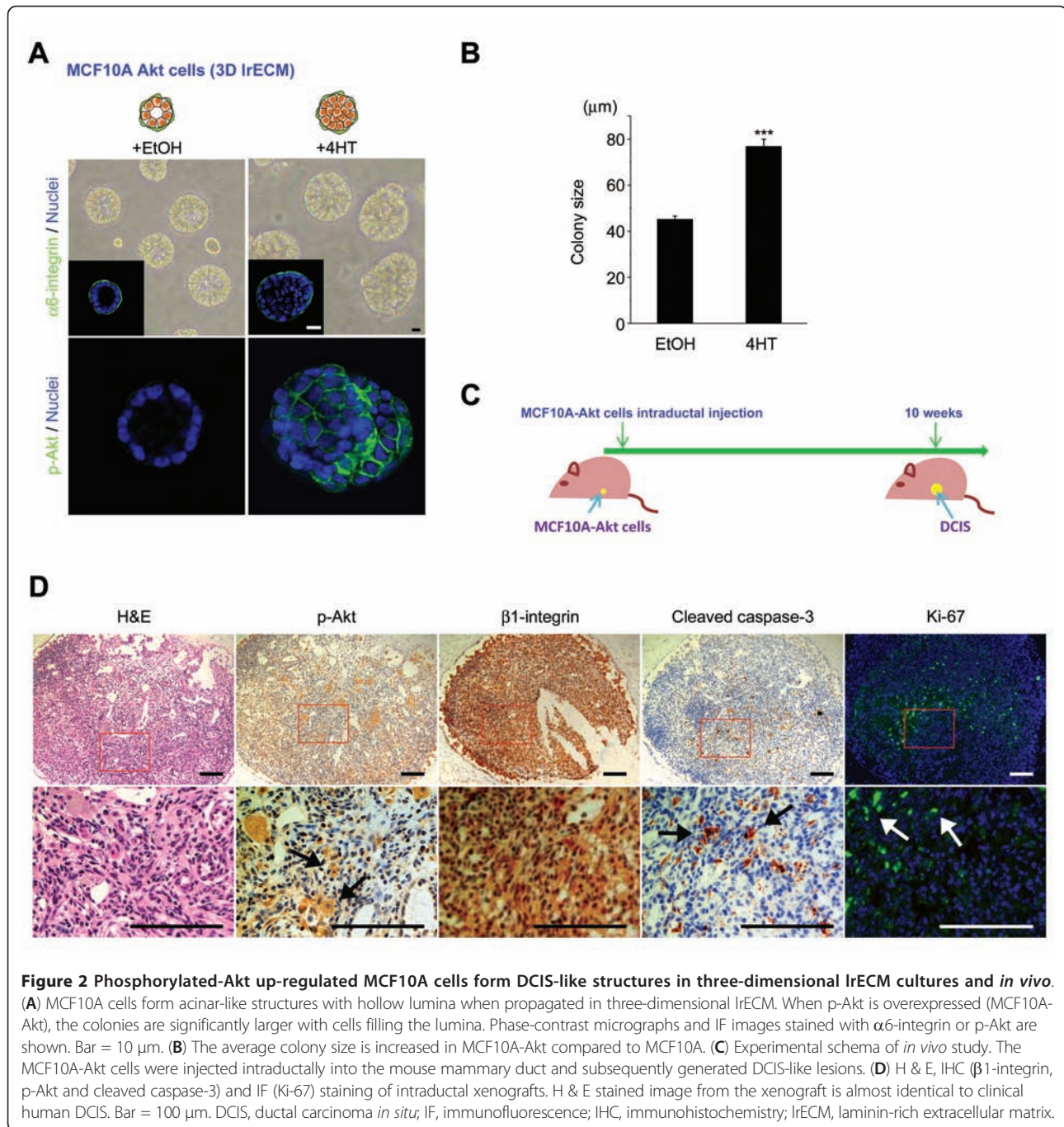


Figure 1 Phosphorylated-Akt is up-regulated in clinical DCIS specimens. (A) IHC of p-Akt in human DCIS specimens. Formalin-fixed, paraffin-embedded DCIS sections from 24 patients were stained with p-Akt. Phosphorylated-Akt intensity score: 0 = none, 1 = light, 2 = moderate, 3 = heavy. Bar = 100 μ m. (B) Intensity and percentage expression pattern for p-Akt in human DCIS specimens. Phospho-Akt percentage score: 0 = <10%, 1 = 10% to 25%, 2 = 25% to 50%, 3 = >50%. (C) Up-regulation of p-Akt is associated with recurrent disease in human DCIS. DCIS, ductal carcinoma *in situ*; IHC, immunohistochemistry.

IR induces apoptosis in an Akt-overexpressing model of human DCIS in three-dimensional IrECM

Using the three-dimensional IrECM culture model, we next investigated the effect of IR on DCIS. To test whether radiation induces apoptosis in the DCIS model, we irradiated the three-dimensional cultured MCF10A-Akt cells at Day 12 and performed IF with anti-cleaved

caspase-3 (Figure 3A). IR induced apoptosis of MCF10A-Akt cells in three-dimensional IrECM in a dose-dependent fashion, detected by cleaved caspase-3 (Figure 3B and Additional file 2). We further quantitated these results using high content image analysis which showed that cleaved caspase-3 positive cells increased with increasing radiation doses (Figure 3C). Interestingly,

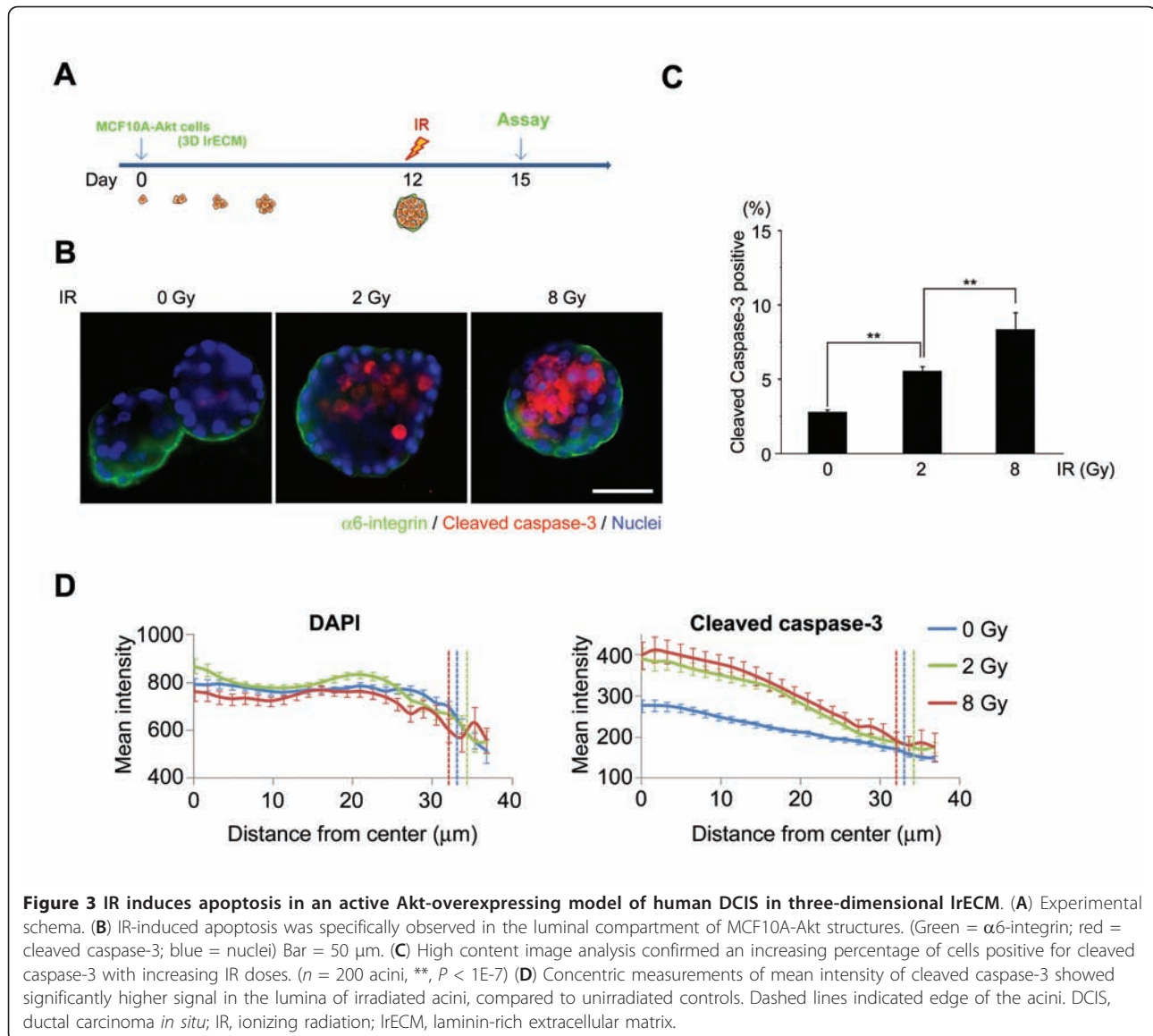


apoptotic cells were significantly increased in lumenally located cells compared to basal cells that were in contact with ECM (Figure 3B and 3D).

An invasive phenotype with high β1-integrin expression emerged from a sub-population of surviving cells post-IR in three-dimensional IrECM

Invasive recurrence remains a significant problem following breast-conserving surgery and radiation for DCIS.

The nature of recurrence remains elusive, and there are currently no models to investigate this aspect of the disease. Thus, we sought to develop a model to investigate the viability of DCIS-like cells that survive significant doses of IR. MCF10A-Akt cells were cultured in three-dimensions for 12 days, followed by sham or 8 Gy IR (Figure 4A). After three days, the MCF10A-Akt cells were released from three-dimensions, dissociated to single cells, and then repropagated in three-dimensional



IrECM (Figure 4A and 4B). Surprisingly, after 12 additional days of culture (or Day 30 of total number of days), we observed a subset of the culture population that exhibited an invasive phenotype (MCF10A-Akt-invasive) (Figure 4B, f, h). Alpha6-integrin showed a disruption in basal polarity with an increase in $\beta 1$ -integrin expression (Figure 4B, j, l). In contrast, the polarity of sham irradiated cells was retained on the second three-dimensional cultures (day 30, Figure 4B, e, g, i) similar to the first three-dimensional cultures (day 15, Figure 4B, a). Matrigel chemoinvasion was increased by 4.57-fold post-IR (Figure 4C), concomitant with an increase in MMP-9 in the conditioned medium of IR treated MCF10A-Akt-invasive cells (Figure 4D). Matrix degradation activity measured by DQ-gelatin matrix was increased by 22-fold post-IR (Figure 4E). Importantly, we also observed the emergence of similar invasive colonies

post-IR using a similar MCF10A-NeoT cell model, validating this phenomenon [see Additional file 3].

FN and $\alpha 5\beta 1$ -integrin are required for invasive progression in MCF10A-Akt cells post-IR

We have previously shown that signaling downstream of $\alpha 5\beta 1$ -integrin and its ligand FN is important for breast cancer cell survival after radiation [10]. In addition to this, we have shown that the expression of $\alpha 5\beta 1$ -integrin, FN and EDA+FN, the FN variant expressing during embryogenesis and wound healing, is up-regulated in highly aggressive metastatic breast cells [10]. In the current study, we investigated whether $\alpha 5\beta 1$ -integrin and FN signaling is involved in the invasive tumor colonies post-IR on MCF10A-Akt in three-dimensional IrECM. At Day 30, the protein expression of $\alpha 5$ -integrin was

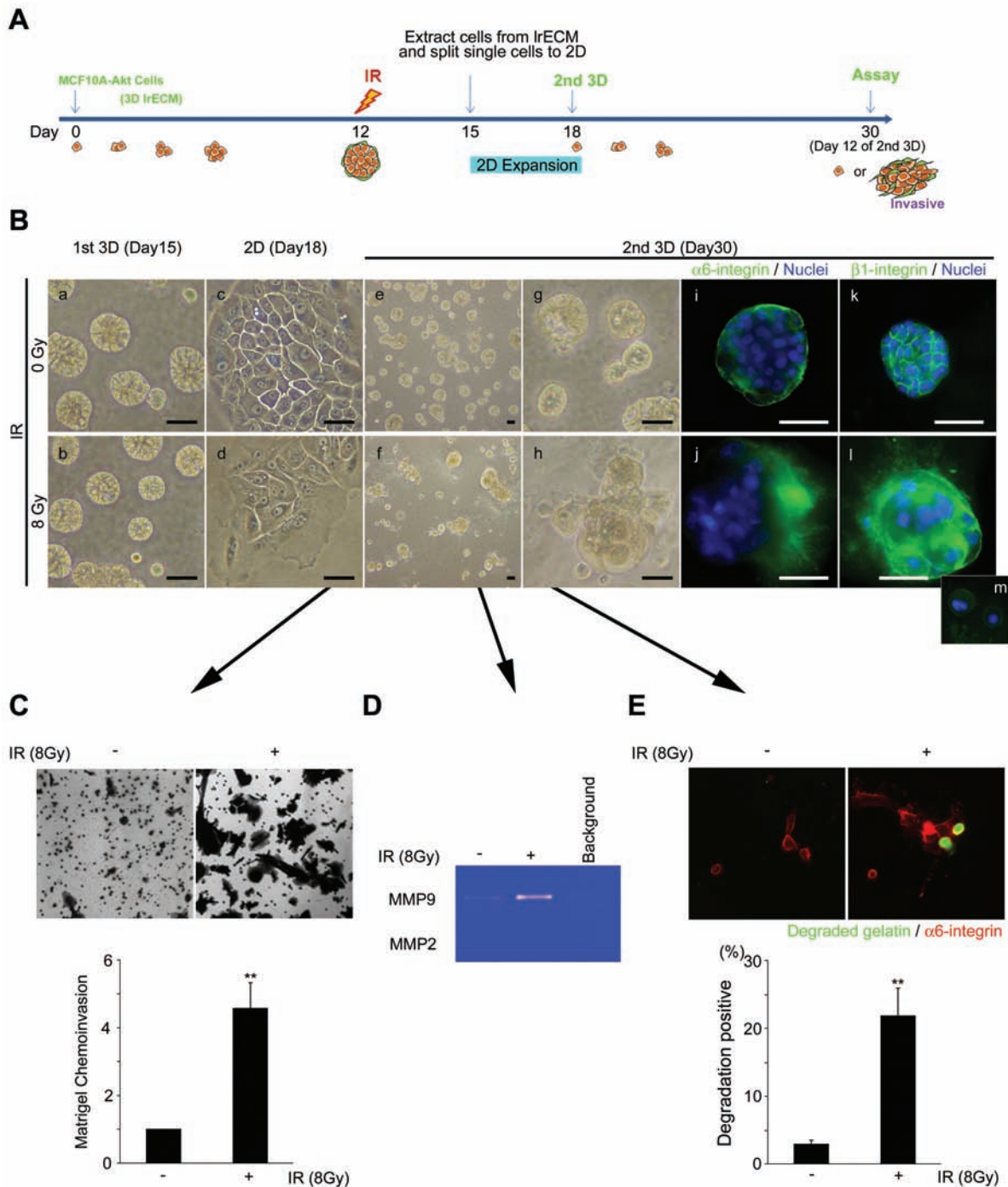


Figure 4 An invasive phenotype emerged from a sub-population of cells surviving post-IR in three-dimensional IrECM. (A) Experimental schema of the recurrence model. At Day 12, cultures were exposed to Sham or 8 Gy IR. On Day 15, the colonies were taken out of three-dimensional IrECM, dissociated to make single cells, and expanded on two dimensional. Single cells were re-plated on three-dimensional IrECM and propagated until Day 30 (12 additional days). **(B)** Phase-contrast micrographs show that a distinct phenotype emerged by Day 30 of culture. Bar = 50 μ m. IF images show α 6-integrin or β 1-integrin (green). Bar = 50 μ m. **(C)** Invasive activity of MCF10A-Akt cells post-IR was quantified using invasion chambers. Graphical representation of the invasive cell numbers were normalized with control, non-irradiated cultures ($n = 3$; **, $P < 0.01$). **(D)** Gelatin zymography shows that MMP-9 secretion was increased in culture medium of IR-treated MCF10A-Akt. **(E)** Matrix degradation activity was confirmed by fluorescently labeled DQ-gelatin matrix. Degraded gelatin is shown in green (22% \pm 7 invasive cells versus 3% \pm 1; $n = 3$; **, $P < 0.01$). DCIS, ductal carcinoma *in situ*; IF, immunofluorescence; IR, ionizing radiation; IrECM, laminin-rich extracellular matrix; MMP-9, matrix metalloproteinase-9.

highly up-regulated and E-cadherin was down-regulated in the irradiated MCF10A-Akt cells in three-dimensional IrECM (Figure 5A). In addition, both total and EDA+FN were higher in the conditioned medium of irradiated cells versus control (Figure 5B).

Since β 1-integrin was highly expressed in the invasive colonies and is a known driver of invasion, we tested whether inhibiting β 1-integrin affected the ability of surviving cells post-IR to acquire invasive features. We found that β 1-integrin inhibitory antibody, AIB2, suppressed the progression of malignancy characterized by matrigel chemoinvasion activity and cancer cell survival after radiation treatment (Figure 5C, D and 5E). Beta1-integrin inhibition induced increased apoptosis (Figure 5D), and abrogated chemoinvasion activity (Figure 5E). We also found that α 5 β 1-integrin inhibitory antibody could suppress the invasive activity (Figure 5F), indicating that α 5 β 1-integrin heterodimer plays a specific role.

NF- κ B activation is involved in the emergence of the invasiveness in surviving MCF10A-Akt cells post-IR

Among the possible molecular mechanisms involved in invasive recurrence downstream of FN and β 1-integrin, our findings pointed to the potential role of NF- κ B. NF- κ B has been reported to induce pro-MMP9 expression downstream of FN and α 5 β 1-integrin [26], and we recently showed its regulation of β 1-integrin via binding to the β 1-integrin promoter post-IR [17]. Thus, we hypothesized that increased FN- α 5 β 1-integrin signaling via NF- κ B/p65 activation could facilitate malignant progression post-IR on the three-dimensional culture model of DCIS. Figure 6A displays a three-fold induction of nuclear NF- κ B/p65 by subcellular fractionation in MCF10A-Akt recurrent cells post-IR. IF also showed the nuclear localization of NF- κ B/p65 in recurrent MCF10A-Akt cells post-IR (Figure 6B). In addition, the binding of NF- κ B to the β 1-integrin promoter region was significantly increased (Figure 6C). Treatment of MCF10A-Akt cells with NF- κ B inhibitor, JSH-23, fully inhibited the IR-induced MCF10A-Akt-invasion activity (Figure 6D), suppressed formation of the invasive colonies (Figure 6E) and was associated with down-regulation of α 5 β 1-integrin (data not shown). Taken together, these data suggest that feedback regulation of β 1-integrin and NF- κ B plays an important role in the emergence of invasive recurrence post-IR in our model of DCIS.

Discussion

RT is commonly used in combination with breast conserving surgery for DCIS to reduce the risk of local recurrence. However, for many women, the risk of potentially life-threatening invasive recurrence remains a significant problem. In the present work, we used MCF10A-Akt cells in three-dimensional IrECM culture to model

human DCIS and to investigate the molecular mechanisms involved in the IR effect on the emergence of invasive disease following treatment.

As others have previously reported [18], we also show that MCF10A-Akt cells form DCIS-like structures with an intact basement membrane and lumina expanded by pleiomorphic cells, reminiscent of DCIS *in vivo*. This model has clinical relevance since Akt has been reported to be overexpressed in human DCIS [24]. Furthermore, we examined p-Akt in several human DCIS cases that either recurred or had not recurred within five years after surgery and RT. Our data show that p-Akt is moderately to highly expressed in 17 of 24 (71%) of DCIS cases, and that five of five (100%) recurrent cases had a high percentage expression of p-Akt; and finally, among cases with the highest nuclear grade, p-Akt was moderately to highly expressed in all cases, providing further support for its role in disease progression. Using the active Akt-overexpressing DCIS model in three-dimensional IrECM, we found that, in response to IR, luminal cells preferentially undergo apoptosis compared to basally located cells. This is the first report, to our knowledge, of the effect of IR in a three-dimensional cell culture model of DCIS. It is well established that contact with ECM confers relative resistance to IR-induced death [27-30], and this has been corroborated in our own previous work [9]. However, the implications of the relative resistance of basally located cells that have more robust mechanotransduced signaling downstream of β 1-integrin post-IR is not known.

To further investigate the fate of cells that remain viable post-IR, we selected for surviving MCF10A-Akt cells at Day 15 of culture. Strikingly, after expanding surviving cells in two dimensions, and repropagating these cells back into three-dimensional IrECM, we observed the growth of malignant colonies characterized by invasive activity by Day 30 of culture. This indicated that a subpopulation of viable cells progresses to invasive cancer; in addition, we believe that the disruption of the basal polarity of the DCIS and the expansion process from single DCIS cells were important in facilitating progression to invasive cancer in this model. In the clinical situation, our data implicate a role for β 1-integrin signaling and NF- κ B regulation in facilitating progression among even single cells that survive IR. In addition to the DCIS-Akt three-dimensional model, we confirmed the emergence of an invasive phenotype with up-regulation of FN/ α 5 β 1-integrin post-IR using a similar non-invasive MCF10A-NeoT cell model, indicating that the phenomenon was not dependent on a specific oncogenic driver.

The invasive colonies that emerged post-IR by Day 30 were characterized by diffuse α 6-integrin distribution, high α 5 β 1-integrin expression, increased MMP9, loss of E-cadherin, and the ability to invade. Beta1-integrin has

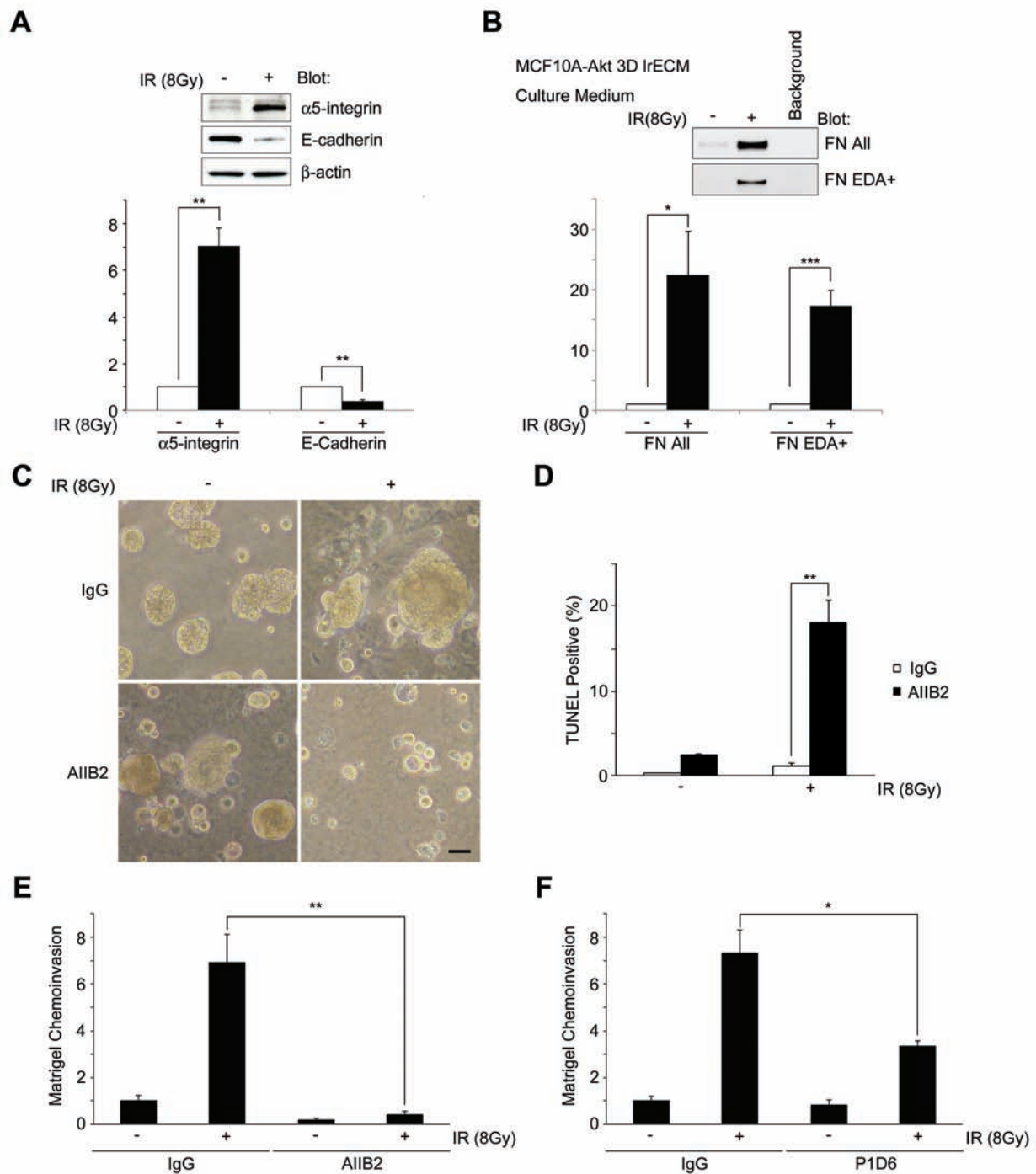


Figure 5 β 1-integrin signaling is targeted to suppress invasive recurrence post-IR in MCF10A-Akt cells in three-dimensional IrECM. (A) On Day 30 cultures, up-regulated α 5 β 1-integrin and down-regulated E-cadherin expression observed on the post-IR cultures. Columns, mean intensity ($n = 3$; **, $P < 0.01$). (B) Up-regulation of FN and EDA+FN was observed in culture medium of cells post-IR. Columns, mean intensity ($n = 5$; *, $P < 0.05$; ***, $P < 0.001$). (C) Phase-contrast images. IR-induced invasive phenotype was abrogated by AlIB2 compared to IgG. The antibodies were added from day 0 of the second three-dimensional cultures. Bar = 50 μ m. (D) Apoptosis was measured by TUNEL-positive cells in AlIB2-treated cultures -/+ IR (mean = 18.1% \pm 3.9, $P < 0.01$). (E-F) Matrigel chemo-invasion was significantly increased in the surviving cells post-IR, and inhibited by β 1-integrin (AlIB2, five-fold, $P < 0.01$) or α 5-integrin (P1D6, two-fold, $P < 0.05$) inhibitory antibodies. DCIS, ductal carcinoma *in situ*; FN, fibronectin; IgG, immunoglobulin G; IR, ionizing radiation; IrECM, laminin-rich extracellular matrix; TUNEL, terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling.

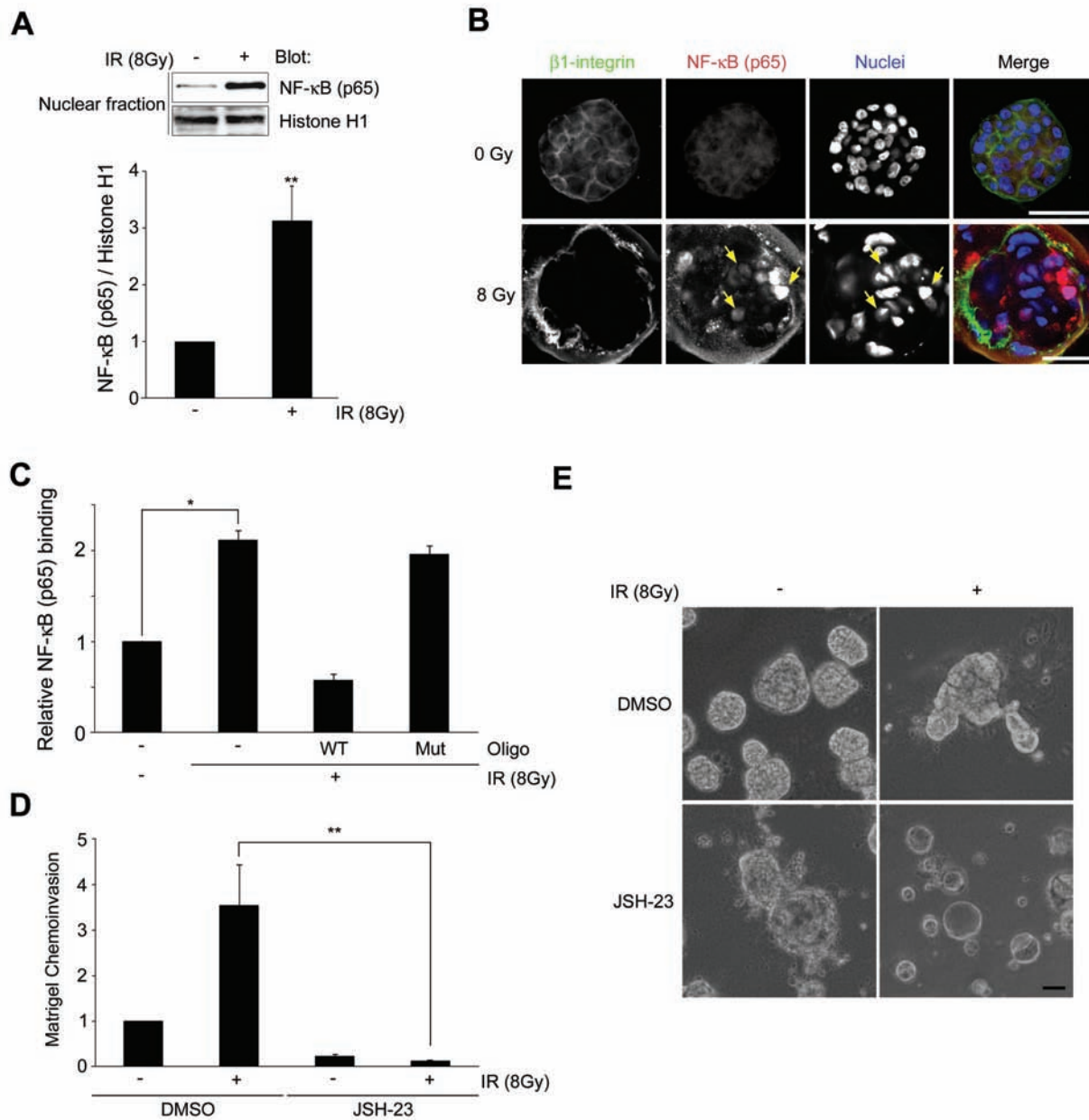


Figure 6 Invasive phenotype emerged in a sub-population of irradiated MCF10A-Akt cells are associated with nuclear translocation of NF-κB. (A) On Day 30 cultures, immunoblotting of nuclear fraction shows up-regulated nuclear translocation of NF-κB p65 in the 8 Gy IR cultures. The intensities of NF-κB were normalized with nuclear protein, Histone H1 (**, $P < 0.01$, $n = 4$). (B) IF images of confocal microscopy show β1-integrin (green), NF-κB (red) and nuclei (blue). Bar = 50 μm. (C) The binding of NF-κB to the β1-integrin promoter region is up-regulated in MCF10A-Akt cells that survived after exposure to radiation (WT, wild-type, Mt, mutated oligonucleotide; $n = 3$, *, $P < 0.05$). (D-E) The NF-κB inhibitor, JSH-23, was added from day 0 of the second three-dimensional cultures. IR-induced chemoinvasion activity, which was inhibited by NF-κB inhibitor, JSH-23. Bar = 50 μm. IF, immunofluorescence; IR, ionizing radiation; NF-κB, nuclear factor-kappaB; TUNEL, terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling.

been implicated in resistance to radiation treatment and mediating survival in breast cancer cells [9,10,29-31]. Thus, we hypothesized that β1-integrin could be targeted to eradicate the invasive colonies that emerged

post-IR in our current DCIS model. As we expected, the β1-integrin inhibitory antibody could suppress the invasive phenotype and induced apoptosis of the cells, indicating the critical role for β1-integrins in the survival

and progression of these cells that have the ability to invade after significant doses of IR.

We showed previously that the $\alpha 5\beta 1$ -integrin and FN were specifically up-regulated in breast cancer cells in three-dimensional IrECM and after IR [10]. Others have shown that FN and $\alpha 5\beta 1$ -integrin signaling via NF- κ B induces MMP-9 expression in carcinoma cells [16,26,32]. MMP expression has been well correlated with invasive activity in breast cancer [33-36]. In the present study, we found that MMP-9 expression was associated with up-regulated expression of $\alpha 5\beta 1$ -integrin and total FN, as well as EDA+FN, and p-Akt in post-irradiated invasive cells. These results indicate a functional regulation between integrin/ECM signaling and proteinase mediated invasive activity, indicating that targeting FN and $\alpha 5\beta 1$ -integrin signaling via p-Akt could be an effective strategy to decrease the risk of invasive recurrence after the radiation treatment in DCIS patients.

Elevated NF- κ B DNA binding activity has been demonstrated both in breast cancer cell lines and primary breast cancer tissues [37,38]. Constitutive activation of NF- κ B contributes to malignant progression, radio- and chemoresistance and increased metastasis of breast tumors [37,39]. In addition, we have demonstrated a forward loop-like regulation of $\beta 1$ -integrin by NF- κ B post-IR, promoting survival in malignant breast cancer cells [17]. Maity *et al.* reported that the induction of MMP-9 expression and invasion of breast cancer cells are mediated by the activation of NF- κ B via a FN/ $\alpha 5\beta 1$ -integrin-dependent mechanism [26]. These reports and our findings indicate the possibility that IR-induced FN/ $\alpha 5\beta 1$ -integrin expression is associated with NF- κ B activity resulting in up-regulation of $\alpha 5\beta 1$ -integrin, MMP-9 secretion and invasiveness in MCF10A-Akt cells. Investigation of the detailed mechanism of the activation and function of NF- κ B in these cells is currently underway.

Conclusions

P-Akt is up-regulated and is associated with recurrent disease in a limited cohort of clinical DCIS specimens. Akt-overexpressing MCF10A-Akt cells form DCIS like structures in three-dimensional IrECM and *in vivo*, and when treated with IR, luminal cells preferentially undergo apoptosis. When cells surviving IR are selected and repropagated into three-dimensional IrECM, an invasive phenotype emerges, characterized by loss of basal polarity, loss of E-cadherin, ability to invade with increased MMP-9 activity, up-regulated FN, $\alpha 5\beta 1$ -integrin and nuclear translocation of NF- κ B. Together, our results suggest that FN- $\beta 1$ -integrin via NF- κ B feedback signaling is critical mediators of invasive progression post-IR, and could be therapeutic targets to suppress invasive recurrence after radiation treatment of DCIS.

Additional material

Additional file 1: High percentage of $\beta 1$ -integrin expression is associated with recurrent cases compared with non-recurrent cases.

(A) IHC of $\beta 1$ -integrin in human DCIS specimens. Formalin-fixed, paraffin-embedded DCIS sections from 24 patients were stained with $\beta 1$ -integrin monoclonal antibody. All slides were counterstained with hematoxylin and representative image is shown for each intensity and percentage score. Scale bar, 100 μ m. (B) Percentage expression pattern for $\beta 1$ -integrin in human DCIS specimens. Beta1-integrin percentage score: 0 = <10%, 1 = 10%-25%, 2 = 25%-50%, 3 = >50%.

Additional file 2: IR induces apoptosis in an Akt-overexpressing model of human DCIS in three-dimensional IrECM.

Western blot from total cell lysates showed increased expression of cleaved caspase-3 in 8 Gy irradiated MCF10A-Akt cells compared to 0 Gy. Equal amounts of protein were subjected to western blotting. The signals of cleaved caspase-3 were normalized with β -actin. Columns, mean intensity of western blot analysis ($n = 3$, *, $P < 0.05$).

Additional file 3: A phenotype of invasive recurrence with high $\alpha 5\beta 1$ -integrin expression emerged from a sub-population of surviving MCF10A-NeoT cells post-IR in three-dimensional IrECM.

(A) Experimental schema of the recurrence model. (B) Phase-contrast micrographs show that an invasive phenotype emerged by Day 30 of culture. (C) Up-regulated FN and $\alpha 5\beta 1$ -integrin protein level were observed on the 8 Gy IR compared to sham irradiated MCF10A-NeoT three-dimensional IrECM cultures.

Abbreviations

ABC: avidin-biotin-peroxidase; DAB: 3, 3'-diaminobenzidine tetrahydrochloride; DAPI: 4', 6-diamino-2-phenylindole; DCIS: ductal carcinoma *in situ*; (D)MEM: (Dulbecco's) modified Eagle's medium; EDTA: ethylenediaminetetraacetic acid; EGF: epidermal growth factor; FN: fibronectin; H & E, hematoxylin and eosin; HRP: horseradish peroxidase; 4-HT: 4-hydroxytamoxifen; IF: immunofluorescence; IgG: immunoglobulin G; IHC: immunohistochemistry; IR: ionizing radiation; MMP-9: matrix metalloproteinase-9; NF- κ B: nuclear factor-kappa B; OR: odds ratio; p-Akt: phosphorylated-Akt; PVDF: polyvinylidene fluoride; RIPA: radioimmunoprecipitation assay; RT: radiation therapy; TBST: Tris-buffered saline Tween-20; three-dimensional IrECM, three-dimensional laminin-rich extracellular matrix; TUNEL: terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling.

Competing interests

CP is a co-founder of OncoSynergy, Ltd. All other authors declare they have no competing interests.

Authors' contributions

CP and JN developed the study concepts and designed the experiments; JN, KA, HZ and YO performed the experiments; SC performed the imaging analysis; KH conducted the pathological analyses; H. Shirato and H. Sabe supervised the clinical study. H. Sabe supervised data analysis; RK collected and organized clinical information of DCIS patients; AO performed statistical analyses; MI provided expertise in radiation physics. Administrative and material support was provided by MI, H. Sabe and H. Shirato; CP and JN prepared the manuscript with the assistance of YO, KA, HZ, SC, AO, RK, H. Shirato and H. Sabe. All authors read and approved the final manuscript.

Acknowledgements

We thank Atsuko Saito and Mariko Sashika for their technical assistance and Keiko Kanno, Midori Tokuda and Etsuko Hayashi for their administrative work. We also thank Dr. Shigeru Hashimoto and Dr. Ari Hashimoto for support. We are grateful to the Department of Breast and Endocrine Surgery of Hokkaido University Hospital for providing us with the clinical specimens. This work was supported by NIH grant 1R01CA124891 to CP and by the American Cancer Society RSG-07-1110-01-CCE to CP. This work was supported in part by the Matching Program for Innovations in Future Drug Discovery and Medical Care from the Japanese Ministry of Education,

Culture, Sports, Science and Technology (MEXT) and by the Grant-in-Aid from the Japan Society for Young Scientists (B) (23791376 and 25861050) to JN.

Authors' details

¹Department of Radiation Medicine, Hokkaido University Graduate School of Medicine, North-15, West-7, Sapporo, Hokkaido, 060-8638, Japan. ²Life Sciences Division, Ernest Orlando Lawrence Berkeley National Laboratory, 1 Cyclotron Road, Berkeley, 94720, USA. ³Department of Molecular Biology, Hokkaido University Graduate School of Medicine, North-15, West-7, Sapporo, Hokkaido, 060-8638, Japan. ⁴Department of Epidemiology and Biostatistics, University of California San Francisco, 1450 3rd Street San Francisco, CA, 94158, USA. ⁵Department of Surgical Pathology, Hokkaido University Hospital, North-15, West-7, Sapporo, Hokkaido, 060-8638, Japan. ⁶Department of Medical Physics and Engineering, Hokkaido University Graduate School of Medicine, North-15, West-7, Sapporo, Hokkaido, 060-8638, Japan. ⁷Department of Radiation Oncology, University of California San Francisco, California Comprehensive Cancer Center, 1600 Divisadero Street H1031, San Francisco, CA 94143-1708, USA.

Received: 23 January 2013 Revised: 4 May 2013 Accepted: 25 July 2013
Published: 25 July 2013

References

1. Polyak K, Kalluri R: **The role of the microenvironment in mammary gland development and cancer.** *Cold Spring Harb Perspect Biol* 2010, **2**:a003244.
2. Solin LJ: **The impact of adding radiation treatment after breast conservation surgery for ductal carcinoma in situ of the breast.** *J Natl Cancer Inst Monogr* 2010, **2010**:187-192.
3. Fisher B, Costantino J, Redmond C, Fisher E, Margolese R, Dimitrov N, Wolmark N, Wickerham DL, Deutsch M, Ore L, Mamounas E, Poller W, Kavanah M: **Lumpectomy compared with lumpectomy and radiation therapy for the treatment of intraductal breast cancer.** *N Engl J Med* 1993, **328**:1581-1586.
4. Brown LF, Guidi AJ, Schnitt SJ, Van De Water L, Iruela-Arispe ML, Yeo TK, Tognazzi K, Dvorak HF: **Vascular stroma formation in carcinoma in situ, invasive carcinoma, and metastatic carcinoma of the breast.** *Clin Cancer Res* 1999, **5**:1041-1056.
5. Orimo A, Weinberg RA: **Stromal fibroblasts in cancer: a novel tumor-promoting cell type.** *Cell Cycle* 2006, **5**:1597-1601.
6. Pavlakis K, Messini I, Vrekoussis T, Yiannou P, Keramopoulos D, Louvrou N, Liakakos T, Stathopoulos EN: **The assessment of angiogenesis and fibroblastic stromagenesis in hyperplastic and pre-invasive breast lesions.** *BMC Cancer* 2008, **8**:88.
7. Desgrosellier JS, Chersesh DA: **Integrins in cancer: biological implications and therapeutic opportunities.** *Nat Rev Cancer* 2010, **10**:9-22.
8. Cordes N, Blaese MA, Meineke V, Van Beuningen D: **Ionizing radiation induces up-regulation of functional beta1-integrin in human lung tumour cell lines in vitro.** *Int J Radiat Biol* 2002, **78**:347-357.
9. Park CC, Zhang H, Pallavicini M, Gray JW, Baehner F, Park CJ, Bissell MJ: **Beta1 integrin inhibitory antibody induces apoptosis of breast cancer cells, inhibits growth, and distinguishes malignant from normal phenotype in three dimensional cultures and in vivo.** *Cancer Res* 2006, **66**:1526-1535.
10. Nam JM, Onodera Y, Bissell MJ, Park CC: **Breast cancer cells in three-dimensional culture display an enhanced radioresponse after coordinate targeting of integrin alpha5beta1 and fibronectin.** *Cancer Res* 2010, **70**:5238-5248.
11. Ahmed KM, Dong S, Fan M, Li JJ: **Nuclear factor-kappaB p65 inhibits mitogen-activated protein kinase signaling pathway in radioresistant breast cancer cells.** *Mol Cancer Res* 2006, **4**:945-955.
12. Cao N, Li S, Wang Z, Ahmed KM, Degnan ME, Fan M, Dynlacht JR, Li JJ: **NF-kappaB-mediated HER2 overexpression in radiation-adaptive resistance.** *Radiat Res* 2009, **171**:9-21.
13. Brach MA, Hass R, Sherman ML, Gunji H, Weichselbaum R, Kufe D: **Ionizing radiation induces expression and binding activity of the nuclear factor kappa B.** *J Clin Invest* 1991, **88**:691-695.
14. Bhoopathi P, Chetty C, Kunigal S, Vanamala SK, Rao JS, Lakka SS: **Blockade of tumor growth due to matrix metalloproteinase-9 inhibition is mediated by sequential activation of beta1-integrin, ERK, and NF-kappaB.** *J Biol Chem* 2008, **283**:1545-1552.
15. Holmes KM, Annala M, Chua CY, Dunlap SM, Liu Y, Hugen N, Moore LM, Cogdell D, Hu L, Nykter M, Hess K, Fuller GN, Zhang W: **Insulin-like growth factor-binding protein 2-driven glioma progression is prevented by blocking a clinically significant integrin, integrin-linked kinase, and NF-kappaB network.** *Proc Natl Acad Sci USA* 2012, **109**:3475-3480.
16. Sil H, Sen T, Chatterjee A: **Fibronectin-integrin (alpha5beta1) modulates migration and invasion of murine melanoma cell line B16F10 by involving MMP-9.** *Oncol Res* 2011, **19**:335-348.
17. Ahmed KM, Zhang H, Park CC: **NF-kappaB regulates radioresistance mediated by beta1-integrin in three-dimensional culture of breast cancer cells.** *Cancer Res* 2013, **73**:3737-3748.
18. Debnath J, Walker SJ, Brugge JS: **Akt activation disrupts mammary acinar architecture and enhances proliferation in an mTOR-dependent manner.** *J Cell Biol* 2003, **163**:315-326.
19. Behbod F, Kittrell FS, LaMarca H, Edwards D, Kerbawy S, Heestand JC, Young E, Mukhopadhyay P, Yeh HW, Allred DC, Hu M, Polyak K, Rosen JM, Medina D: **An intraductal human-in-mouse transplantation model mimics the subtypes of ductal carcinoma in situ.** *Breast Cancer Res* 2009, **11**:R66.
20. Debnath J, Muthuswamy SK, Brugge JS: **Morphogenesis and oncogenesis of MCF-10A mammary epithelial acini grown in three-dimensional basement membrane cultures.** *Methods* 2003, **30**:256-268.
21. Lee GY, Kenny PA, Lee EH, Bissell MJ: **Three-dimensional culture models of normal and malignant breast epithelial cells.** *Nat Methods* 2007, **4**:359-365.
22. Nam JM, Onodera Y, Mazaki Y, Miyoshi H, Hashimoto S, Sabe H: **CIN85, a Cbl-interacting protein, is a component of AMAP1-mediated breast cancer invasion machinery.** *EMBO J* 2007, **26**:647-656.
23. Park CC, Georgescu W, Polyzos A, Pham C, Ahmed KM, Zhang H, Costes SV: **Rapid and automated multidimensional fluorescence microscopy profiling of 3D human breast cultures.** *Integr Biol* 2013, **5**:681-691.
24. Bose S, Chandran S, Mirocha JM, Bose N: **The Akt pathway in human breast cancer: a tissue-array-based analysis.** *Mod Pathol* 2006, **19**:238-245.
25. Yao ES, Zhang H, Chen YY, Lee B, Chew K, Moore D, Park C: **Increased beta1 integrin is associated with decreased survival in invasive breast cancer.** *Cancer Res* 2007, **67**:659-664.
26. Maity G, Choudhury PR, Sen T, Ganguly KK, Sil H, Chatterjee A: **Culture of human breast cancer cell line (MDA-MB-231) on fibronectin-coated surface induces pro-matrix metalloproteinase-9 expression and activity.** *Tumour Biol* 2011, **32**:129-138.
27. Meineke V, Gilbertz KP, Schilperoot K, Cordes N, Sendler A, Moede T, van Beuningen D: **Ionizing radiation modulates cell surface integrin expression and adhesion of COLO-320 cells to collagen and fibronectin in vitro.** *Strahlenther Onkol* 2002, **178**:709-714.
28. Cordes N, Seidler J, Durzok R, Geinitz H, Brakebusch C: **beta1-integrin-mediated signaling essentially contributes to cell survival after radiation-induced genotoxic injury.** *Oncogene* 2006, **25**:1378-1390.
29. Cordes N, Blaese MA, Plasswilm L, Rodemann HP, Van Beuningen D: **Fibronectin and laminin increase resistance to ionizing radiation and the cytotoxic drug Ukrain in human tumour and normal cells in vitro.** *Int J Radiat Biol* 2003, **79**:709-720.
30. Nam JM, Chung Y, Hsu HC, Park CC: **Beta1 integrin targeting to enhance radiation therapy.** *Int J Radiat Biol* 2009, **85**:923-928.
31. Park CC, Zhang HJ, Yao ES, Park CJ, Bissell MJ: **Beta1 integrin inhibition dramatically enhances radiotherapy efficacy in human breast cancer xenografts.** *Cancer Res* 2008, **68**:4398-4405.
32. Sen T, Dutta A, Maity G, Chatterjee A: **Fibronectin induces matrix metalloproteinase-9 (MMP-9) in human laryngeal carcinoma cells by involving multiple signaling pathways.** *Biochimie* 2010, **92**:1422-1434.
33. Deryugina EI, Quigley JP: **Matrix metalloproteinases and tumor metastasis.** *Cancer Metastasis Rev* 2006, **25**:9-34.
34. Stetler-Stevenson WG: **Matrix metalloproteinases in angiogenesis: a moving target for therapeutic intervention.** *J Clin Invest* 1999, **103**:1237-1241.
35. Sun Y, Lu N, Ling Y, Gao Y, Chen Y, Wang L, Hu R, Qi Q, Liu W, Yang Y, You Q, Guo Q: **Oroxilin A suppresses invasion through down-regulating the expression of matrix metalloproteinase-2/9 in MDA-MB-435 human breast cancer cells.** *Eur J Pharmacol* 2009, **603**:22-28.
36. Kim ES, Kim JS, Kim SG, Hwang S, Lee CH, Moon A: **Sphingosine 1-phosphate regulates matrix metalloproteinase-9 expression and breast cell invasion through S1P3-Galpaq coupling.** *J Cell Sci* 2011, **124**:2220-2230.

37. Biswas DK, Iglehart JD: Linkage between EGFR family receptors and nuclear factor kappaB (NF-kappaB) signaling in breast cancer. *J Cell Physiol* 2006, **209**:645-652.
38. Sovak MA, Bellas RE, Kim DW, Zanieski GJ, Rogers AE, Traish AM, Sonenshein GE: Aberrant nuclear factor-kappaB/Rel expression and the pathogenesis of breast cancer. *J Clin Invest* 1997, **100**:2952-2960.
39. Wu JT, Kral JG: The NF-kappaB/IkappaB signaling system: a molecular target in breast cancer therapy. *J Surg Res* 2005, **123**:158-169.

doi:10.1186/bcr3454

Cite this article as: Nam *et al.*: β 1-integrin via NF- κ B signaling is essential for acquisition of invasiveness in a model of radiation treated *in situ* breast cancer. *Breast Cancer Research* 2013 **15**:R60.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

